**Research Paper** 

# Multiple dietary supplements do not affect metabolic and cardiovascular health

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Abstract: Dietary supplements are widely used for health purposes. However, little is known about the metabolic and cardiovascular effects of combinations of popular over-the-counter supplements, each of which has been shown to have anti-oxidant, anti-inflammatory and pro-longevity properties in cell culture or animal studies. This study was a 6-month randomized, single-blind controlled trial, in which 56 non-obese (BMI 21.0-29.9 kg/m<sup>2</sup>) men and women, aged 38 to 55 yr, were assigned to a dietary supplement (SUP) group or control (CON) group, with a 6-month follow-up. The SUP group took 10 dietary supplements each day (100 mg of resveratrol, a complex of 800 mg each of green, black, and white tea extract, 250 mg of pomegranate extract, 650 mg of quercetin, 500 mg of acetyl-l-carnitine, 600 mg of lipoic acid, 900 mg of curcumin, 1 g of sesamin, 1.7 g of cinnamon bark extract, and 1.0 g fish oil). Both the SUP and CON groups took a daily multivitamin/mineral supplement. The main outcome measures were arterial stiffness, endothelial function, biomarkers of inflammation and oxidative stress, and cardiometabolic risk factors. Twenty-four weeks of daily supplementation with 10 dietary supplements did not affect arterial stiffness or endothelial function in nonobese individuals. These compounds also did not alter body fat measured by DEXA, blood pressure, plasma lipids, glucose, insulin, IGF-1, and markers of inflammation and oxidative stress. In summary, supplementation with a combination of popular dietary supplements has no cardiovascular or metabolic effects in non-obese relatively healthy individuals.

### **INTRODUCTION**

Non-vitamin, non-mineral dietary supplements are widely used for health purposes and sometimes as a substitute to a healthy diet or conventional medical treatments. Nearly 1 in 7 adults takes supplements regularly, and approximately 40% have taken one or more dietary supplements during their life [1]. However, despite the widespread and growing use of these over-the-counter products, insight into the potential beneficial or harmful biological effects of these compounds in humans is frequently lacking. Some individuals, especially heavy supplement users, typically consume combinations of dietary supplements because they believe that multiple compounds can act through complimentary, additive or synergistic mechanisms to convey a greater biologic effect than can be achieved by any individual supplement [2]. The present study was a randomized clinical trial to evaluate the effectiveness of supplementation with a combination of some of the most self-prescribed dietary supplements (i.e. resveratrol, curcumin, green/black/white tea extract, quercetin, acetyl-l-carnitine, lipoic acid, pomegranate, cinnamon bark, sesamin, and fish oil), in

lean and overweight middle-aged men and women eating a Western diet. It has been reported that these compounds exert powerful protective effects against inflammation, oxidative stress/free radical damage, insulin resistance, and protein glycation in cell culture and laboratory animal studies [3-24]. We evaluated the combined effects of these supplements on arterial stiffness, endothelial function, markers of inflammation, oxidative stress, glucose and lipid metabolism, and blood pressure.

### **RESULTS**

#### **Study Participants**

Screening, enrollment, and follow-up information is presented in Fig. 1. The study participants were generally healthy, as reflected by the following: BMI,  $25.0\pm2.3 \text{ kg/m}^2$ ; total cholesterol,  $187\pm32 \text{ mg/dL}$ ; triglycerides,  $79\pm53 \text{ mg/dL}$ ; fasting glucose,  $82\pm9$ mg/dL; and systolic and diastolic blood pressures of  $109\pm12$  and  $68\pm8$  mmHg, respectively. The proportions of men and women did not differ between groups (p=0.65). The average age of participants was slightly lower in the SUP group (means $\pm$ SD:  $44\pm6$  vs.  $47\pm5$  yr, p=0.05). BMI did not differ (p=0.65) between the SUP ( $25.2\pm2.0 \text{ kg/m}^2$ ) and control ( $24.9\pm2.5 \text{ kg/m}^2$ ) groups. Based on monthly pill compliance queries, 96% of the participants in the SUP group took all of the prescribed doses of the supplements. All participants in both groups complied with the vitamin/mineral supplementation regimen.

There were no differences between groups in PWV, AI, blood pressure, and endothelium-dependent and endothelium-independent brachial artery vasodilation (Table 1). No changes in body weight or % total body fat occurred in either study group (Table 2). Plasma lipid concentrations and indices of glucoregulation did not change in either group (Table 2). No differences between groups were observed for markers of inflammation, oxidative stress and glycation, and blood counts (Table 2). Serum insulin and IGF-1 concentrations did not change in either group (Table 2). Since completing the randomized trial, all but one of the participants in the SUP group continued for another 6 months on the supplementation regimen. Even with this longer, 12 month period of supplementation, no changes in any outcome were observed (data not shown).

