# Longitudinal decline of leukocyte telomere length in old age and the association with sex and genetic risk

Kari Berglund<sup>1</sup>, Chandra A. Reynolds<sup>2</sup>, Alexander Ploner<sup>1</sup>, Lotte Gerritsen<sup>1,3</sup>, Iiris Hovatta<sup>4,5</sup>, Nancy L. Pedersen<sup>1</sup>, and Sara Hägg<sup>1</sup>

<sup>1</sup>Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, 171 77 Stockholm, Sweden <sup>2</sup>Department of Psychology, University of California-Riverside, Riverside, CA 92521, USA <sup>3</sup>Department of Clinical Psychology, Faculty of Social Sciences, Utrecht University, Utrecht, the Netherlands <sup>4</sup>Department of Biosciences, Viikki Biocenter, University of Helsinki, Helsinki, Finland <sup>5</sup>Department of Health, National Institute for Health and Welfare, Helsinki, Finland

**Key words:** telomere length, longitudinal decline, aging, genetic risk score, latent growth curve **Received:** 04/19/16; Accepted: 06/20/16; Published: 07/07/16 **Correspondence to:** Sara Hägg, PhD; **E-mail:** <u>Sara.Hagg@ki.se</u>

Abstract: Telomeres are DNA-protein structures at the ends of chromosomes. Leukocyte telomere length (LTL) shortening has been associated with advanced age. However, most studies use cross-sectional data, hence, the aim of our study was to model longitudinal trajectories of LTL attrition across 20 years at old age. Assessments of LTL were done by qPCR in SATSA (Swedish Adoption/Twin Study of Aging; N=636 individuals). Cross-sectional and longitudinal associations with age were estimated, the latter using latent growth curve analysis. A genetic risk score (GRS) for LTL was further assessed and included in the models. We confirmed an inverse cross-sectional association of LTL with age (B=-0.0022 T/S-ratio; 95% CI: -0.0035, -0.0009, p-value=0.0008). Longitudinal LTL analyses adjusted for sex (1598 samples;  $\leq 5$  measurements) suggested modest average decline until 69 years of age but accelerating decline after 69 years, with significant inter-individual variation. Women had on average ~6% T/S-ratio units longer LTL at baseline, and inclusion of the GRS improved the model where four risk alleles was equivalent to the effect size difference between the sexes. In this cohort of old individuals, baseline LTL varied with age, sex and genetic background. The rate of change of LTL accelerated with age and varied considerably between individuals.

### **INTRODUCTION**

Telomeres are DNA-protein structures of tandem hexanucleotide repeats at the ends of eukaryotic chromosomes, providing protection from degradation and recombination during cell division [1]. In somatic cells, telomeres are inadequately replicated, and, as a result, telomere length declines with each cell division [2]. Leukocyte telomere length (LTL) in humans is inversely associated with age and is influenced by genetic [3] and environmental factors [4]. Moreover, telomeres are shorter in men than in women; a consequence of an accelerated attrition rate in men throughout the life course [5-9]. Shorter telomeres have been implicated in several diseases, including cardiovascular disease, but it is unclear what direct impact, if any, telomeres have on the development of age-related diseases [8, 10-14].

A recent meta-analysis of 124 cross-sectional studies has shown that telomere lengths are significantly shorter with age [15], and at least six longitudinal studies report a decrease in telomere length with advancing age [16-21]. However, previous efforts have had shorter followup time and only included two to three time points per study subject, which is insufficient for estimating trajectories of change. In the present study, we examine both the cross-sectional and longitudinal associations between LTL and chronological age in elderly Swedish twins with up to five LTL measurements per individual across 20 years. Thus, we sought to characterize LTL trajectories with advancing age, and to test whether individual differences in trajectories may be accounted for in part by sex and a genetic risk score (GRS) for LTL identified in a prior genome-wide association study [3].

### **RESULTS**

#### **Cross-sectional analysis**

The Swedish Adoption/Twin Study of Aging (SATSA) [22] is a longitudinal study started in 1984 including twins from mid-life and onwards. It consists of twin pairs reared apart matched with twin pairs reared together, and is collected from all over Sweden. The individuals are followed up to 30 years with in-person testing (IPT) of cognitive ability, physical fitness, anthropometrics, health data and questionnaires up to 10 times. Data on LTL are available from IPT 3. 5. 6. 8. and 9, and were attained by qPCR techniques (see Methods for more details). Basic characteristics of a cross-sectional sample of LTL measures at baseline (n=636 individuals from same sex twin pairs with 84 monozygotic and 152 dizygotic complete pairs) are presented in Table 1. Through linear regression adjusted for family relatedness and sex, an inverse association between LTL and age was found (B=-0.0022 T/Sratio/year; 95% confidence interval [CI]: -0.0035, -0.0009, p-value=0.0008) and female sex was associated with a greater overall LTL (0.0499; 95% CI: 0.0200, 0.0797; p-value=0.001). Stratifications on zygosity did not significantly impact the model (data not shown). By using genotype data, an un-weighted GRS summarizing risk alleles from seven single nucleotide polymorphisms (SNPs) associated with LTL [3] was calculated per individual (M=8.42, SD=1.43; Supplementary Table 1).

The overall effect of the GRS on LTL in SATSA (-0.046 SD-decrease in T/S-ratio per additional risk allele; 95% CI: -0.111, 0.019) was similar to the effect seen in the original LTL genome-wide association study (GWAS) (-0.070; 95% CI: -0.077, -0.063) although the CI's were wider (Supplementary Figure 1, upper panel). Hence, the GRS did not significantly contribute to the cross-sectional model and did not alter the estimates markedly (Supplementary Table 2). Additional sensitivity analyses are presented in Supplementary Tables 2 and 3.

	SATSA
	(N = 636)
Years of data collection	1992 - 2012
Telomere length	$0.70 \pm 0.17$
$(\text{mean} \pm \text{sd})^{\$}$	$0.70 \pm 0.17$
Age (mean $\pm$ sd)	$68.86 \pm 9.67$
Women (%)	372 (58%)
GRS (mean $\pm$ sd)*	$8.42 \pm 1.43$

Table 1. SATSA cross-sectional characteristics

<sup>S</sup>Mean telomere length is adjusted for batch effect and re-scaled back to T/S-ratio. \*Genetic information was available for 585 individuals. GRS: genetic risk score.

