Research Paper

Accelerated epigenetic aging as a risk factor for chronic obstructive pulmonary disease and decreased lung function in two prospective cohort studies

Miyuki Breen¹, Jamaji C. Nwanaji-Enwerem^{2,3,4}, Stefan Karrasch^{5,6,7}, Claudia Flexeder⁶, Holger Schulz^{6,7}, Melanie Waldenberger⁸, Sonja Kunze⁸, Markus Ollert^{9,10}, Stefan Weidinger¹¹, Elena Colicino¹², Xu Gao¹³, Cuicui Wang², Jincheng Shen¹⁴, Allan C. Just¹², Pantel Vokonas¹⁵, David Sparrow¹⁵, Lifang Hou¹⁶, Joel D. Schwartz², Andrea A. Baccarelli¹³, Annette Peters⁶, Cavin K. Ward-Caviness¹⁷

¹Oak Ridge Institute for Science and Education (ORISE), Center for Public Health and Environmental Assessment, US Environmental Protection Agency, Chapel Hill, NC 27709, USA

²Department of Environmental Health, Harvard T.H. Chan School of Public Health, Boston, MA 02115, USA

³ MD-PhD Program, Harvard Medical School, Boston, MA 02115, USA

⁴Belfer Center for Science and International Affairs, Harvard Kennedy School of Government, Cambridge, MA 02138, USA

⁵Institute and Outpatient Clinic for Occupational, Social and Environmental Medicine, Inner Clinic, University Hospital of Munich, Ludwig-Maximilians-Universität, Munich, Germany

⁶Institute of Epidemiology, Helmholtz Zentrum München, Neuherberg, Germany

⁷Comprehensive Pneumology Center Munich (CPC-M), Member of the German Center for Lung Research (DZL), Munich, Neuherberg, Germany

⁸Research Unit Molecular Epidemiology, Helmholtz Zentrum München, Neuherberg, Germany

⁹Department of Infection and Immunity, Luxembourg Institute of Health, Esch-sur-Alzette, Luxembourg

¹⁰Odense Research Center for Anaphylaxis, Department of Dermatology and Allergy Center, University of Southern Denmark, Odense, Denmark

¹¹Department of Genetic Dermatology, University of Kiel, Kiel, Germany

¹²Department of Environmental Medicine and Public Health, Icahn School of Medicine at Mount Sinai, New York, NY 10029, USA

¹³Department of Environmental Health Sciences, Mailman School of Public Health, Columbia University, New York, NY 10032, USA

¹⁴Department of Population Health Sciences, University of Utah, School of Medicine, Salt Lake City, UT 84132, USA
 ¹⁵Veterans Affairs Normative Aging Study, Veterans Affairs Boston Healthcare System, Department of Medicine, Boston University School of Medicine, Boston, MA 02118, USA

¹⁶Department of Preventive Medicine, Feinberg School of Medicine, Northwestern University, Chicago, IL 60611, USA
 ¹⁷Center for Public Health and Environmental Assessment, US Environmental Protection Agency, Chapel Hill, NC
 27709, USA

Correspondence to: Cavin K. Ward-Caviness; email: ward-caviness.cavin@epa.govKeywords: DNA methylation age, COPD, lung function, accelerated agingReceived: April 15, 2020Accepted: July 14, 2020Published: August 3, 2020

Copyright: Breen et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Chronic obstructive pulmonary disease (COPD) is a frequent diagnosis in older individuals and contributor to global morbidity and mortality. Given the link between lung disease and aging, we need to understand how molecular indicators of aging relate to lung function and disease. Using data from the population-based KORA

(Cooperative Health Research in the Region of Augsburg) surveys, we associated baseline epigenetic (DNA methylation) age acceleration with incident COPD and lung function. Models were adjusted for age, sex, smoking, height, weight, and baseline lung disease as appropriate. Associations were replicated in the Normative Aging Study. Of 770 KORA participants, 131 developed incident COPD over 7 years. Baseline accelerated epigenetic aging was significantly associated with incident COPD. The change in age acceleration (follow-up – baseline) was more strongly associated with COPD than baseline aging alone. The association between the change in age acceleration between baseline and follow-up and incident COPD replicated in the Normative Aging Study. Associations with spirometric lung function parameters were weaker than those with COPD, but a meta-analysis of both cohorts provide suggestive evidence of associations. Accelerated epigenetic aging, both baseline measures and changes over time, may be a risk factor for COPD and reduced lung function.

