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SUPPLEMENTARY DATA

Supplementary Table 1. Antibodies used for immunofluorescence analysis

| Usage | Antibodies | Manufacturer | Catalogue number | Dilution |
|----------------------------------|------------------|------------------------------|------------------|----------|
| Western Blotting | Lamin A/C (N-18) | Santa Cruz Biotechnology, CA | Sc-6215 | 1:500 |
| | Beta-Actin | Santa Cruz Biotechnology, CA | Sc-47778 | 1:500 |
| IPS characterization | OCT4 | Stemgent, Cambridge, MA | 09-0023 | 1:100 |
| | SSEA-4 | | 09-0006 | |
| | Tra1-60 | | 09-0010 | |
| | Nanog | | 09-0020 | |
| CM staining | Alpha-actinin | Sigma-Aldrich, St. Louis, MO | A7811 | 1:200 |
| EC staining | Lectin | Sigma-Aldrich, St. Louis, MO | L9006 | 1:100 |
| | vWF | Millipore | AB7568 | 1:100 |
| Fibroblast staining | Fibronectin | Santa Cruz Biotechnology, CA | Sc-69777 | 1:100 |
| | Vimentin | | Sc-6260 | 1:100 |
| Nuclear blebbing analysis | Lamin A/C (N-18) | Santa Cruz Biotechnology, CA | Sc-6215 | 1:100 |

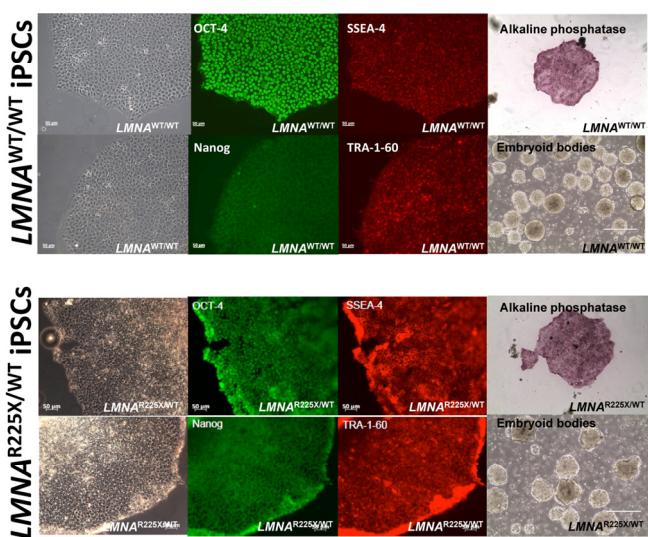
Supplementary Table 2. PCR primers and conditions for reprogramming transgene silencing analysis

| Gene | Accession no. | Forward/reverse (5'→3') | Annealing temperature (°C) | Product (bp) | Cycles |
|------------|----------------|---|----------------------------|--------------|--------|
| Endo-OCT4 | NM_001159542.1 | 5'- GACAACAATGAAAATCTTCAGGAGA -3' 5' - TTCTGGCGCCGGTTACAGAACCA -3' | 57 | 223 | 30 |
| Endo-NANOG | NM_024865.2 | 5'-AAGACAAGGTCCCCTGCAAG 5'- CCTAGTGGTCTGCTGTATTAC | 57 | 583 | 30 |
| *Exo-OCT4 | NM_001159542.1 | 5'-TCAAGCCTCAGACAGTGGTC3' 5'-GGCCCGATTCTGGCCCTCA3' | 57 | 236 | 30 |
| *Exo-NANOG | NM_024865 | 5'-TCAAGCCTCAGACAGTGGTC-3' 5'-CTTCAAAGCAAGGCAAGCTT-3' | 57 | 296 | 30 |
| GAPDH | NM_011406 | 5'- AGCCACATCGCTCAGACACC -3' 5'- GTACTCAGCGGCCAGCATCG -3' | 60 | 157 | 30 |

Abbreviation: OCT4 : octamer-binding transcription factor 4 ; GAPDH: Glyceraldehyde 3-phosphate dehydrogenase.

*Forward primer was probed on EF1-alpha coding region, which is upstream of the OCT-4/NANOG cDNA sequence in the lentiviral reprogramming vector.