#### Longitudinal analysis

Longitudinal sample characteristics are reported by measurement occasions and sex in Table 2. Women in general have a higher mean LTL than men at similar ages, whether by measurement occasions or by IPT's (Supplementary Table 4). The distribution of longitudinal samples across the 636 individuals was 31.1%

	Measurement Occasion							
-	1 <sup>st</sup> Measure	2 <sup>nd</sup> Measure	<sup>d</sup> Measure 3 <sup>rd</sup> Measure		5 <sup>th</sup> Measure			
Men	(N=264)	(N=181)	(N=128)	(N=66)	(N=22)			
Telomere length (mean±sd) <sup>\$</sup>	0.68±0.18	0.70±0.12	0.70±0.15	0.68±0.09	0.70±0.19			
Age (mean±sd)	67.44±8.79	71.84±8.70	74.36±7.93	76.52±7.82	78.42±5.74			
Women	(N=372)	(N=257)	(N=174)	(N=98)	(N=36)			
Telomere length (mean±sd) <sup>\$</sup>	0.72±0.16	0.72±0.13	0.72±0.13	0.73±0.13	0.68±0.16			
Age (mean±sd)	69.86±10.13	73.75±9.45	76.55±8.85	78.73±8.52	80.91±8.07			

Table 2. SATSA longitudinal characteristics by sex and measurement occasions

<sup>\$</sup>Mean telomere length is adjusted for batch effect and re-scaled back to T/S-ratio.

(N=198) with one measurement; 21.4% (N=136) with two; 21.7% (N=138) with three; 16.7% (N=106) with four; and 9.1% (N=58) with five measurements, hence, 47% had at least three measurements available.

An inverse relationship between LTL and age was seen when plotting individual trajectories of all samples stratified by sex (Figure 1). The overall trend, presented here as a smoothing curve, is close to linear for both men and women, with similar slopes, but with a slightly lower intercept and accelerating attrition in older age in men. Individual trajectories with linear trends, including a split at the centering age defined as the median age at IPT3 (69.3 years), are displayed in the supplement (Supplementary Figure 2). To quantify telomere attrition longitudinally, latent growth curve (LGC) models were tested (Figure 2, Methods, and Supplementary Data). In brief, an LGC model is a mixed model including three variables: 1) the intercept with the LTL value at the age of 69.3 years, 2) a linear slope before the centering age (69.3 years), and 3) a linear slope after the centering age. Variances and cross-correlations were presented for all three variables and twin relatedness were adjusted for by estimating between- and within-pair variances. At first, an intercept-only (no change) model (Model 1) was compared to a one-slope model without any change point included (Model 2), allowing for a linear rate of change across the whole age span. The improvement in fit was significant (p-value= $2 \cdot 10$ -8; Table 3). Model 2 also

Table 3. Latent growth curve models for leukocyte telomere length and age (N=1598)

Models tested								
Model	-2LL	Parms	AIC	BIC	Δ-2LL	df		
1: Intercept only	-1865.9	5	-1855.9	-1836.2				
2: Intercept + slope 1	-1910.5	10	-1892.5	-1857.1	44.6 <sup>§</sup>	5***		
3: Intercept + slope 1 & 2	-1930.4	17	-1900.4	-1841.4	19.9 <sup>£</sup>	7**		

<sup>§</sup>Model 1 vs. model 2. <sup>f</sup>Model 2 vs. model 3. Chi-square tests for deviance: \*\*p-value<0.01; \*\*\*p-value<0.001. In all models, sex as covariate as well as random effects between twin pairs and between individuals were included. LL: log-likelihood. Parms: parameters; AIC: Akaike Information Criterion; BIC: Bayesian Information Criterion; *df*: degrees of freedom.





revealed significant variation between twin pairs in LTL change, suggestive of familial factors (Supplementary Table 5). Next, the one-slope model (Model 2) was compared to a two-slope model allowing for two linear rates, one before and one after age 69.3, respectively (Model 3). Again, the fit was substantially improved (pvalue=0.006; Table 3). The fixed effect (average) for slope 1 in model 3 (young-old age<69.3) was not significant (p-value=0.30), but it was for slope 2 (oldold age>69.3; B=-0.0021 T/S-ratio/year; 95% CI: -0.0034, -0.0008, p-value=0.002). The variance around the intercept for both familial (between twin pairs) and environmental (within twin pair) effects contributed strongly (p-value<0.001 for both effects) to the model (Table 4). However, there was also variation detected for both slope 1 and 2 for the within-pair random effect (p-value<0.01 and <0.05 respectively), suggesting that non-familial/environmental factors are important to rate of change at both young- and old-old age, although the effect was bigger at young-old age. Indeed, sensitivity analyses indicated that it was necessary to retain the sets of between and within-pair random effects for both slope 1 and slope 2 ( $\Delta \chi^2$  p-value<0.0005), indicating individual differences in change across both age periods. Altogether, the average difference between slope 1 and 2 in model 3 was not great, as was obvious from the raw trajectories depicted (Figure 1 and Supplementary Figure 2). Rather, the variation around the slopes was appreciable and larger for slope 1 than slope 2 (see Supplementary Table 6 for complete model estimates). Further sensitivity analyses suggested that the same trends remained when altering the centering age to be five years earlier or later, and again, as in the



**Figure 2.** Latent growth curve analysis of telomere length attrition with a group intercept (I) and two slopes (S<sub>1</sub>) and (S<sub>2</sub>) as well as sex as covariate (C). Observed data are denoted by Y<sub>0</sub> through Y<sub>4</sub>. M<sub>1</sub>: mean intercept; M<sub>S1</sub>: mean slope 1; M<sub>S2</sub>: mean slope 2; M<sub>c</sub>: mean covariate. V stands for variance of each component respectively. r denotes the correlation between the intercept and the slopes. B1t through B2t represent the age basis coefficients with change over time t. U<sub>0</sub> through U<sub>4</sub> represent random components from the telomere length measurements, constrained to be equal for each assessment.

		Fixed Ef	fects			
Interce (estimate (9		Slope 1 (estimate (95% CI))	Slop (estimate (		Sex (estimate (95% CI))	
0.7287 (0.7095-0.7479)***		-0.0012 (-0.0033-0.0010)	-0.0021 (-0.00340.0008)**		-0.0409 (-0.06370.0182)**	
		Random H	Effects			
varI <sub>t</sub>	varI <sub>p</sub>	varS <sub>1t</sub>	varS <sub>1p</sub>	varS <sub>2t</sub>	varS <sub>2p</sub>	
0.0060***	0.0082***	0.00006**	* 0.00001 0.00002*		2* 0.00002	

Table 4. Fixed and random effects of the full spline two-slope model (Model 3) from latent growth curve analysis of leukocyte telomere length and age

\*p-value<0.05; \*\*p-value<0.01; \*\*\*p-value<0.001;

Note. Random effects: varI = variance around the intercept; varS<sub>1</sub> = variance around slope 1; varS<sub>2</sub> = variance around slope 2; In all models, sex as covariate as well as random effects between twin pairs (subscript 'p') and within twin pairs (subscript 't') were included.

cross-sectional analysis, zygozity did not matter (data not shown). Finally, women had on average 0.0409 higher T/S-ratio than men (p-value=0.0005; Table 4), corresponding to a 6% difference in baseline T/S-ratio

at 69.3 years. The estimate was similar although smaller to the cross-sectional estimate (0.0499). However, no interaction was seen between sex and the rate of decline in old-old age (Table 5).