INTRODUCTION

Lung disease continues to be a primary determinant of morbidity and mortality worldwide. A study on the global burden of disease indicates that although the contribution of asthma to global mortality decreased by 11.7% between 2005 and 2015, asthma still contributed to > 397,000 deaths in 2015. Chronic obstructive pulmonary disease (COPD) is a substantially greater contributor to global mortality with 3,188,300 deaths linked to COPD in 2015, and, unlike asthma, COPDattributable deaths appear to be increasing [1]. A globally aging population is likely a primary contributor to the increased mortality burden of COPD, a trend which may exacerbate as the population of older adults (particularly in developed nations) rapidly expands.

Though lung function and lung disease are closely linked with aging [2], relatively little is known about how molecular indicators of biological age relate to lung disease and lung function. One of the most widely used biomarkers of biological age is telomere length [3]. Telomeres are repetitive sequences that cap the ends of chromosomes and prevent the loss of genetic information during DNA replication. Decreased telomere length is an indicator of increased biological age, and is associated with pulmonary fibrosis and emphysema, suggesting a link between molecular biomarkers of aging and lung disease [4-7]. Recently, DNA methylation has emerged as a novel biomarker for the aging process [8]. DNA methylation-based aging biomarkers are associated with mortality, cancer, hemostasis, frailty, and other ageassociated traits and diseases [9]. However, relatively little is known about how lung disease and function relate to DNA methylation-derived aging biomarkers. In a study of two adult cohorts, DNA methylation loci associated with changes in lung function were correlated with DNA methylation age and age acceleration difference (difference between DNA methylation age and chronological age) [10]. In a study of adult women, accelerated DNA methylation aging was associated with incident lung cancer [11] and there have been two smaller studies examining asthma in children [12] and lung function in adults aged 70+ [13] showing positive associations. However, there has still been no large, prospective cohort study of lung function and lung disease risks using epigenetic aging biomarkers. Here we use the Horvath DNA methylation age (DNAmAge) biomarker [14] to determine if accelerated epigenetic aging is related to lung function measurements and incident COPD in two prospective, population-based cohorts.

RESULTS

The characteristics of KORA are shown in Table 1. The mean age at baseline was 47.4 yr and there was an average of 7.09 yr between baseline and follow-up examinations. We observed 131 cases of incident COPD in KORA. The mean age acceleration difference (AAD) at baseline was 1.59 yr while the mean extrinsic epigenetic age acceleration difference (EEAD) at baseline was 2.90 yr. At follow-up, the mean lung function measures were 4.25 liters for FVC, 3.30 liters for FEV₁, 2.93 liters for FEF25-75 and 0.78 for FEV₁/FVC. The characteristics of the replication NAS cohort are shown in Supplementary Table 1. We define AAD as the difference between Horvath epigenetic age and chronological age. EEAD is defined as the difference between extrinsic epigenetic age and chronological age.

Adjustment models for the analyses are given in Table 2 and described in the methods. Higher baseline EEAD was associated with higher odds of incident of COPD (odds ratio [OR] = 1.01 per 5-year higher EEAD; 95% confidence interval = 1.00 - 1.03) while AAD did not show an association with incident COPD (Table 3; Supplementary Table 2). There was a modest, negative correlation between the difference in AAD between baseline and follow-up (Δ AAD) and AAD as well as between the difference in EEAD between baseline and follow-up (Δ EEAD) and EEAD at baseline indicating that those with lower baseline AAD and EEAD had larger changes in these aging biomarkers over time (Supplementary Figure 1). Analyses of $\triangle AAD$ and $\triangle EEAD$ in KORA revealed stronger associations than baseline measures alone, and the overall pattern of associations for both $\triangle AAD$ and $\triangle EEAD$ was

concordant with associations observed for AAD and EEAD, i.e. stronger associations for Δ EEAD than Δ AAD and a slight attenuation of the estimates with inclusion of additional confounders. Δ EEAD was