No serious adverse events occurred. Serum markers of liver and kidney function were unaffected by supplementation. Other adverse events were limited to mild gastrointestinal discomfort associated with taking the large number of oral supplements in 19% of the participants.



Figure 1. Consort diagram reflecting flow of study participants through the study

	Supplements(n= 28)	Control (n=26)	Adjusted Difference Between Groups	Between Group P value
Pulse wave velocity, m/s				
Baseline	$5.4 \pm 0.3$	$5.2 \pm 0.2$		
6 months	$4.8 \pm 0.3$	$5.3 \pm 0.2$		
Change	$-0.5 \pm 0.4$	$0.1 \pm 0.3$	$-0.3 \pm 0.4$	0.36
Augmentation index, %				
Baseline	$11.7 \pm 2.6$	$12.3 \pm 2.8$		
6 months	$12.5 \pm 2.8$	$13.0 \pm 2.3$		
Change	$0.8 \pm 1.1$	$0.7 \pm 1.1$	$0.4 \pm 1.5$	0.77
Flow-mediated dilation, %				
Baseline	$5.0 \pm 0.5$	$4.1 \pm 0.4$		
6 months	$4.5 \pm 0.4$	$4.4 \pm 0.3$		
Change	$-0.5 \pm 0.5$	$0.3 \pm 0.3$	$-0.3 \pm 0.5$	0.54
GTN-mediated dilation, %				
Baseline	$16.1 \pm 1.3$	$14.6 \pm 1.2$		
6 months	$14.2 \pm 1.6$	$13.3 \pm 0.9$		
Change	$-1.9 \pm 1.1$	$-1.3 \pm 1.4$	$-0.2 \pm 1.7$	0.93
Systolic BP, mmHg				
Baseline	$108 \pm 2$	$109 \pm 2$		
6 months	$108 \pm 2$	$107 \pm 3$		
Change	$0 \pm 1$	$-2 \pm 1$	$1\pm 2$	0.48
Diastolic BP, mmHg				
Baseline	$69 \pm 1$	$66 \pm 2$		
6 months	$69 \pm 1$	$64 \pm 2$		
Change	$0 \pm 1$	$-2 \pm 1^*$	$2 \pm 1$	0.08

Table 1. Effect of 6 months of nutritional supplements or control on indices of arterial stiffness, vasomotor

Values are arithmetic means  $\pm$  SE except for mean differences between groups, which have been adjusted for baseline values. Between-group P values reflect the between-group comparison change-scores from ANCOVAs that included baseline values as the covariate. \*Significant (p≤0.05) within-group change. GTN, nitroglycerine; BP, blood pressure. Several subjects in the supplement group (n=15) and control group (n=14) did not undergo testing for GTN-mediated dilation because systolic blood pressure was below 100 mmHg, which is a contraindication to GTN administration.

### DISCUSSION

Our findings indicate that daily use of multiple dietary supplements has no effect on arterial stiffness (i.e. pulse wave velocity and augmentation index), endothelial brachial arterv flow-dependent function (i.e. vasodilatation) or blood pressure in nonobese men and women. Furthermore, supplementation with these compounds did not affect key metabolic variables implicated in the biology of aging, and in the pathogenesis of cardiovascular disease, including plasma

markers of inflammation, oxidative stress and glycation, plasma lipids, growth factors, or body composition.

Reports from studies conducted mainly on cells in culture or in experimental animals suggest that these compounds promote metabolic health and may have anti-aging effects [4-24]. In particular, resveratrol, quercetin, curcumin, acetyl-L-carnitine, lipoic acid, fish oil, sesamin, pomegranate, cinnamon bark, and green tea extracts have all been reported to have powerful anti-oxidant and/or anti-inflammatory properties [4-24].