Table 5. Additional adjusted two-slope models (Model 3) of leukocyte telomere length and age (N=1504)

	Models tested						
Model	-2LL	Parms	AIC	BIC	Reference model	Δ-2LL	df
3	-1834.2	17	-1802.2	-1740.1			
3+slope2*sex	-1834.4	18	-1800.4	-1734.4	3	0.2	1
3+GRS	-1843.1	18	-1809.1	-1743.2	3	8.9	1**
3+GRS+slope2*GRS	-1843.9	19	-1807.9	-1738.0	3+GRS	0.8	1
3+GRS+sex*GRS	-1843.2	19	-1807.2	-1737.3	3+GRS	0.1	1

Chi-square tests for deviance: **\*\***p-value<0.01; In all models, sex as covariate as well as random effects between twin pairs and between individuals were included. LL: log-likelihood. Parms: parameters; AIC: Akaike Information Criterion; BIC: Bayesian Information Criterion; *df*: degrees of freedom; GRS: Genetic Risk Score.



**Figure 3.** Predicted trajectories for men and women based on parameter estimates from the two-slope model of leukocyte telomere length (LTL) including sex and genetic risk score (GRS) effects. Male sex and addition of risk alleles in the GRS each result in shorter LTL. A decline after the centering age of 69.3 years is apparent for both men and women. Age in years is on the x-axis, and the plate-adjusted residuals of LTL re-scaled back to T/S-ratio is on the y-axis. The dashed line indicates centering age and the left corner panel is a zoomed version.



**Figure 4.** Individual relative leukocyte telomere length (LTL) change in the longitudinal cohort. The difference in LTL measurement between any two time-points in the same individual is on the x-axis. The frequency is on the y-axis. Telomere elongation is exhibited in samples with delta LTL>0, and telomere attrition is exhibited in samples with delta LTL<0. The sample distribution shows an overall elongation in 46%.

#### Genetic risk scores in longitudinal models

To test whether genetic predisposition towards shorter LTL had an impact in the longitudinal model, the GRS for LTL was entered into the fixed effects of the twoslope model (Supplementary Table 7). The fit improved markedly (p-value=0.003), but GRS did not interact with sex or age (Table 5). The different trajectories seen for model 3 depending on age, sex and GRS parameter estimates are displayed in Figure 3. Men have generally shorter LTL than women, and the trajectories are parallel. One risk allele increase of the GRS results in additional -0.011 decrease in T/S-ratio (95% CI: -0.018, -0.004, p-value=0.003), and four additional risk alleles are equivalent to the difference in T/S-ratio in men compared to in women. Overall, the GRS estimate from the LGC model is similar to the estimate from the crosssectional model; however, the CI's are smaller (Supplementary Figure 1, lower panel). We further investigated effects of individual genetic variants (those included in the GRS) on the two-slope model and found the TERC locus to be the most important contributor with significant effect on its own (B=-0.020; 95% CI: -0.037, -0.003, p-value=0.02, Supplementary Figure 1, lower panel). However, none of the SNPs was individually as good as the GRS was for improving the

#### **Telomere elongation**

Longitudinally, many individuals exhibited telomere elongation from one occasion to the next (Figure 4). Elongation was seen in 46% of the within-individual sample comparisons. including all possible combinations, ranging from 44-47% depending on the number of years between measurements (Supplementary Figure 3). The coefficient of variation from the qPCR analyses was ~7%, suggesting that the elongation seen was likely a biological phenomenon although technical bias from measurement imprecision and/or possible differences in sample collection between IPT's could not be completely ruled out.

### **DISCUSSION**

In the current study, we first examined the crosssectional associations between LTL and age, and, like previous reports, we found an inverse relationship with increasing age. Second, using LGC analysis with up to five measurements across 20 years, we found that LTL decreases with age in a two-slope model with a small acceleration of decline after 69.3 years of age. Men have shorter telomere lengths than women, and genetic variation has an additional influence on overall LTL. Several earlier studies have reported an inverse association between age and telomere length [15-20, 23], as did we, and we further demonstrated that women have longer LTL, which is in line with earlier research [5-7, 15, 23]. Taking our results and prior literature together, shorter telomeres in men could result from very small but consistent attrition throughout adulthood rather than a steeper decline compared to women in old age. Moreover, previous literature from cross-sectional and longitudinal studies has suggested a linear relationship between telomere length and age [15-20]. We found both the one-slope and the two-slope models to be significant, with a substantially better fit of the latter. While the overall average trend was linear, there was systematic variability around the average trend, better described in a two-slope model accounting for more individual differences.

The magnitude of this age-related decline was small overall, and with slight acceleration in the old-old. This observation is in line with earlier research in the field where faster decline in LTL is believed to take place in childhood and old age [23]. The age-related telomere loss in SATSA was similar in both cross-sectional and longitudinal analyses, and somewhat smaller than earlier longitudinal estimates of T/S-ratio attrition rate [19]. A likely reason for differing results may be measurement imprecision from using the qPCR technique and fluctuations in lymphocyte subpopulations. The two-slope trajectory analyses supported both familial and non-familial influences on LTL, with equal contributions to average LTL level (at age 69) and non-familial sources featuring more prominently in the change before age 69 than after age 69. This suggests that in young-old age, individualspecific lifestyle factors may prove more relevant to accelerated LTL shortening above and beyond familial and environmental contributions to overall LTL; however, in old-old age, familial factors may become increasingly salient to accelerated LTL shortening. Moreover, we note that the variation in rate of change was larger in young-old age; hence, evaluating variation in trajectories beyond the assumption of simple linearity and average trends is important for understanding etiological underpinnings.

Previous studies have investigated the heritability of LTL; a meta-analysis of five European ancestry cohorts reported it to be 0.70 [24], our study contributes new information on how telomere attrition *rates* may vary between twins. The same meta-analysis also looked at LTL correlations in spouses and found it to be substantial with larger effects in old age, in line with our findings of environmental contributions to the slopes.

Moreover, the GRS significantly improved the twoslope model when added to the fixed effect, suggesting that genetic background contributes to the overall LTL, but not its rate of change: we did not observe any significant interaction between slope 2 and GRS. In addition, when testing the individual gene effects on the LTL trajectories, we found them to be proportional to the genetic variant effect sizes, with the TERC gene (telomerase RNA component) as the most important contributor to the model. The estimated decreased effect in T/S-ratio from one additional risk allele increase in GRS was similar in both longitudinal (-0.011) and cross-sectional models (-0.015). Furthermore, we could not find any interaction between sex and GRS, which indicates that the genetic effect on LTL attrition is equal in both men and women. However, the genetic risk component can have larger effects than sex on telomere shortening; the addition of four risk alleles cancel out the sex effect indicating that individual predisposition to LTL attrition is of great importance.