N 770*	Baseline	Follow-up
<i>N</i> = 770 *	Mean (SD)	Mean (SD)
Age (yr)	47.4 (5.57)	54.5 (5.60)
Follow-up time (yr)	-	7.09 (0.42)
Weight (kg)	77.8 (15.7)	-
Height (cm)	169 (9.13)	-
DNAmAge (yr)	50.0 (6.62)	54.9 (5.74)
AAD (yr)	2.59 (4.64)	0.36 (4.43)
EEAD (yr)	1.90 (5.65)	8.87 (5.78)
FVC (liter)	-	4.25 (0.97)
FEV ₁ (liter)	-	3.30 (0.77)
FEF25-75 (liter/s)	-	2.93 (1.01)
FEV ₁ /FVC	-	0.78 (0.06)
Binary Variables	N (%)	N (%)
Female	406 (52.7)	406 (52.7)
Incident COPD	-	131 (18.0)
Asthma	52 (6.92)	81 (10.5)
Never Smoker	295 (38.4)	294 (38.2)
Former Smoker	279 (36.3)	328 (42.4)
Current Smoker	195 (25.4)	148 (19.2)

AAD = age acceleration difference; BMI = body mass index; COPD = Chronic obstructive pulmonary disease; DNAmAge = DNA methylation age; EEAA = extrinsic epigenetic age acceleration; EEAD = extrinsic epigenetic age acceleration difference; FEV₁ = forced expiratory volume in 1 s; FVC = forced vital capacity; FEF25-75 = forced expiratory flow at 25-75% of FVC; GOLD = Global Initiative for Chronic Obstructive Lung Disease; GLI = Global Lungs Initiative. * = only 726 participants had values available for lung function outcomes.

Table 2. Model descriptions.

Outcome	Exposure (Epigenetic Aging Measure)			
Outcome	AAD, EEAD	ΔΑΑΦ, ΔΕΕΑΦ		
Lung Function (FEV ₁ , FVC,	age, sex, smoking status, height,	age, sex, smoking status, height, weight, baseline COPD,		
FEV ₁ /FVC, FEF25-75)	weight, baseline COPD, baseline	baseline asthma, years between examinations, baseline		
	asthma	epigenetic age (AAD, EEAD)		
COPD	age, sex, smoking status, height,	age, sex, smoking status, height, weight, years between		
	weight	examinations, baseline epigenetic age (AAD, EEAD)		

List of the covariates adjusted for in each combination of outcomes and epigenetic aging measures (exposures) examined. Age, sex, smoking status, height, and weight were included in all models. Models for lung function variables additionally included baseline COPD and asthma. Models for the change in epigenetic age over time (ΔAAD , $\Delta EEAD$) additionally adjusted for baseline epigenetic age (AAD or EEAD for ΔAAD and $\Delta EEAD$ models respectively) as well as the follow up time (years between examinations. AAD = age acceleration difference; COPD = chronic obstructive pulmonary disease; EEAD = extrinsic epigenetic age acceleration difference; FEF27-75 = forced expiratory flow at 25-75% of pulmonary volume; FEV₁ = forced expiratory volume in 1 second; FVC = forced vital capacity.

Epigenetic aging measure	OR	LCI	UCI	Р
AAD	1.00	0.99	1.02	0.63
EEAD	1.01	1.00	1.03	0.04
ΔΑΑD	1.01	0.98	1.03	0.62
ΔEEAD	1.02	1.00	1.04	0.04

Table 3. Associations between epigenetic aging measures and COPD.

Adjustments for the epigenetic aging models are shown in Table 2. Associations are presented per 5-year change in the epigenetic aging measure. AAD = Age Acceleration Difference; EEAD = Extrinsic Epigenetic Age Acceleration Difference; LCI = lower 95% confidence interval; OR = odds ratio; P = p-value; UCI = upper 95% confidence interval; ΔAAD = change in AAD between baseline and follow-up; $\Delta EEAD$ = change in EEAD between baseline and follow-up.

associated with incident COPD (Table 3). Of the COPD associations with P < 0.05 in KORA, the Δ EEAD and COPD association replicated in NAS (Supplementary Table 3) with a nearly identical effect estimate and 95% confidence interval across the two studies and a smaller 95% confidence interval in the meta-analysis (OR = 1.02; 95% CI = 1.01-1.03; P = 7.1x10⁻⁴; Figure 1).