	Supplements (n=28)	Control (n=26)	Adjusted Difference Between Groups	Between Group P value
Body Mass and Composition				
Body mass, kg				
Baseline	$73.8 \pm 2.3$	$72.8 \pm 1.8$		
6 months	$74.2 \pm 2.5$	$73.1 \pm 1.7$		
Change	$0.3 \pm 0.4$	$0.2 \pm 0.4$	$0.1 \pm 0.6$	0.91
Body fat, %				
Baseline	$30.1 \pm 1.1$	$30.2 \pm 1.7$		
6 months	$29.9 \pm 1.0$	$30.2 \pm 1.8$		
Change	$-0.2 \pm 0.3$	$0.2 \pm 0.3$	$-0.2 \pm 0.4$	0.70
ipids				
Triglycerides, mg/dL				
Baseline	$79 \pm 12$	$78 \pm 8$		
6 months	$75 \pm 6$	$78\pm8$ $87\pm8$		
Change	$-4 \pm 11$	$8 \pm 4$	$-12 \pm 8$	0.16
Chunge	$+ \pm 11$	0 = 4	$12 \pm 0$	0.10
Total cholesterol, mg/dL				
Baseline	$186 \pm 6$	$189 \pm 6$		
6 months	$186 \pm 6$	$192 \pm 5$		
Change	$-1 \pm 4$	$4 \pm 4$	$-5 \pm 5$	0.32
LDL-cholesterol, mg/dL				
Baseline	$110 \pm 6$	$114 \pm 6$		
6 months	$110 \pm 0$ $111 \pm 5$	$111 \pm 0$ $115 \pm 5$		
Change	$1\pm 3$	$115 \pm 5$ $1 \pm 4$	$-1 \pm 5$	0.80
-				
HDL-cholesterol, mg/dL	(1 + 2)	(1 + 2)		
Baseline	$61 \pm 3$ $59 \pm 3$	$61 \pm 3$ $60 \pm 3$		
6 months	$59 \pm 3$ -2 ± 2	$60 \pm 3$	1 + 2	0.59
Change	- <i>∠</i> ± <i>∠</i>	$-1 \pm 2$	$-1 \pm 2$	0.59
<b>Elucoregulatory Function</b>				
Fasting glucose, mg/dL		o.		
Baseline	$80 \pm 2$	$84 \pm 2$		
6 months	$83 \pm 2$	$85 \pm 2$		
Change	$3\pm 2$	$1\pm 2$	$-1 \pm 2$	0.70
Fasting insulin, µU/mL				
Baseline	$3.3 \pm 0.3$	$2.8 \pm 0.3$		
6 months	$3.6 \pm 0.4$	$2.0 \pm 0.3$ $2.9 \pm 0.3$		
Change	$0.3 \pm 0.3$	$0.1 \pm 0.2$	$0.3 \pm 0.4$	0.41
HOMA-IR				
Baseline	$0.64 \pm 0.06$	$0.60 \pm 0.06$		
6 months Change	$\begin{array}{c} 0.74 \pm 0.09 \\ 0.10 \pm 0.06 \end{array}$	$0.63 \pm 0.06 \\ 0.03 \pm 0.04$	$0.08 \pm 0.08$	0.32

Table 2. Risk factors for cardiovascular disease and diabetes, and circulating markers of oxidative stress,

, , , , , , , , , , , , , , , , , , , ,	in response to 6 months of nutritional supplementation or control. Adjusted				
	Supplements (n=28)	Control (n=26)	Difference Between Groups	Betweer Group P value	
			Gloups		
flammatory Cytokines and Ox	idative Stress Markers				
CRP, mg/L					
Baseline	$1.69 \pm 0.48$	$1.21 \pm 0.52$			
6 months	$1.51 \pm 0.20$	$1.44 \pm 0.56$			
Change	$-0.17 \pm 0.49$	$0.23 \pm 0.10*$	$-0.20 \pm 0.43$	0.46	
TNFα, pg/mL					
Baseline	$1.92 \pm 0.09$	$2.10 \pm 0.18$			
6 months	$1.94 \pm 0.09$	$2.13 \pm 0.19$			
Change	$0.02 \pm 0.05$	$0.03 \pm 0.14$	$-0.05 \pm 0.14$	0.70	
IL-6, pg/mL					
Baseline	$1.26 \pm 0.26$	$1.39 \pm 0.31$			
6 months	$1.26 \pm 0.26$ $1.35 \pm 0.23$	$1.59 \pm 0.31$ $1.68 \pm 0.32$			
6 months Change	$1.35 \pm 0.23$ $0.08 \pm 0.08$	$1.68 \pm 0.32$ $0.29 \pm 0.20$	$-0.22 \pm 0.20$	0.27	
Change	$0.08 \pm 0.08$	$0.29 \pm 0.20$	$-0.22 \pm 0.20$	0.27	
Protein carbonyl, nmol/mg					
Baseline					
6 months	$0.83 \pm 0.03$	$0.79 \pm 0.03$			
Change	$0.76 \pm 0.03$	$0.82 \pm 0.03$			
	$-0.07 \pm 0.04$	$0.03 \pm 0.03$	$-0.06 \pm 0.04$	0.14	
AGEs, ng/mL					
Baseline	$307 \pm 14$	$272 \pm 13$			
6 months	$287 \pm 12$	$270 \pm 12$			
Change	$-20 \pm 13$	$-2 \pm 12$	$1 \pm 14$	0.96	
hite Cell Counts and Growth 1	Factors				
WBC, k/cumm					
Baseline	$5.1 \pm 0.2$	$4.8 \pm 0.2$			
6 months	$5.0 \pm 0.2$	$5.1 \pm 0.3$			
Change	$-0.1 \pm 0.2$	$0.3 \pm 0.2$	$-0.3 \pm 0.3$	0.20	
Lymphocytes, k/cumm					
Baseline	$1.50 \pm 0.05$	$1.42 \pm 0.09$			
6 months	$1.42 \pm 0.06$	$1.42 \pm 0.09$ $1.44 \pm 0.10$			
Change	$-0.08 \pm 0.04*$	$0.02 \pm 0.05$	$-0.10 \pm 0.06$	0.11	
IGF-1, ng/mL					
Baseline	$145 \pm 6$	$148 \pm 7$			
6 months	$143 \pm 0$ $151 \pm 7$	$140 \pm 7$ $144 \pm 6$			
Change	$6\pm 5$	$-4 \pm 4$	$9\pm 6$	0.15	