### **Telomere elongation**

Telomere elongation has been observed and discussed in other longitudinal cohorts with LTL assayed by qPCR technique [25-29]. They all report evidence of small increases in telomere length, consistent with the lengthening that we observed. It still remains unclear what causes leukocyte telomere elongation. However, a recent mouse study demonstrated that, although increased stress levels accelerate telomere attrition, telomeres recover back to normal length when the stress subsides [30]. Nonetheless, the interesting question is whether the individual cells extend their telomeres by up-regulating the telomerase enzyme or whether there is renewal of the leukocyte population from the stem cell pool, leading to overall longer LTL due to faster turnover and larger number of young cells. In our sample, the elongation exceeds what would be expected from the coefficient of variation (6.98%) of the qPCR method, but technical variability could also be inferred from differences in sample collection over time and changes in leukocyte turnover. Additional research is necessary to explore why telomere elongation has been observed in this and in other studies

#### Strengths and limitations

The strength of this study is the inclusion of repeated LTL samples (up to five time points) over a 20-year time period, allowing exploration of trajectories as opposed to the maximum of three time points in previous studies [17, 18]. Moreover, the study design with twin pairs allowed us to investigate the familial and non-familial contributions to individual differences;

at the same time, inclusion of GRS enabled quantification of genetic propensity to overall telomere shortening.

Limitations of the study include the generalizability of LTL to other tissues. Although LTL is strongly correlated with telomere length in muscle, skin, and subcutaneous fat, it is unclear what sort of longitudinal decline telomeres may experience in other cell types that undergo mitosis at different frequencies [31]. Additionally, because of our focus on older Swedish adults, we cannot generalize our results to younger adults or other ancestries. Finally, our slope estimates in both the cross-sectional and longitudinal models are based on relative attrition measures due to use of the qPCR-based LTL measurement. This limits the comparability of our results with other studies using other measurement techniques.

In summary, we present for the first time trajectories of longitudinal telomere decline in old age using a twin design including investigation of genetic contributions. The annual decline is small and linear with slightly accelerated decline after 69 years. Male sex and inclusion of a GRS for LTL in the models were independently associated with shorter telomeres.

## **METHODS**

Study samples. The Swedish Twin Registry (STR) [32] is a population-based national registry established in the late 1950s and consists of twins born 1886-2000. SATSA (The Swedish Adoption/Twin Study of Aging) is a sub-study of STR and was started in 1984 with a first questionnaire sent out to twins reared apart matched with twins reared together (N=2018) [22]. Two years later the first in-person testing (IPT) was made on 645 individuals (303 pairs) over the age of 50 years. Following approximately every third year, a new IPT was administered and a total of ten IPTs were done until 2014 with questionnaires, anthropometric measures, blood sampling, physical function measurements and cognitive testing. More information on the SATSA study, including a list of references, can be found online (http://ki.se/en/meb/satsa-the-swedish-adoptiontwinstudy-of-aging). LTL assessments were done in individuals attending at least one of IPT 3, 5, 6, 8 or 9 (N=636 individuals corresponding to 1598 samples). Cross-sectional LTL data was based on the first available measure. Genotype information was gathered through the Illumina CardioMetabochip [33] and was available for 585 individuals.

<u>Telomere assessment.</u> LTL measurements were derived from blood leukocytes using a qPCR assay, comparing a telomere length PCR product (T) against a PCR product of a reference gene (S) to produce a T/S-ratio [34]. A detailed description of the LTL assessment is presented in the Supplementary Data.

<u>Genetic risk score calculation.</u> An individual, nonweighted GRS was calculated using seven genetic variants previously known to be associated with LTL [3]; directly genotyped (rs2736100, rs2281929) and imputed variants (rs11125529, rs10936599, rs7675998, rs9420907, rs8105767) (Supplementary Table 1). The SATSA study was not included in the original GWAS. For each individual, all risk alleles for shorter LTL were counted and summed. The GRS variable was centered on its mean before inclusion into the models.

<u>Statistical analyses.</u> In the telomere samples, plate ID was used to adjust for batch effect and residuals were re-scaled back to T/S-ratio by adding the mean to each value (Supplementary Data, Supplementary Figure 4). Hence, the weighted estimates from the models are reported as B values not to be confused with standardized  $\beta$  values. Moreover, samples were excluded if the T/S-ratio exceeded four SD's from the mean within the study (cross-sectional: [N=6]) or within a given age group (longitudinal: 50-59 years [N=1], 60-69 years [N=3], 70-79 years [N=4], 80+ years [N=3]).

Linear regressions with re-scaled LTL residuals as the outcome variable and sex as a covariate were performed on the cross-sectional data to evaluate age and LTL relationships. Twinness was controlled for by robust sandwich estimates of standard errors using *PROC SURVEYREG* including cluster on twin pair in SAS 9.4 (SAS Inc, Cary, NC).

Latent growth curve models have been used previously with SATSA data to model age changes in cognitive functioning [35, 36]. Specifically, regression models were fitted to each individual's longitudinal LTL trajectory, resulting in an average model for the sample (fixed effects) plus individual deviations from the average model (random effects). Familial influences on LTL were accounted for by allowing for random effects between twin pairs and non-familial/environmental factors estimated via within-pair random effects. Sex was included in all models. A best-fit model was selected via chi-square difference tests ( $\Delta \chi^2$ ), comparing the -2 log-likelihood (-2LL) of an intercept-only model, a one-slope model, and a two-slope model, with age variables centered at the median age at IPT3 (69.3 vears). The one-slope model estimated a single linear rate of change effect across age, while the two-slope model allowed for two linear rates, before and after age 69.3, respectively. Comparisons of additional two-slope models with GRS were carried out likewise. Individual trajectories were plotted together with a smoothing curve defined by a locally weighted regression model, summarizing the average behavior of LTL as a function of age without explicit assumptions about its overall shape. Predicted trajectory curves were calculated and further plotted from the parameter estimates of the two-slope model. All LGC analyses were performed using *PROC MIXED* in SAS 9.4, and R version 3.2 was used to create figures. The cut-off level used for overall significance were p<0.05 as is commonly used for these types of models [37].

#### ACKNOWLEDGEMENTS

We thank Jenni Lahtinen for technical help in performing the telomere length measurements.

#### Funding

This work was supported by NIH grants R01 AG04563, AG10175, AG028555, the MacArthur Foundation Research Network on Successful Aging, the Swedish Council for Working Life and Social Research (FAS/FORTE) (97:0147:1B, 2009-0795, 2013-2292), the Swedish Research Council (825-2007-7460, 825-2009-6141, 521-2013-8689, 2015-03255), Loo & Hans Osterman Foundation and the Foundation for Geriatric Diseases. LG was supported by a Marie Curie intra-European Fellowship of the European Community's Seventh Framework Programme under contract number PIEF-GA-2011-300355. Telomere assessments were performed through support of a Distinguished Professor Award from the Karolinska Institutet to NLP.

#### **Conflict of Interests Statement**

The authors have no conflict of interests to declare.

#### **REFERENCES**

**1.** Blackburn EH. Structure and function of telomeres. Nature. 1991; 350:569-73.

**2.** Olovnikov AM. A theory of marginotomy. The incomplete copying of template margin in enzymic synthesis of polynucleotides and biological significance of the phenomenon. Journal of theoretical biology. 1973; 41:181-90.