In KORA, we observed little to no evidence for associations between lung function measures (FVC, FEV₁, FEF25-75, and FEV₁/FVC) measured at baseline and epigenetic aging biomarkers measured at follow up (Supplementary Table 2). Unlike with COPD, lung function associations did not improve substantially when examining $\triangle AAD$ or $\triangle EEAD$ and many of the associations weakened (Supplementary Table 2). In NAS we also observed weak evidence for associations between lung function and aging measures, with the exception of FEV₁/FVC which was associated with both Δ EEAD and Δ AAD (Supplementary Table 3). Though weak to null in KORA and NAS individually, associations between epigenetic aging and lung function were often consistent between the two cohorts, and in a meta-analysis, associations with FEV₁ and FEV₁/FVC and epigenetic aging measures were observed (Supplementary Table 4).

DISCUSSION

We observed significant associations between epigenetic aging and COPD in analyses of independent, prospective cohorts. In KORA, a 5 yr difference in EEAD at baseline was associated with an increased risk of incident COPD, a though the magnitude of the associations was smaller than associations seen with other traits [9]. The change in EEAD between the baseline and follow-up examinations (Δ EEAD) was more strongly associated with COPD at follow-up than baseline EEAD alone. We observed an inverse correlation between baseline epigenetic aging and the change in epigenetic age over time, which may represent regression towards the mean. The association between Δ EEAD and COPD replicated in NAS, an older, all male cohort from the USA, and was has similar magnitude with a smaller 95% confidence interval in the meta-analysis than in either cohort alone. As the aging rate has been stated to be constant across adulthood [15], it is possible that Δ EEAD measured between baseline and follow-up is largely reflective of the aging rate at baseline in these adult populations. Future studies should examine baseline epigenetic aging rates to determine if they are risk factors for COPD, independent of baseline epigenetic age itself.

Associations for lung function in KORA were substantially weaker than associations for incident COPD and were often consistent with a null association. Only in a meta-analysis of KORA and NAS were associations between epigenetic aging and lung function observed for EEAD and FEV₁ (-0.96% change in FEV₁ from median per 5 year higher EEAD, 95% confidence interval = -1.80 - -0.11; Supplementary Table 4). Thus, while there is some evidence for an association between epigenetic aging and lung function, these associations appear weaker than those observed for COPD and may be best explored in meta-analyses or significantly larger cohorts.

Accelerated epigenetic aging has been associated with mortality and several health outcomes, and while the drivers of accelerated epigenetic aging have not been conclusively determined, a number of environmental exposures are associated with epigenetic aging [9]. EEAD was used here to measure accelerated epigenetic aging and calculated as the difference between extrinsic epigenetic age and chronological age. Unlike AAD, which is tissue agnostic, EEAD is valid only in blood and strongly correlated with blood immune cell counts and is considered a biomarker for epigenetic aging related to the immune system. COPD and lung function are associated with local or systemic inflammation and alterations regarding the immune system [16–18], which may account for why EEAD uniformly showed stronger associations with COPD and lung function than AAD.

There have been two other studies to date to examine lung outcomes and epigenetic aging. In the first study repeated measures of epigenetic age and a variety of physical fitness related measures, including FEV_1 , at ages 70, 73, and 76 in the Lothian Birth Cohort (a cohort recruited out of the Lothian area of Scotland in 1936). In

this study, accelerated epigenetic age, equivalent to the AAD measure here, was associated with decreased FEV₁ [13]. Extrinsic epigenetic age was not developed until after this study and thus was not tested. The second study examined a cohort of children as part of Project Viva, a mother-child pair cohort recruited from Boston, Massachusetts. In this cohort extrinsic epigenetic age acceleration measured at mid-childhood (mean age = 7.8vr) was associated with asthma in cross-sectional associations, however extrinsic epigenetic age acceleration at birth was not associated with asthma at mid-childhood [12].