Values are arithmetic means  $\pm$  SE except for mean differences between groups which have been adjusted for baseline values. Between-group P values reflect the between-group comparison change-scores from ANCOVAs that included baseline values as the covariate. \*Significant (p $\leq$ 0.05) within-group change. Triglyceride data were also adjusted for a significant effect of age on the baseline to follow up changes. Within-group P values are from paired t-tests. LDL, low density lipoprotein; HDL, high density lipoprotein; HOMA-IR, homeostasis model assessment of insulin resistance; CRP, C-reactive protein, TNF $\alpha$ , tumor necrosis factor  $\alpha$ ; IL-6, interleukin-6; AGEs, advanced glycation end products; WBC, white blood cells; IGF-1, insulin-like growth factor-1. To convert units to SI units, multiply the conventional units by the following conversion factors: triglycerides x 0.0113 = mmol/L; total, LDL-, and HDL-cholesterol x 0.0259 = mmol/L; glucose x 0.0555 = mmol/L; insulin x 6.945 = pmol/L.

However, in our study we did not see any reduction in markers of inflammation or oxidative stress. Oxidative stress and inflammation are major players in the pathogenesis of arterial aging and endothelial dysfunction, which is a precursor of atherosclerosis [25-29]. Nevertheless, pulse wave velocity, augmentation index and endothelium-dependent vasorelaxation did not improve in the supplement group. Furthermore, supplementation with these compounds did not alter other well-accepted cardiometabolic risk factors, including blood pressure, insulin resistance, and serum cholesterol, triglyceride and advanced glycation endproducts concentrations. Endothelial function can be improved with just a single infusion of the anti-oxidant, vitamin C [30]. Because all the supplements that we used have been reported to have powerful antiinflammatory and anti-oxidant effects, it is interesting that no biological change in endothelial function was observed. In contrast, it has been shown that 6-mo or 1yr supplementation with DHEA-s results in significant improvements of glucose tolerance, arterial stiffness and inflammation [31].

One possible explanation for the lack of beneficial metabolic effects of these over-the counter dietary supplements could be the low phytochemical bioavailability or inadequate supplement potency of the phytochemicals contained in some of these compounds which are available without prescription [32]. Nonetheless, the fact remains that millions of individuals in USA and Europe are consuming these supplements, sometimes instead of a healthy diet and conventional medical treatment, which might contain hazard trace amounts of pesticides and heavy metals, including lead, arsenic, mercury, and cadmium [1,33].

A placebo was not provided to control group participants in the present study, which might be viewed as a limitation. However, while a "placebo effect" might have been an explanation for significant changes in the supplement group, we did not observe changes. Therefore, in this context, the lack of a placebo is not a limitation. Moreover, although the sample size was relatively small, there was no suggestion of clinically significant differences from the between-group estimates, despite multiple comparisons. Finally, it is conceivable that some supplements might have had beneficial effects that where counteracted by negative effects of others. However, this seems unlikely and people rarely take isolated supplements, and many of these compounds are contained in foods and beverages that are consumed regularly by millions of people. Moreover, heavy supplement users believe that these compounds work better in combination, because as with food and beverages, they provide a mix of phytochemicals that interact and potentiate their effect (e.g. resveratrol in wine, cathechins in green and black tea, pomegranate in fruits, curcumin in spices, etc.).

Findings from the present study suggest that a combination of several popular nutritional supplements, which are commonly taken with the intention of promoting healthy longevity and preventing chronic disease, have no effects on arterial stiffness, endothelial function, inflammation, oxidative stress, and other chronic disease risk factors in non-obese men and women. Additional randomized controlled studies are still needed to assess the potential benefits of these and other dietary supplements in obese metabolically abnormal individuals.

### **METHODS**

<u>Participants.</u> Non-obese (BMI 21.0-29.9 kg/m<sup>2</sup>) men and women, aged 38-65y were recruited from the Saint Louis area. Participants were free of chronic disease based on a medical history, physical examination, blood and urine chemistries, and electrocardiogram. Exclusion criteria included chronic use of medications or dietary supplements, tobacco use, alcohol abuse, and habitual vigorous exercise. Participants consented to participate in the study, which was approved by the Washington University Institutional Review Board.