**3.** Codd V, Nelson CP, Albrecht E, Mangino M, Deelen J, Buxton JL, Hottenga JJ, Fischer K, Esko T, Surakka I, Broer L, Nyholt DR, Mateo Leach I, et al. Identification of seven loci affecting mean telomere length and their association with disease. Nat Genet. 2013; 45:422-27.

**4.** Valdes AM, Andrew T, Gardner JP, Kimura M, Oelsner E, Cherkas LF, Aviv A and Spector TD. Obesity, cigarette smoking, and telomere length in women. Lancet. 2005; 366:662-64.

**5.** Hoffmann J, Erben Y, Zeiher AM, Dimmeler S and Spyridopoulos I. Telomere length-heterogeneity among myeloid cells is a predictor for chronological ageing. Experimental gerontology. 2009; 44:363-66.

**6.** Benetos A, Okuda K, Lajemi M, Kimura M, Thomas F, Skurnick J, Labat C, Bean K and Aviv A. Telomere length as an indicator of biological aging: the gender effect and relation with pulse pressure and pulse wave velocity. Hypertension. 2001; 37:381-85.

**7.** Moller P, Mayer S, Mattfeldt T, Muller K, Wiegand P and Bruderlein S. Sex-related differences in length and erosion dynamics of human telomeres favor females. Aging (Albany NY). 2009; 1:733-39. doi: 10.18632/aging.100068

**8.** Bekaert S, De Meyer T, Rietzschel ER, De Buyzere ML, De Bacquer D, Langlois M, Segers P, Cooman L, Van Damme P, Cassiman P, Van Criekinge W, Verdonck P, De Backer GG, et al. Telomere length and cardiovascular risk factors in a middle-aged population free of overt cardiovascular disease. Aging cell. 2007; 6:639-47.

**9.** Okuda K, Bardeguez A, Gardner JP, Rodriguez P, Ganesh V, Kimura M, Skurnick J, Awad G and Aviv A. Telomere length in the newborn. Pediatric research. 2002; 52:377-81.

**10.** Samani NJ, Boultby R, Butler R, Thompson JR and Goodall AH. Telomere shortening in atherosclerosis. Lancet. 2001; 358:472-73.

**11.** Fyhrquist F and Saijonmaa O. Telomere length and cardiovascular aging. Annals of medicine. 2012; 44:138-42.

**12.** Bhupatiraju C, Saini D, Patkar S, Deepak P, Das B and Padma T. Association of shorter telomere length with essential hypertension in Indian population. American journal of human biology : the official journal of the Human Biology Council. 2012; 24:573-78.

**13.** Weischer M, Bojesen SE, Cawthon RM, Freiberg JJ, Tybjaerg-Hansen A and Nordestgaard BG. Short telomere length, myocardial infarction, ischemic heart disease, and early death. Arteriosclerosis, thrombosis, and vascular biology. 2012; 32:822-29.

**14.** Fitzpatrick AL, Kronmal RA, Gardner JP, Psaty BM, Jenny NS, Tracy RP, Walston J, Kimura M and Aviv A. Leukocyte telomere length and cardiovascular disease in the cardiovascular health study. Am J Epidemiol. 2007; 165:14-21.

**15.** Muezzinler A, Zaineddin AK and Brenner H. A systematic review of leukocyte telomere length and age in adults. Ageing Res Rev. 2013; 12:509-19.

**16.** Rius-Ottenheim N, Houben JM, Kromhout D, Kafatos A, van der Mast RC, Zitman FG, Geleijnse JM, Hageman GJ and Giltay EJ. Telomere length and mental well-being in elderly men from the Netherlands and Greece. Behavior genetics. 2012; 42:278-86.

**17.** Chen W, Kimura M, Kim S, Cao X, Srinivasan SR, Berenson GS, Kark JD and Aviv A. Longitudinal versus cross-sectional evaluations of leukocyte telomere length dynamics: age-dependent telomere shortening is the rule. J Gerontol A Biol Sci Med Sci. 2011; 66:312-19.

**18.** McCracken J, Baccarelli A, Hoxha M, Dioni L, Melly S, Coull B, Suh H, Vokonas P and Schwartz J. Annual ambient black carbon associated with shorter telomeres in elderly men: Veterans Affairs Normative Aging Study. Environmental health perspectives. 2010; 118:1564-70.

**19.** Kuznetsova T, Codd V, Brouilette S, Thijs L, Gonzalez A, Jin Y, Richart T, van der Harst P, Diez J, Staessen JA and Samani NJ. Association between left ventricular mass and telomere length

in a population study. American journal of epidemiology. 2010; 172:440-50.

**20.** Ehrlenbach S, Willeit P, Kiechl S, Willeit J, Reindl M, Schanda K, Kronenberg F and Brandstatter A. Influences on the reduction of relative telomere length over 10 years in the population-based Bruneck Study: introduction of a well-controlled high-throughput assay. International journal of epidemiology. 2009; 38:1725-34.

**21.** Steenstrup T, Hjelmborg JV, Mortensen LH, Kimura M, Christensen K and Aviv A. Leukocyte telomere dynamics in the elderly. European journal of epidemiology. 2013; 28:181-87.

**22.** Pedersen NL, McClearn GE, Plomin R, Nesselroade JR, Berg S and DeFaire U. The Swedish Adoption Twin Study of Aging: an update. Acta Genet Med Gemellol. 1991; 40:7-20.

**23.** Sanders JL and Newman AB. Telomere length in epidemiology: a biomarker of aging, age-related disease, both, or neither? Epidemiol Rev. 2013; 35:112-31.

**24.** Broer L, Codd V, Nyholt DR, Deelen J, Mangino M, Willemsen G, Albrecht E, Amin N, Beekman M, de Geus EJ, Henders A, Nelson CP, Steves CJ, et al. Meta-analysis of telomere length in 19,713 subjects reveals high heritability, stronger maternal inheritance and a paternal age effect. Eur J Hum Genet. 2013; 21:1163-68.

**25.** Aviv A, Chen W, Gardner JP, Kimura M, Brimacombe M, Cao X, Srinivasan SR and Berenson GS. Leukocyte telomere dynamics: longitudinal findings among young adults in the Bogalusa Heart Study. Am J Epidemiol. 2009; 169:323-29.

**26.** Farzaneh-Far R, Lin J, Epel E, Lapham K, Blackburn E and Whooley MA. Telomere length trajectory and its determinants in persons with coronary artery disease: longitudinal findings from the heart and soul study. PLoS One. 2010; 5:8612.

**27.** Nordfjall K, Svenson U, Norrback KF, Adolfsson R, Lenner P and Roos G. The individual blood cell telomere attrition rate is telomere length dependent. PLoS Genet. 2009; 5:1000375.