Figure 1. Associations between epigenetic aging measures and COPD in KORA and NAS. Associations were performed independently in KORA and NAS using the adjustment models laid out in Table 2 (including adjustment for baseline epigenetic age in the ΔAAD and $\Delta EEAD$ models) and are presented per 5-year change in the epigenetic aging measure. A fixed effect meta-analysis was used to combine results across cohorts. Only $\Delta EEAD$ is considered to have replicated as it had association with P < 0.05 in the discovery cohort (KORA) and in the replication cohort (NAS). For all epigenetic aging measures we consistently observe a 1-2% increase in the odds of COPD per 5-year change (elevation) in epigenetic aging. AAD = age acceleration difference; ΔAAD = change in age acceleration difference; EEAD = extrinsic epigenetic age acceleration difference; FE = Fixed effect.

Our study has several advantages over these previous analyses. It is the first study to examine both COPD and lung function for multiple epigenetic aging measures in the same cohort. This enabled a direct comparison of the relationships revealing that associations for COPD were stronger than those for lung function measures in this cohort. Another strength of our study is the sample size. With more than 700 participants in KORA and more than 400 in NAS, our study is larger than both previous studies which had samples sizes < 1000. Additionally, KORA had a younger age distribution (mean age at baseline = 47.4vr) as compared to either previous study, and thus may be more generalizable to an average adult population. Our study also benefits from replication across independent cohorts sampled from different countries and with different demographic distributions, e.g. age and sex. KORA and NAS also differed in the mean change in epigenetic age seen between baseline and follow-up (though all odds ratios are expressed per 5year higher epigenetic age), another potential source of heterogeneity. Despite these differences, associations were often concordant between KORA and NAS. highlighting the consistency of the observed associations. A limitation of our study and the previous studies is the limited ethnic diversity. Both KORA and the Lothian birth cohort are single ethnicity cohorts (German and Scottish respectively) and while Project Viva is a multi-ethnic cohort, the majority of the participants are of Caucasian ancestry, as is also the case with NAS. This limits the ability of current studies to verify that associations generalize to multiple ethnicities. The repeated DNA methylation samplings in KORA allowed us to compute the change in DNA methylation over time and associate that with lung function and disease at follow-up. The Lothian Birth Cohort also had longitudinal DNA methylation samplings but was only able to examine longitudinal associations with FEV₁, and did not compare baseline associations with changes in epigenetic aging over time. Our observation that the change in epigenetic age over time may be more strongly associated with COPD than baseline epigenetic age acceleration provides useful information on alterations in risk as epigenetic age acceleration changes.

Potential mechanisms

Though we are still at the beginning of understanding the impacts of epigenetic aging, it is important to consider the potential mechanisms that might underlie associations between epigenetic aging biomarkers and disease to put these associations in a more complete context. There are several potential mechanisms linking epigenetic age with both COPD and lung function. Cellular senescence has been previously linked to the

pathogenesis of lung disease. Telomere length (a molecular marker of aging and cellular senescence) is linked to emphysema, COPD, and pulmonary fibrosis [4, 5, 7]. Epigenetic aging presents a further potential biomarker of cellular aging that linked to risk of lung disease and possibly declines in lung function. It is also possible that epigenetic aging is a mediator of environmental exposures linked to lung disease and function. Elevated air pollution exposure is associated with higher DNA methylation age [9, 19, 20], and there have been a few studies suggesting that DNA methylation alterations may mediate the effects of air pollution exposures on lung function [21–23]. Thus, it is possible that one of the links between epigenetic aging and lung function / disease lies in epigenetic mediation of environmental exposures like air pollution.

CONCLUSIONS

We used adult participants from two independent, longitudinal cohort studies to uncover prospective associations between COPD and lung function and epigenetic aging. The results expand on the literature suggesting that accelerated epigenetic aging is a risk factor for chronic, age-related diseases, even after controlling for chronological age. The results supplement work which establish epigenetic mechanisms as an important part of the pathophysiologic pathways underlying lung function decrements and incident COPD. Further work is needed to functionally validate the results, establish if modifications of epigenetic aging biomarkers can alter future disease risk, and expand associations to other ethnic groups.