Study Design. The study was a 6-month single-blind controlled trial in which participants were randomized (1:1 ratio with stratification for sex) to a nutritional supplement (SUP) group or control (CON) group. At the end of the RCT, the participants who had been randomized to SUP continued taking the supplements for another 6-mo, while those who had been in the CON group crossed over to the supplementation regimen for Technicians who performed 6-mo. outcomes assessments were blinded to study group assignments. Participants fasted overnight (12-hr) and refrained from exercise for 24-hr before testing. For follow-up tests, participants in the SUP group were instructed to take their nutritional supplements upon waking in the morning prior to testing. The primary outcome, carotidfemoral pulse wave velocity (PWV), was measured by using Doppler flow measures. Another index of arterial stiffness, carotid artery augmentation index (AI) was measured using applanation tonometry. Endotheliumdependent and endothelium-independent brachial artery vasodilation were evaluated by using ultrasound imaging to measure flow-mediated dilation and glyceryl trinitrate- (GTN)-mediated dilation, respectively.

<u>Intervention</u>. Participants in the SUP group took the following dietary supplements: resveratrol (100 mg/day), quercetin (650 mg/day), acetyl-l-carnitine HCL (500/mg/day), alpha-lipoic acid (600 mg/day),

curcumin complex (900 mg/day; standardized to 95% total curcuminoids, plus piperine 5 mg), pomegranate extract (250 mg/day; standardized to 70% ellagic acid), fish oil (1 g/day, containing 300 mg of eicosapentaenoic acid and 200 mg docosahexaenoic acid), cinnamon bark (1.7 g/day), green/white/black tea complex (800 mg/day each of green, black, and white tea extract) and sesamin (1 g/day; standardized to 500 mg sesamin lignans). Sesamin was formulated by Scivation, Inc. (Burlington, NC, USA) and all others by Swanson Health Products (Fargo, ND, USA). Both groups received a daily multivitamin/mineral supplement (Daily Multi-Vitamin & Mineral, Swanson, Fargo, ND, USA) and were advised to maintain their usual diet and physical activity. Participants met with a member of the research team each month to receive supplements and to answer questions about compliance with the supplementation regimen, changes in diet, physical activity, medical conditions, and medication use, and adverse events.

<u>Body weight and composition.</u> Weight and height were measured with a calibrated balance beam scale and wall-mounted stadiometer, respectively, with the participant wearing only a hospital gown and underwear. BMI was calculated (kg/m<sup>2</sup>). Body composition was evaluated using dual energy X-ray absorptiometry (Delphi W, software version 11.2, Hologic Corp., Waltham, MA). Bilateral brachial artery blood pressure was measured with a calibrated monitor (Dinamap 1846 SX, Critkon, Inc, Tampa, FL) according to JNC 7 guidelines.

Circulating biomarkers of disease risk and aging. Blood samples were collected from a forearm vein into separate tubes containing sodium heparin, EDTA, and no additives. Plasma and serum were isolated by centrifugation (3500 g, 15 min, 4° C) and stored at -80° C for later batch analyses. Samples were analyzed for plasma concentrations of glucose (glucose oxidase method, 2300 Stat Plus, YSI Inc., Yellow Springs, OH) and insulin (chemiluminescence assay, Immulite 1000, Siemens USA. Malvern, PA). The homeostasis model assessment of insulin resistance (HOMA-IR) was calculated from fasting glucose and insulin. ELISA assay kits were used to measure serum concentrations of inflammatory cytokines (tumor necrosis factor a (TNFα), interleukin-6 (IL-6), C-reactive protein (CRP), Quantikine, R&D Systems, Minneapolis, MN). oxidative stress markers (protein carbonyls (Cell Biolabs, San Diego, CA) and advanced glycation endproduct (AGE) N-1-carboxymethyl lysine (MBL International, Woburn, MA)), and insulin-like growth factor-1 (IGF-1, Diagnostic Systems Laboratories, Webster, TX). Plasma lipids and blood counts were

measured by a CLIA-certified clinical laboratory at the medical center.