A



B

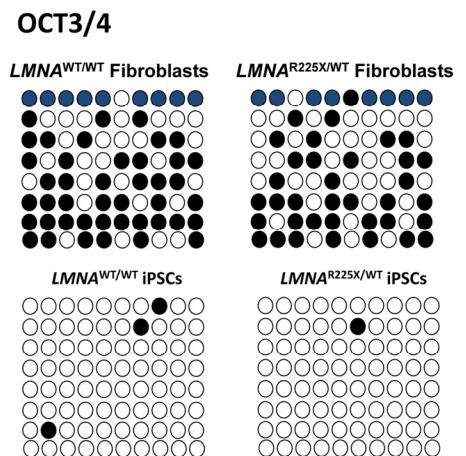
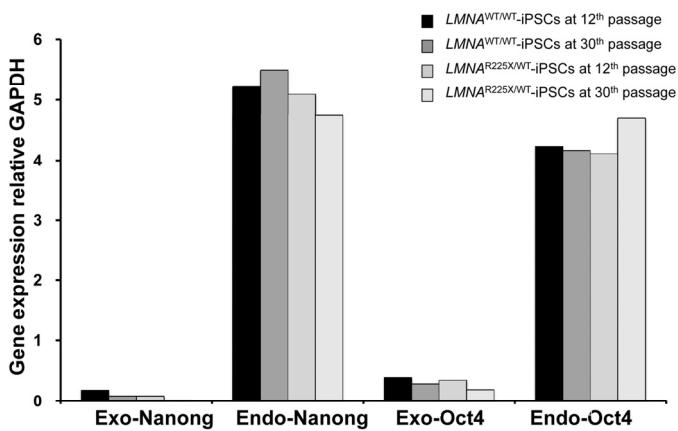


Figure 1. Generation of patient-specific iPSC lines. (A) Immunofluorescence analysis of pluripotent markers OCT4 (Green), SSEA4 (Red), NANOG (Green), and TRA-1-60; the expression of alkaline phosphatase; and embryoid body formation in representative iPSC clones derived from the proband (II:7) and the healthy control, **(B)** Oct-4 promoter methylation analysis with bisulfate pyro-sequencing in two parent fibroblast lines ($LMNA^{R225X/WT}$ and $LMNA^{WT/WT}$), and their iPSC lines.

C



D

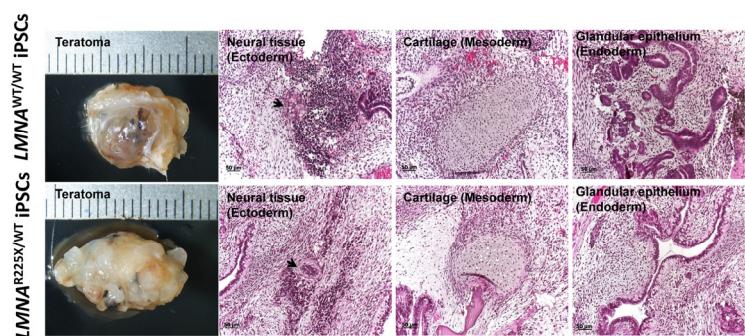
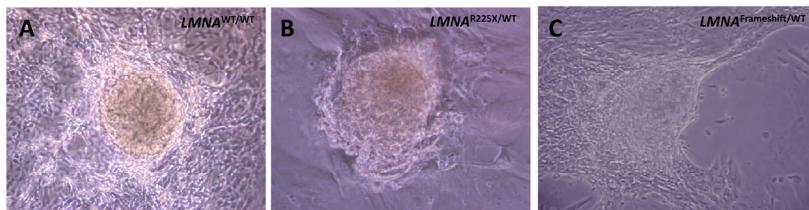


Figure 1. Generation of patient-specific iPSC lines. (C) RT-PCR analyses of the endogenous and exogenous level of OCT4 and Nanog of LMNA^{R225X/WT} and LMNA^{WT/WT} iPSC lines at 12th and 30th passages, (D) Teratoma formation and the histological section of teratoma formed 4-6 weeks after subcutaneous injection of LMNA^{R225X/WT} and LMNA^{WT/WT} iPSC lines into NOD/SCID mice.