MATERIALS AND METHODS

Study population

The discovery data for this study were taken from the fourth baseline survey of the Cooperation for Health Research in the Region of Augsburg (KORA S4) and their follow-up (KORA F4). These data consist of crosssectional population-based surveys from registered German residents in Augsburg and its two adjacent counties (Southern Germany) [24]. The KORA S4 study was conducted between 1999 and 2001 with 4261 participants with ages from 25 to 74 yr. Of these participants, 3080 also participated in the 7 yr follow-up KORA F4 study between 2006 and 2008. Information on lifestyle and medical history questionnaire were gathered by trained medical staff during a standardized interview. as well as an extensive standardized medical examination including the collection of blood samples for later clinical chemistry and genomic analyses. The details of this cohort have been previously published [25-27]. The Bavarian Medical Association Ethics

Committee approved these studies and all participants provided written informed consent.

The Normative Aging Study (NAS) was used to replicate associations from the KORA cohort. The NAS was established by the U.S. Department of Veterans Affairs and is an ongoing longitudinal cohort study of male volunteers within the Greater Boston area. In 1963, the NAS began enrolling participants free of any chronic medical conditions. The NAS is now a closed cohort, but study participants continually return for onsite, detailed medical examinations every 3-5 years. During these study visits, anthropomorphic measurements are performed, biological specimens are collected, and data on smoking status and additional risk factors that may impact health are documented. Participants provided written informed consent to the VA Institutional Review Board (IRB). The Harvard T.H. Chan School of Public Health and the VA IRBs granted human subjects' approval. Eligibility for our study sample required complete data for the exposures, outcomes, and covariates determined a priori for the present analysis. Given the analysis structure, for inclusion, participants were also required to have at least two study visits.

Epigenetic aging measures

DNA methylation was assessed in KORA S4 and F4 and NAS using the Illumina Infinium HumanMethylation 450k array, and DNAmAge was calculated using the online calculator provided by the laboratory of Dr. Steve Horvath [14]. Ouality control for the methylation data and use of the online calculator for KORA was previously described in detail by Ward-Caviness CK et al. [19]. DNA methylation was assessed similarly in NAS as it was in KORA [28]. Briefly for both cohorts study staff measured DNA methylation on whole blood DNA collected at each study visit for the participants. After performing bisulfite conversion on the DNA (EZ-96 DNA Methylation Kit, Zymo Research, Orange, CA, USA), DNA methylation was quantified using the Illumina HumanMethylation450k platform (Infinium HD Methylation protocol, Illumina, San Diego, CA, USA). In NAS, to ensure a similar age distribution across chips/plates and minimize batch effects, a twostage age-stratified algorithm was used to randomize samples across plates. Samples where >5% of probes had a beadcount < 3 or > 1% of probes had a detection P-value >0.05 were removed. The remaining samples were pre-processed with Illumina-type background correction without normalization and normalized with dye-bias and beta-mixture quantile normalization adjustments. In KORA samples with detection P-values > 0.05 for 1% of probes and probes with detection Pvalues > 0.05 for 1% of samples were removed. Methylation data was normalized using quantile

normalization and beta-mixture quantile normalization. For both cohorts methylation beta values were calculated as the ratio of the intensity of the methylated signal over the total (methylated and unmethylated) probe intensity signal with a constant of 100 added to the denominator to prevent large values with low intensity probes, per standard procedures.

For both KORA and NAS the methylation beta values were used to estimate epigenetic aging measures. For our analysis, we focused on two epigenetic measures: Age Acceleration Difference (AAD) and Extrinsic Epigenetic Age Acceleration Difference (EEAD). AAD assesses differences between DNAmAge and chronological age and is tissue agnostic [14]. EEAD is a blood-specific measures of aging that is the difference between chronological age and Extrinsic Epigenetic Age which correlates with immune cell counts [11]. Both epigenetic aging measures assess "aging" as the difference between the epigenetic age measure (DNAmAge and Extrinsic Epigenetic Age for AAD and EEAD respectively) and chronological age.