Indices of arterial stiffness. Carotid artery augmentation index (AI) was measured by using applanation tonometry according to published guidelines [34]. Twenty digital pulse waves were recorded with a tonometer (Millar Instruments, Inc., Model #TCB-500, Houston, TX) and analyzed with Windaq software (version 2.31, DATAQ Instruments, Inc., Akron, OH). The the maximum and minimum voltage on each wave form was identified and used to calculate pulse pressure (PP). The second derivative of the pulse wave was generated and used to identify the "shoulder" on the upstroke of the raw wave form. Augmentation pressure (AP) was calculated as the difference between the peak voltage and the voltage at the shoulder. Augmentation index was calculated as  $AI = 100 \times AP/PP$  for each of the 20 waveforms; the resulting values were averaged. Because AI is dependent on heart rate, AI values were adjusted to a standardized heart rate of 75 beats/min based on the inverse relationship of 4.8 AI units per 10 beats/min HR [35]. The technician visually inspected all waveforms to ensure that the landmarks had been properly identified and to omit waveforms that were of suboptimal quality due to artifacts or irregular heartbeats. When analyses were questionable (e.g. large variation in AI values among waveforms), the waveforms were re-analyzed by another technician. If discrepancies between the analyses occurred, the technicians reviewed the analyses together and if the differences could not be remediated, the data were excluded.

Pulse wave velocity (PWV) was determined according to standard procedures1 by using transcutaneous Doppler flow measurements (Model 806-CB, Parks Medical Electronics, Inc., Aloha, OR) at the right common carotid artery and the right femoral artery. Twenty Doppler wave forms were simultaneously recorded (Windaq software, version 2.31, DATAQ Instruments, Inc., Akron, OH) at the two sites. Pulse transit time was determined as the difference in pulse arrival times (based on the "foot" of the pulse wave, as determined from the second derivative of the pulse wave) for the carotid and femoral sites. Pulse transit distances between the aorta and the carotid site and the aorta and the femoral site were measured over the skin using the second intercostal space as a landmark for the aorta, with the difference between these distances being considered the pulse propagation distance [36]. PWV was calculated for each of the 20 pulses as the quotient of the propagation distance (in meters) and transit time (in seconds) and the 20 values were averaged. Quality

control procedures were identical to those described above for the AI method.

Vascular function. Brachial artery flow mediated dilation (FMD) was used to measure endotheliumdependent vasodilation and glyceryl trinitrate- (GTN)mediated dilation was used to measure endotheliumindependent vasodilation. The participant's right arm was immobilized with the shoulder abducted at 70-90° and elbow fully extended. Ultrasound images were acquired using an ultrasound system (Agilent SONOS 5500, Andover, Massachusetts) equipped with an 11-3L linear array transducer. The probe was secured using a stereotactic clamp (Noga Engineering, Ltd.) to maintain constant positioning over the artery throughout the procedure. Ultrasound images were fed to a computer for real time quantification of arterial diastolic diameter  $(30 \pm 2 \text{ measures/sec})$  using Vascular Imaging Analysis software (VIA, version 9.60) [37]. Baseline diameter was recorded for 2 minutes before a pneumatic cuff (Hokanson E20 Rapid Cuff Inflator and AG101 Cuff Inflator Air Source, PMS Instruments, Ltd., Maidenhead, UK) was inflated to 200 mmHg on the right forearm to occlude blood flow. After 5 minutes of occlusion, the pressure in the cuff was rapidly released and arterial diameter was recorded continuously for 5 minutes. FMD was calculated as the percent increase in diameter from baseline to peak diameter, where baseline diameter was the average diastolic diameter over the 2minute baseline and peak diameter was recorded as the 10-second average of the highest diastolic diameter after cuff deflation. After the FMD assessment, and after the diameter of the artery returned to baseline, a second 2-minute baseline data collection was performed, followed by administration of 0.4 mg sublingual GTN spray. Brachial artery diameter was evaluated again 5 minutes after GTN administration. GTN-mediated dilation was calculated as the percentage increase in arterial diameter from baseline to 5 minutes after GTN administration.

Statistical Power and Analyses. Sample size estimates were calculated for PWV, as a measure of arterial stiffness and for interleukin-6 (IL-6) and C-reactive protein (CRP) as inflammatory markers. Calculations were based on data collected in our laboratory during previous studies. These studies included a randomized controlled trial on the effects of 50 mg/d oral DHEA supplementation on markers of aging in older adults [31] and cross-sectional [38] and longitudinal studies [39] on the effects of calorie restriction on markers of aging in middle-aged adults. For all sample size calculations, the alpha error rate was set at 0.05, tests were designated as two-tailed, desired power was set to 0.80, and the ratio of participants to be randomized to the intervention and no-treatment control groups was 1:1. Results indicate that 21 subjects per group would be sufficient for detecting a 3.1 m/sec improvement in pulse wave velocity, 27 subjects per groups would be sufficient for detecting a 0.86 ng/mL reduction in IL-6, and 12 subjects per groups would be sufficient for detecting a 1.60 mg/L reduction in CRP. Therefore, our proposed sample size of 56 participants (28 per group) was sufficient to detect biologically relevant changes in arterial stiffness and inflammation.