**28.** Hovatta I, de Mello VD, Kananen L, Lindstrom J, Eriksson JG, Ilanne-Parikka P, Keinanen-Kiukaanniemi S, Peltonen M, Tuomilehto J and Uusitupa M. Leukocyte telomere length in the Finnish Diabetes Prevention Study. PLoS One. 2012; 7:34948.

**29.** Epel ES, Blackburn EH, Lin J, Dhabhar FS, Adler NE, Morrow JD and Cawthon RM. Accelerated telomere shortening in response to life stress. Proc Natl Acad Sci U S A. 2004; 101:17312-15.

**30.** Cai N, Chang S, Li Y, Li Q, Hu J, Liang J, Song L, Kretzschmar W, Gan X, Nicod J, Rivera M, Deng H, Du B, et al. Molecular signatures of major depression. Curr Biol. 2015; 25:1146-56.

**31.** Daniali L, Benetos A, Susser E, Kark JD, Labat C, Kimura M, Desai K, Granick M and Aviv A. Telomeres shorten at equivalent rates in somatic tissues of adults. Nat Commun. 2013; 4:1597.

**32.** Magnusson PK, Almqvist C, Rahman I, Ganna A, Viktorin A, Walum H, Halldner L, Lundstrom S, Ullen F, Langstrom N, Larsson H, Nyman A, Gumpert CH, et al. The Swedish Twin Registry: establishment of a biobank and other recent developments. Twin research and human genetics : the official journal of the International Society for Twin Studies. 2013; 16:317-29.

**33.** Voight BF, Kang HM, Ding J, Palmer CD, Sidore C, Chines PS, Burtt NP, Fuchsberger C, Li Y, Erdmann J, Frayling TM, Heid IM, Jackson AU, et al. The metabochip, a custom genotyping array for genetic studies of metabolic, cardiovascular, and anthropometric traits. PLoS Genet. 2012; 8:1002793.

**34.** Cawthon RM. Telomere measurement by quantitative PCR. Nucleic Acids Res. 2002; 30:47.

**35.** Finkel D, Reynolds CA, McArdle JJ, Gatz M and Pedersen NL. Latent growth curve analyses of accelerating decline in cognitive abilities in late adulthood. Developmental psychology. 2003; 39:535-50.

**36.** Reynolds CA, Gatz M, Prince JA, Berg S and Pedersen NL. Serum lipid levels and cognitive change in late life. J Am Geriatr Soc. 2010; 58:501-09.

**37.** Singer & Willett. Applied Longitudinal Data Analysis: Modeling Change and Event Occurrence. 2003.

### SUPPLEMENTARY DATA

Leukocyte telomere length measurement
Batch effect adjustments of leukocyte telomere length.
Additional cross-sectional models tested
Latent Growth Curve (LGC) Analysis
References
Supplementary Tables
Supplementary Figures
Appendix

#### Leukocyte telomere length measurement

Leukocyte telomere length (LTL) was measured from DNA extracted from peripheral blood. We used a qPCR-based method [1], as described previously [2-4]. We used  $\beta$ -hemoglobin (Hgb) as a single copy reference gene. Separate reactions for telomere and Hgb reaction were carried out in paired 384-well plates in which matched sample well positions were used. Ten nanograms of DNA was used for each reaction, performed in triplicate. Every plate included a 7-point standard curve, which was used to create a standard curve and to perform absolute quantification of each sample. Samples and standard dilutions were transferred into the plates using a multichannel pipet and dried overnight at room temperature. Specific reaction mix for telomere reaction included 270 nM tel1b primer (5'-CGGTTT(GTTTGG)5GTT-3') and 900 nM tel2b primer (5'-GGCTTG(CCTTAC)5CCT-3'), 0.2X SYBR Green I (Invitrogen), 5 mM DTT (Sigma-Aldrich), 1% DMSO (Sigma-Aldrich), 0.2 mM of each dNTP (Fermentas), and 1.25 U AmpliTaq Gold DNA polymerase (Applied Biosystems) in a total volume of 15 µl AmpliTag Gold Buffer II supplemented with 1.5 mM MgCl<sub>2</sub>. Hgb reaction mix included 300 nM Hgb1 primer (5'-GCTTCTGACACAACTGTGTTCACTAGC-3') and Hgb2 primer (5'-CACCAACTTCATCCACGT TCACC-3') in a total volume of 15 µl of iQ SyBrGreen supermix (BioRad). The cycling conditions for telomere amplification were: 10 minutes at 95 °C followed by 25 cycles at 95 °C for 15 s and 54°C for 2 min with signal The cycling conditions for Hgb acquisition. amplification were: 95 °C for 10 min followed by 35 cycles at 95 °C for 15 s, 58 °C for 20 s, 72 °C for 20 s with signal acquisition. Reactions were performed with CFX384 Real-Time PCR Detection System (Bio-Rad). Melt-curve analysis was carried out in the end of the run to ensure specific primer binding.

We used the Bio-Rad CFX Manager software to perform quality control, and samples with standard deviation of >0.5 between triplicates were omitted from the analysis. Plate effect was taken into account by analyzing five genomic DNA control samples on every plate. We normalized the telomere and Hgb signal values separately to the mean of these control samples before taking the T/S ratio. A few samples failed in the qPCR assay (N=48,  $\sim$ 3%) and were thus excluded from further analysis.

# Batch effect adjustments of leukocyte telomere length

LTL measurements by qPCR techniques are sensitive to differences in the analyses protocols. In our samples, DNA extraction was done in one lab with one protocol, and the LTL assessments were then made in another lab using the protocol described above. Nevertheless, the T/S ratio reported may vary depending on which plate a given sample was run on, known as a batch effect. To explore a potential batch effect, linear regressions with LTL T/S-ratio as dependent variable and age, sex and dummy variables for the different plate IDs as independent variables revealed two plates with significant technical variation. A box plot divided by the different plate IDs were made to illustrate the batch effect (Supplementary Figure 4A). Thus, the T/S-ratio measurements were adjusted for plate number (two plates out of 22 had significant variation) by calculating residuals from the linear regression of T/S-ratio on plate ID (Supplementary Figure 4B), and residuals were then re-scaled back to T/S-ratio by adding the global mean to each value.

#### Additional cross-sectional models tested

In all additional models tested, LTL decreased with advancing age (Supplementary Table 2). Model 1, the crude model, and Model 3, including sex and GRS, showed effect estimates similar to the main model with sex as a covariate, presented in the main text. However, Model 2, which also included an interaction term between sex and age, did not fit well. Overall, sex contributes to the model but the GRS does not, nor are there significant interactions.

In sex-stratified models, LTL decreased with advancing age for both men and women (p-value  $\leq 0.02$  for both models). Men experienced a slightly accelerated decline in LTL; however, the differences in yearly telomere attrition rates between men and women were not significant (Supplementary Table 3).