Beating Embryoid Body



Beating Cluster

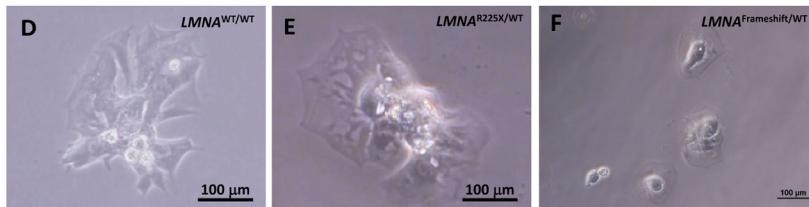


Figure 2. (A, B, & C) Beating embryoid bodies derived from LMNA^{R225X/WT}, LMNA^{Frameshift/WT}, and LMNA^{WT/WT} iPSCs; (D, E, & F) Spontaneously beating cell clusters after dissociation. Videos of these beating embryoid bodies and clusters were available in supplemental materials.

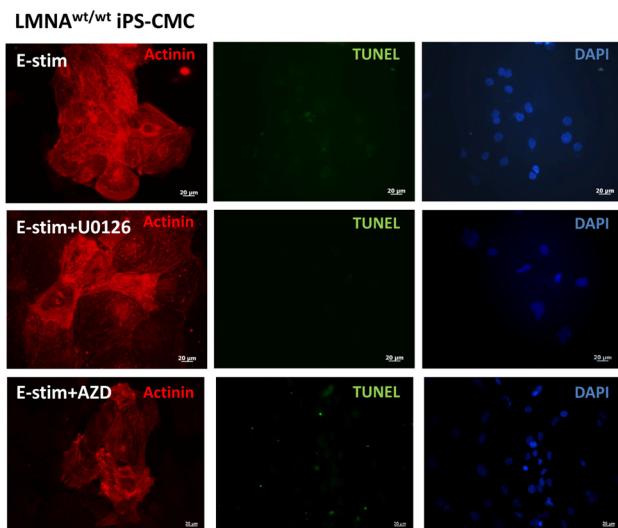
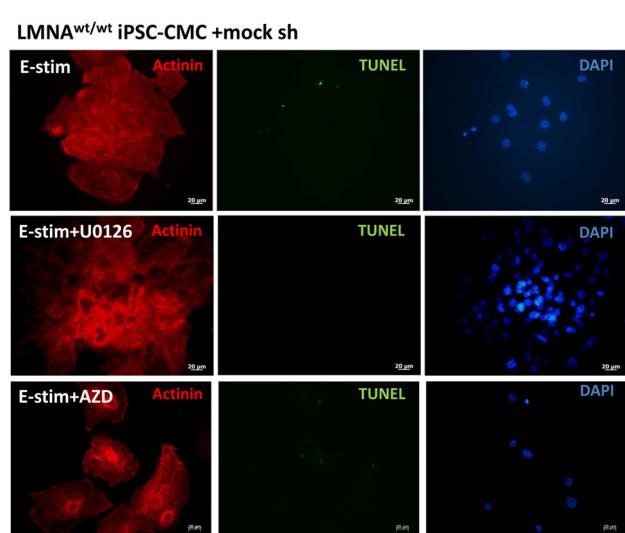
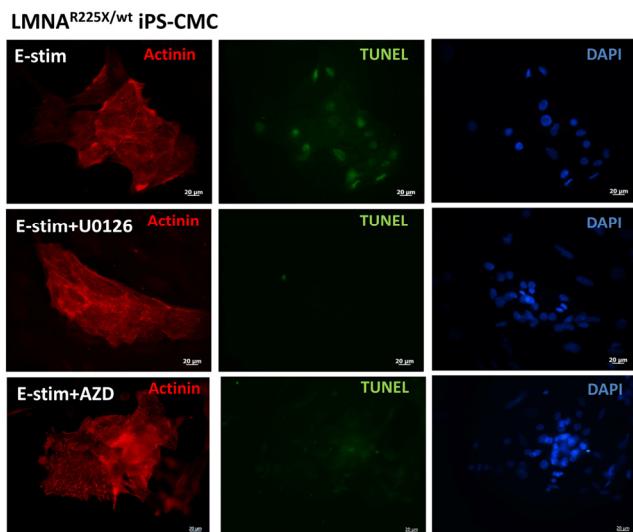
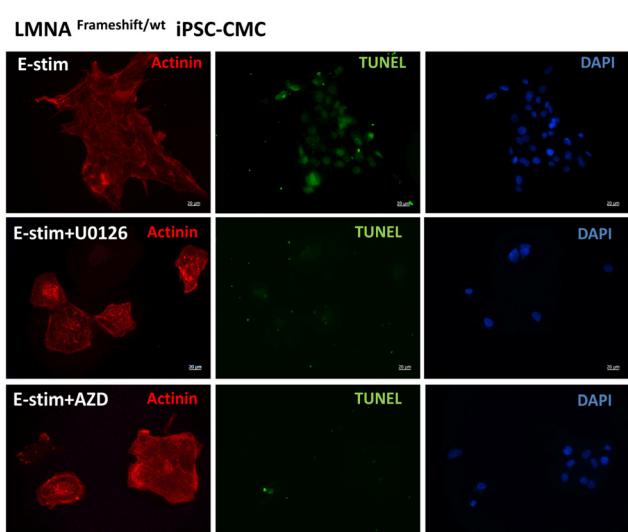
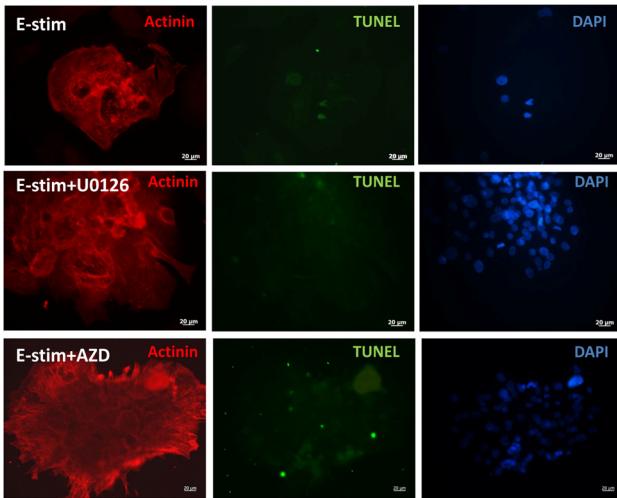
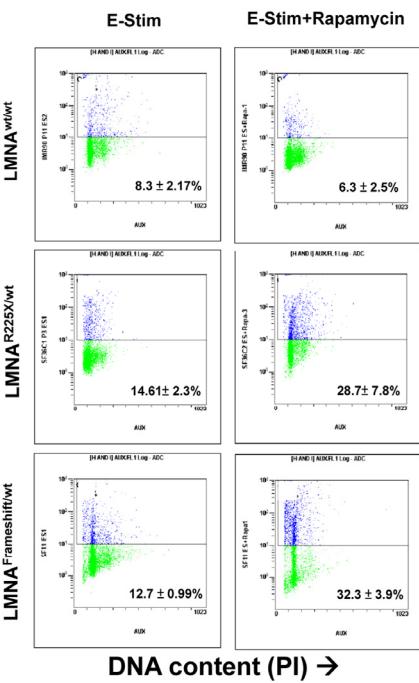
A**B****C****D**