Power analyses were performed for PWV, IL-6, and CRP to determine sample size. Respective results indicated that sample sizes of 21, 27, and 12 subjects per group would be adequate for detecting significant effects. Therefore, 28 subjects per group was chosen. Baseline characteristics were compared with independent t-tests and chi-square tests. Outcomes were analyzed with analysis of covariance, in which baseline values were included as a covariate. Paired t-tests were used for within-group paired comparisons. Significance was accepted at  $p \le 0.05$  and tests were two-tailed. Analyses were conducted with SAS for Windows XP Pro (version 9.3, SAS Institute, Cary, NC).

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#### **Conflicts of Interest Statement**

The authors declare no conflicts of interest.

### REFERENCES

**1.** Blendon RJ, Benson JM, Botta MD, Weldon KJ. Users' views of dietary supplements. JAMA Intern Med 2013; 173:74-6.

**2.** Denham BE. Dietary supplements--regulatory issues and implications for public health. JAMA 2011; 306:428-9.

**3.** Almeida IM, Barreira JC, Oliveira MB, Ferreira IC. Dietary antioxidant supplements: benefits of their combined use. Food Chem Toxicol 2011; 49:3232-7.

**4.** Baur JA, Sinclair DA. Therapeutic potential of resveratrol: the in vivo evidence. Nature reviews. Drug discovery. 2006; 5:493-506.

**5.** Das S, Das DK. Anti-inflammatory responses of resveratrol. Inflammation & allergy drug targets. 2007; 6:168-173.

**6.** Chan MM. Inhibition of tumor necrosis factor by curcumin, a phytochemical. Biochem. Pharmacol. 1995; 49:1551-1556.

**7.** Ruby AJ, Kuttan G, Babu KD, Rajasekharan KN, Kuttan R. Antitumour and antioxidant activity of natural curcuminoids. Cancer Lett. 1995; 94:79-83.

8. Balentine D, Paetau-Robinson I. Tea as a source of dietary antioxidants with a potential role in prevention of chronic

diseases. In: Mazza G, BD O, eds. Herbs, Botanicals & Teas. Lancaster: Technomic Publishing Co., Inc.; 2000:265-287.

**9.** Aneja R, Odoms K, Denenberg AG, Wong HR. Theaflavin, a black tea extract, is a novel anti-inflammatory compound. Crit. Care Med. 2004; 32:2097-2103.

**10.** Bors W, Heller W, Michel C, Saran M. Flavonoids as antioxidants: determination of radical-scavenging efficiencies. Methods Enzymol. 1990; 186:343-355.

**11.** García-Mediavilla V, Crespo I, Collado PS, Esteller A, Sánchez-Campos S, Tuñón MJ, González-Gallego J. The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. Eur. J. Pharmacol. 2007; 557:221-229.

**12.** Mackraj I, Govender T, Ramesar S. The antihypertensive effects of quercetin in a salt-sensitive model of hypertension. J. Cardiovasc. Pharmacol. 2008; 51:239-245.

**13.** Rivera L, Moron R, Sanchez M, Zarzuelo A, Galisteo M. Quercetin ameliorates metabolic syndrome and improves the inflammatory status in obese Zucker rats. Obesity. 2008; 16:2081-2087.

**14.** Hagen TM, Liu J, Lykkesfeldt J, Wehr CM, Ingersoll RT, Vinarsky V, Bartholomew JC, Ames BN. Feeding acetyl-L-carnitine and lipoic acid to old rats significantly improves metabolic function while decreasing oxidative stress. Proc. Natl. Acad. Sci. U. S. A. 2002; 99:1870-1875.

**15.** Aviram M, Dornfeld L, Rosenblat M, Volkova N, Kaplan M, Coleman R, Hayek T, Presser D, Fuhrman B. Pomegranate juice consumption reduces oxidative stress, atherogenic modifications to LDL, and platelet aggregation: studies in humans and in atherosclerotic apolipoprotein E-deficient mice. Am. J. Clin. Nutr. 2000; 71:1062-1076.

**16.** Aviram M, Dornfeld L. Pomegranate juice consumption inhibits serum angiotensin converting enzyme activity and reduces systolic blood pressure. Atherosclerosis. 2001; 158:195-198.

**17.** Lin CC, Wu SJ, Chang CH, Ng LT. Antioxidant activity of Cinnamomum cassia. Phytother. Res. 2003; 17:726-730.

**18.** Reddy AM, Seo JH, Ryu SY, Kim YS, Min KR. Cinnamaldehyde and 2 methoxycinnamaldehyde as NF-kappaB inhibitors from Cinnamomum cassia. Planta Med. 2004; 70:823-827.

**19.** Qin B, Nagasaki M, Ren M, Bajotto G, Oshida Y, Sato Y. Cinnamon extract (traditional herb) potentiates in vivo insulin-regulated glucose utilization via enhancing insulin signaling in rats. Diabetes Res. Clin. Pract. 2003; 62:139-148.