#### Latent Growth Curve (LGC) Analysis

All LGC analyses were performed using PROC MIXED in SAS 9.4, and the SAS code is provided as an appendix at the end of the Supplementary Data. First, the centering age was selected to be 69.3 years because it was the median age at baseline (IPT3). (This was later tested using sensitivity analyses of the models with centering ages +/-5 years from 69.3 year as well as with exclusion of IPT3 measures.) Next, the data was coded so that all samples contributing to slope 1 (age < 69.3years) were marked and likewise for all samples contributing to slope 2 (age > 69.3 years). An intercept only model was run were all samples at baseline contributed, even the individuals with only one timepoint available contributed to this model. Then a simple linear slope was added to the model which improved model fit as described in the main text. Here all samples with more than one time point contributed to the slope. Finally a two-slope model was tested which had an even better fit to the data. To contribute to both slopes, an individual had to have samples from both before and after the centering age (69.3 years) and also at least three time points, many had up to five time points available. An individual with only two time points contributed to one of the slopes only. For each new model run, the improved model fit was confirmed using log-likelihood ratio test with comparisons to the prior model. All models included sex as covariate and adjustments for twinness by inclusion of random effects for within- and between twin pairs. When the genetic

for within- and between twin pairs. When the genetic risk score was added to the model it was further improved, but only for level of TL at age 69.3, which meant that the contribution was on the intercept only and not on change. Because of lack of samples with genetic markers was available, the genetic model was run and tested on a smaller set of individuals and comparisons had to be made to the same smaller set of individuals in the prior comparison model(s) as well.

#### RERERENCES

**1.** Cawthon RM. Telomere measurement by quantitative PCR. Nucleic Acids Res. 2002; 30:47.

**2.** Eerola J, Kananen L, Manninen K, Hellstrom O, Tienari PJ and Hovatta I. No evidence for shorter leukocyte telomere length in Parkinson's disease patients. J Gerontol A Biol Sci Med Sci. 2010; 65:1181-84.

**3.** Kananen L, Surakka I, Pirkola S, Suvisaari J, Lonnqvist J, Peltonen L, Ripatti S and Hovatta I. Childhood adversities are associated with shorter telomere length at adult age both in individuals with an anxiety disorder and controls. PLoS One. 2010; 5:10826.

**4.** Kao HT, Cawthon RM, Delisi LE, Bertisch HC, Ji F, Gordon D, Li P, Benedict MM, Greenberg WM and Porton B. Rapid telomere erosion in schizophrenia. Mol Psychiatry. 2008; 13:118-19.

#### SUPPLEMENTARY TABLES

SNP	Chr	Position	Gene	Effect allele	Other allele	Effect allele frequency	Imputed*
rs11125529	2	54329370	ACYP2	С	А	0.83	Yes
rs10936599	3	170974795	TERC	Т	С	0.25	Yes
rs7675998	4	164227270	NAF1	А	G	0.19	Yes
rs2736100	5	1339516	TERT	А	С	0.48	No
rs9420907	10	105666455	OBFC1	А	С	0.89	Yes
rs8105767	19	22007281	ZNF208	А	G	0.68	Yes
rs2281929	20	61892524	RTEL1	Т	С	0.89	No

#### Supplementary Table 1. Information on genetic variants of LTL used in SATSA

\*Imputed SNPs with proper\_info>0.4; LTL, leukocyte telomere length

# Supplementary Table 2. Crude and adjusted models including interactions with leukocyte telomere length as outcome variable for SATSA cross-sectional

	Model 1 <sup>§</sup>	Model 2 <sup>£</sup>	Model 3 <sup>†</sup>
	estimate (95% CI)	estimate (95% CI)	estimate (95% CI)
SATSA			
Age (years)	-0.0019 (-0.00320.0006)**	-0.0043 (-0.00920.0007)	-0.0022 (-0.00360.0009)**
Sex		-0.0357 (-0.2353-0.1639)	0.0488 (0.0170-0.0807)**
Age*Sex		0.0013 (-0.0016-0.0041)	
GRS			-0.0145 (-0.0942-0.0652)
Age*GRS			0.0001 (-0.0008-0.0010)
Sex*GRS			-0.0010 (-0.0260-0.0240)

\*\*p-value<0.01; <sup>§</sup>Model adjusted for twinness <sup>£</sup>Sex included as a covariate <sup>†</sup>Sex and GRS included as covariates. Reference group for sex is men. GRS, genetic risk score.

# Supplementary Table 3. Sex-stratified models of the association between leukocyte telomere length and age for SATSA cross-sectional

	Model 1-Men	Model 2-Women
	estimate (95% CI)	estimate (95% CI)
SATSA	N=264	N=372
Age	-0.0030 (-0.00540.0007)*	-0.0018 (-0.00330.0002)*

\*p-value<0.05; Models adjusted for twinness.

#### Supplementary Table 4. SATSA longitudinal characteristics by sex and IPT occasion

In-person testing (IPT)	IPT3	IPT5	IPT6	IPT8	IPT9
Year	1992-1994	1999-2001	2002-2004	2008-2010	2010-2012
Men	(N=162)	(N=149)	(N=126)	(N=122)	(N=102)
Telomere length (mean±sd) <sup>§</sup>	0.66±0.20	0.71±0.11	0.70±0.14	0.70±0.13	0.69±0.15
Age, yrs (mean±sd)	67.42±8.09	68.96±9.43	71.61±9.01	74.39±8.05	76.51±7.53
Women	(N=206)	(N=238)	(N=167)	(N=175)	(N=151)
Telomere length (mean±sd) <sup>\$</sup>	0.72±0.20	0.72±0.13	0.72±0.13	0.73±0.11	0.71±0.14
Age, yrs (mean±sd)	69.64±9.51	71.20±10.41	73.01±9.57	76.95±8.82	79.08±8.77

<sup>\$</sup>Mean telomere length is adjusted for batch effect and re-scaled back to T/S-ratio.

# Supplementary Table 5. Fixed and random effects of the 1 slope model (Model 2) from latent growth curve analysis of leukocyte telomere length and age

			<b>Fixed Effects</b>				
Intercept (estimate (95% CI)) Slope 1 (estimate (95% CI)) Sex (estimate (95% CI))							
0.7256 (	0.7094-0.7418)**	* -0.0	-0.0018 (-0.00280.0009)**			60.0173)**	
	Random Effects						
varI <sub>t</sub>	varIp	varS <sub>1t</sub>	varS <sub>1p</sub>	$Ri_ts_{1t}$	$Ri_{p}s_{1p}$	Residual	
0.0025**	0.0078***	0.000005	0.00002**	-1.0	-0.45**	0.012***	

\*\*p-value<0.01. \*\*\*p-value<0.001. Random effects: varl = variance around the intercept; varS<sub>1</sub> = variance around slope 1;  $Ri_{[]}s_1$  = correlation between the random intercepts and slope 1; residual = residual variance reflecting within-wave variance. In all models, sex as covariate as well as random effects between twin pairs (subscript 'p') and within twin pairs (subscript 't') were included

# Supplementary Table 6. Fixed and random effects of the full two-slope model (Model 3) from latent growth curve analysis of leukocyte telomere length and age including correlation and residual estimates.