Figure 3. Electrical stimulation inducing apoptosis in cardiomyocytes derived from LMNA^{R225X/WT} & LMNA^{Frameshift/WT} iPSCs. Representative TUNEL assay and co-immunofluorescence staining of alpha-actinin in cardiomyocytes derived from (A) LMNA^{WT/WT} iPSCs, (B) LMNA^{R225X/WT} iPSCs, (C) LMNA^{Frameshift/WT} iPSCs, (D) LMNA^{WT/WT} iPSCs treated with mock shRNA.

E

LMNA^{wt/wt} + shLMNA iPSC-CMC

F



G

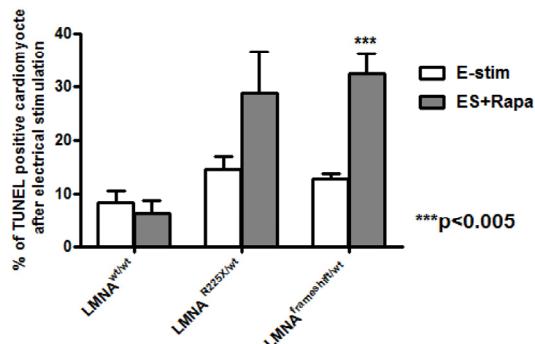


Figure 3. Electrical stimulation inducing apoptosis in cardiomyocytes derived from LMNA & LMNA iPSCs. (E) LMNA iPSCs treated with shLMNA, (F and G) Quantification of apoptotic cardiac differentiated iPSCs in presence of rapamycin by APO-BrdU TUNEL assay at baseline and after electrical stimulation. The percentage of cardiomyocytes with apoptosis was determined by FACS analysis by FL-1 positive gating. Unpaired t-test was performed between treatment and baseline n=3.