**20.** Qin B, Nagasaki M, Ren M, Bajotto G, Oshida Y, Sato Y. Cinnamon extract prevents the insulin resistance induced by a high-fructose diet. Horm. Metab. Res. 2004; 36:119-125.

**21.** Ashakumary L, Rouyer I, Takahashi Y, Ide T, Fukuda N, Aoyama T, Hashimoto T, Mizugaki M, Sugano M. Sesamin, a sesame lignan, is a potent inducer of hepatic fatty acid oxidation in the rat. Metabolism. 1999; 48:1303-1313.

**22.** Arachchige PG, Takahashi Y, Ide T. Dietary sesamin and docosahexaenoic and eicosapentaenoic acids synergistically increase the gene expression of enzymes involved in hepatic peroxisomal fatty acid oxidation in rats. Metabolism. 2006; 55:381-390.

**23.** Hirose N, Inoue T, Nishihara K, Sugano M, Akimoto K, Shimizu S and Yamada H. Inhibition of cholesterol absorption and synthesis in rats by sesamin. J. Lipid Res. 1991; 32:629-638.

**24.** Calder PC. Polyunsaturated fatty acids, inflammation, and immunity. Lipids. 2001; 36:1007-1024.

**25.** Noma K, Goto C, Nishioka K, Jitsuiki D, Umemura T, Ueda K, Kimura M, Nakagawa K, Oshima T, Chayama K, Yoshizumi M, Liao JK, Higashi Y. Roles of rho-associated kinase and oxidative stress in the pathogenesis of aortic stiffness. J Am Coll Cardiol. 2007; 49:698-705.

**26.** Abramson JL, Weintraub WS and Vaccarino V. Association between pulse pressure and C-reactive protein among apparently healthy US adults. Hypertension 2002; 39: 197-202.

**27.** Mattace-Raso FUS, van der Cammen TJM, van der Meer IM, Schalekamp MADH, Asmar R, Hofman A and Witteman JCM. C-reactive protein and arterial stiffness in older adults: The Rotterdam Study. Atherosclerosis 2004; 176: 111-116.

**28.** Davignon J, Ganz P. Role of endothelial dysfunction in atherosclerosis. Circulation 2004; 109:III27-32.

**29.** Hürlimann D, Forster A, Noll G, Enseleit F, Chenevard R, Distler O, Béchir M, Spieker LE, Neidhart M, Michel BA, Gay RE, Lüscher TF, Gay S, Ruschitzka F. Anti-tumor necrosis factor-alpha treatment improves endothelial function in patients with rheumatoid arthritis. Circulation 2002; 106:2184-7.

**30.** Eskurza I, Monahan KD, Robinson JA, Seals DR. Effect of acute and chronic ascorbic acid on flow-mediated dilatation with sedentary and physically active human ageing. J Physiol 2004; 556:315-24.

**31.** Weiss EP, Villareal DT, Ehsani AA, Fontana L, Holloszy JO. Dehydroepiandrosterone replacement therapy in older adults improves indices of arterial stiffness. Aging Cell 2012; 11: 876-84.

**32.** Williamson G, Manach C. Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. Am J Clin Nutr 2005; 81:243-55.

**33.** Office UGA. Herbal dietary supplements: examples of deceptive or questionable marketing practices and potentially dangerous advice. Publication No GAO-10-662T. http://www.gao.gov/new.items/d10662t.pdf (12 August 2013)

**34.** Laurent S, Cockcroft J, Van Bortel L, Boutouyrie P, Giannattasio C, Hayoz D, Pannier B, Vlachopoulos C, Wilkinson I, Struijker-Boudier H; European Network for Non-invasive Investigation of Large Arteries. Expert consensus document on arterial stiffness: methodological issues and clinical application. Eur Heart J 2006; 27:2588-2605.

**35.** Wilkinson IB, Mohammad NH, Tyrrell S, Hall IR, Webb DJ, Paul VE, Levy T, Cockcroft JR. Heart rate dependency of pulse pressure amplification and arterial stiffness. Am J Hypertens 2002; 15:24-30.

**36.** Karamanoglu M. Errors in estimating propagation distance in pulse wave velocity. Hypertension 2003; 41:e8.

**37.** Sidhu JS, Newey VR, Nassiri DK, Kaski JC. A rapid and reproducible on line automated technique to determine endothelial function. Heart 2002; 88:289-92.

**38.** Fontana L, Meyer TE, Klein S, Holloszy JO. Long-term calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. Proc Natl Acad Sci USA 2004; 101:6659-63.

**39.** Fontana L, Villareal DT, Weiss EP, Racette SB, Steger-May K, Klein S, Holloszy JO; Washington University School of Medicine CALERIE Group.Calorie restriction or exercise: effects on coronary heart disease risk factors. A randomized, controlled trial. Am J Physiol Endocrinol Metab 2007; 293:197-202.