		Fixe	d Effects				
Intercep (estimate (959		Slope 1 (estimate (95% CI)) (esti		Slope 2 (estimate (95% CI))		Sex (estimate (95% CI))	
0.7287	0.7287 -0.001			-0.0021		-0.0409	
(0.7095-0.747	79)*** (	-0.0033-0.001	0) (-0.0	(-0.00340.0008)**		70.0182)***	
		Rando	om Effects				
varI <sub>t</sub>	varIp	varS	lt Va	arS <sub>1p</sub>	varS <sub>2t</sub>	varS <sub>2p</sub>	
0.0060***	0.0082***	0.0000	6** 0.0	00001	0.00002*	0.00002	
$Ri_ts_{1t}$	$Ri_{p}s_{1p}$	$Ri_ts_{2t}$	$Ri_{p}s_{2p}$	$Rs_{1t}s_{2t}$	$Rs_{1p}s_{2p}$	Residual	
0.43*	-0.18	-1.00	-0.51**	1.00	-0.41	0.011***	

\*p-value<0.05; \*\*p-value<0.01; \*\*\*p-value<0.001; Random effects: varl = variance around the intercept; varS<sub>1</sub> = variance around slope 1; varS<sub>2</sub> = variance around slope 2; Ris = correlation between the random intercept and slope 1 or slope 2; Rs1s2 = correlation between slope 1 and slope 2; residual = residual variance reflecting within-wave variance. In all models, sex as covariate as well as random effects between twin pairs (subscript 'p') and within twin pairs (subscript 't') were included.

# Supplementary Table 7. Fixed and random effects of the full spline two-slope model (model 3) from latent growth curve analysis of leukocyte telomere length and age with GRS

Fixed Effects						
Intercept	S	Slope 1		e 2	Sex	GRS
(estimate (95%	CI)) (estimat	(estimate (95% CI))		95% CI))	(estimate (95% CI))	(estimate (95% CI))
0.7306	-0	.0010	-0.00	)23	-0.0383	-0.0108
(0.7106-0.7507)	)*** (-0.003	32-0.0013)	(-0.00370	).0010)**	(-0.06180.0148)**	(-0.01790.0038)**
Random Effects						
varI <sub>t</sub>	varIp	varS <sub>1</sub>	t	varS <sub>1p</sub>	varS <sub>2t</sub>	varS <sub>2p</sub>
0.0053**	0.0093***	0.0000	6*	0.00002	0.00002	0.00002
$Ri_ts_{1t}$	Ri <sub>p</sub> s <sub>1p</sub>	Ri <sub>t</sub> s <sub>2t</sub>	Ri <sub>p</sub> s <sub>2p</sub>	ŀ	$Rs_{1t}s_{2t}$ $Rs_{1p}s_{2p}$	, Residual
0.56**	-0.16	-1.00	-0.58**		0.64 -0.24	0.011***

\*p-value<0.05; \*\*p-value<0.01; \*\*\*p-value<0.001; GRS, Genetic Risk Score; Random effects: varI = variance around the intercept; varS1 = variance around slope 1; varS2 = variance around slope 2; Ris = correlation between the random intercept and slope 1 or slope 2; Rs1s2 = correlation between slope 1 and slope 2; residual = residual variance reflecting within-wave variance. In all models, sex as covariate as well as random effects between twin pairs (subscript 'p') and within twin pairs (subscript 't') were included.



**Supplementary Figure 1.** Genetic associations between every genetic variant alone and in combination in a genetic risk score (GRS) with leukocyte telomere length (LTL). Left panels are cross-sectional associations in the SATSA study (n=585) with adjustments for family structure, age and sex. Upper panel shows z-transformed LTL values. Lower panel shows LTL values adjusted for batch effect and rescaled back to T/S-ratio scale. Upper right panel presents genetic estimates for LTL from the Codd et al paper (Nature Genet 2013). Lower right panel presents genetic estimates from longitudinal LTL analysis using latent growth curve models.



**Supplementary Figure 2.** Plot of individual trajectories across measurement points longitudinally. The x-axis represents age at each in-person testing (IPT), and the y-axis represents the plate-adjusted residuals of telomere length reported by T/S-ratio. Left panel is men only and right is women only. Each line represents a participant's trajectory from raw data; individuals with only one measurement occasion are presented as single points. In addition, two linear regression lines are shown, before and one after the centering age (69.3 years).



**Supplementary Figure 3.** Individual relative leukocyte telomere length (LTL) elongation in the longitudinal cohort. The difference in telomere length measurement between two time points is on the x-axis. The frequency is on the y-axis. Telomere elongation is exhibited in samples with delta LTL >0, and telomere attrition is exhibited in samples with delta LTL<0. The four different panels represent the number of years that differs between the two time points of LTL measurements (0->5; 5->10; 10->15; 15->20 years).



**Supplementary Figure 4A.** Boxplots demonstrating the measures of telomere length by T/S-ratio over different plates. (A) Before adjusting for plate effect.



**Supplementary Figure 4B.** Boxplots demonstrating the measures of telomere length by T/S-ratio over different plates. (**B**) After adjusting for plate effects showing residuals for the association between telomere length and plate ID.

### Appendix

SAS code for LGC models used with random effects for within-, and between twinpairs.

```
/*Defining centering age and ages for slopes 1 and 2.*/
data allsubjs;
set allsubjs;
 cage=69.3; /*centering age*/
 iage=round((age-cage),.5); /*centering around cage*/
 Cage1=round((age-cage),.5); if age > 69.3 then Cage1=0; /*Slope 1*/
 Cage2=round((age-cage),.5); if age < 69.3 and age ne . then Cage2=0; /*Slope
2*/
run;
/*Intercept model.*/
title1 'Random Effects Model Fitting';
title2 'centering=69.13: selected=0';
title3 'Regular model: Intercept with sex';
proc mixed data=allsubjs noitprint covtest IC noclprint method=ML;
           class pairid twinid sex;
           model tlresidual=sex/solution ddfm=bw CL;
           random intercept/sub=twinid(pairid) type=unr;
           random intercept/SUB=pairid type=unr;
run;
/*One-slope model.*/
title3 'Regular model: L&S with sex';
proc mixed abs scoring=5 maxfunc=300 data=allsubjs itdetails covtest IC
noclprint method=ML;
          class pairid twinid sex;
           model tlresidual=iage sex/solution ddfm=bw outp=bpred CL;
           random intercept iage/SUB=twinid(pairid)type=unr;
           random intercept iage/SUB=pairid TYPE=unr;
run;
/*Two-slope model.*/
title3 'Two-Slope Model with sex';
proc mixed abs data=allsubjs scoring=300 maxfunc=300 itdetails covtest IC
noclprint method=ML;
           class pairid twinid sex;
           model tlresidual=Cage1 Cage2 sex/solution ddfm=bw outp=bpred CL;
           random intercept Cage1 Cage2/sub=twinid(pairid) type=unr;
           random intercept Cage1 Cage2/sub=pairid TYPE=unr;
run;
```