COPD and lung function

Incident COPD at follow up (KORA F4) was the primary outcomes with secondary analysis of lung function measures. COPD (including chronic bronchitis) was assessed via a questionnaire provided at baseline to the KORA participants, and all participants reporting COPD were removed from the incident COPD analyses. At follow-up a more robust assessment of COPD was possible thanks to the availability of spirometric indices. The spirometric indices available at follow up were forced expiratory volume in 1 s (FEV₁), forced vital capacity (FVC), FEV₁/FVC, and forced expiratory flow between 25-75% of FVC (FEF25-75). In KORA spirometry was performed in accordance with standard recommendations [29]. Briefly, spirometry was performed on sitting subjects wearing noseclips using a pneumotachograph-type spirometer (Masterscope PC; CareFusion, Höchberg, Germany) with a resistance of 0.05 kPa·l-1·s-1 at 10 l·s-1 and a volume accuracy of ± 5 ml [30]. Spirometry was performed according to international standards [29, 31] and according to American Thoracic Society guidelines [32]. In NAS, spirometry was measured in standing participants with a nose clip using a 10-litre water-filled survey-recording spirometer and an Eagle II minicomputer (Warren E. Collins, Braintree, Massachusetts). Asthma and smoking status (current, former, and never) were assessed via self-report or the via the American Thoracic Society (ATS) questionnaire [33].

To identify all potential cases, incident COPD was assessed as a positive indication via spirometry, i.e. the definitions based on Global Initiative for Chronic Obstructive Lung Disease (GOLD) or the Global Lungs Initiative (GLI) methods, as well as self-report via questionnaire for chronic bronchitis. The GOLD and GLI methods are defined as $FEV_1/FVC < 0.70$ [34] and FEV_1/FVC z-score < -1.6445 (5% lower limit of normal) [35], respectively.

In NAS only lung function measures were available thus we identified incident COPD cases as a positive via either the GOLD or GLI methods.

Covariates

Demographic, anthropometric, and lifestyle data were obtained during cohort enrollment and at follow-up. Smoking status was categorized as smokers (including occasional smokers), ex-smokers and never smokers via self-report. In KORA S4, physical examinations were conducted to obtain height and weight, and body mass index (BMI) was calculated as weight (kg) divided by height (m) squared. Similarly, in NAS demographic, anthropomorphic, and lifestyle data were collected at baseline and at follow up including smoking status, age, sex, height, and weight. Baseline asthma and COPD status were collected via self-report in KORA. COPD was assessed in NAS using the GOLD and GLI spirometry based methods, as there baseline questionnaire assessment of COPD was not available in NAS.

Statistical analysis

We conducted all statistical analyses using the R statistical language (R Core Development Team, version 3.4.3) [36]. Logistic and linear regression models were implemented for binary and continuous outcome variables, respectively. We analyzed each epigenetic aging measure separately for associations with COPD and lung function and used a nominal significance threshold of P < 0.05 to determine significance given the limited number of outcomes examined and correlations between outcome measures.

We performed two statistical analyses. First, a prospective analysis was conducted using baseline (KORA S4) epigenetic aging measures and COPD as well as lung function assessed at follow-up (KORA F4). Second, we assessed if the change (Δ) in AAD and EEAD between baseline and follow-up was associated with COPD or lung function assessed at follow-up. This analysis included an adjustment for baseline epigenetic aging. For the analyses we adjusted for age, sex, smoking status, height, and weight. We additionally considered inclusion of body mass index (height [m]/weight [kg]²) however this did not modify

associations after including height and weight, so we decided to use the more parsimonious model.

For the secondary analyses of lung function, we allowed participants with COPD at baseline to participate in the analyses to maximize sample size. To prevent confounding by baseline COPD status we added an adjustment for both baseline COPD as well as baseline asthma (assessed via self-report) to the models. For models with ΔAAD and $\Delta EEAD$ as predictors, we added the follow-up time to all models to observe if adjusting for variations in follow-up times altered associations, as well as adjustment for baseline epigenetic age. Table 2 details the models used for each analysis.

Associations with P < 0.05 in KORA were replicated in the Normative Aging Study (NAS). We chose a nominal p-value as the replication threshold because lung function and COPD are highly correlated (as COPD was in part defined by lung function). Thus, the tests were not independent and a Bonferroni multiple testing correction would be overly conservative. We additionally used the metafor package [37] in R to perform fixed-effects meta-analyses for associations across the two cohorts. Though we examined association in NAS and under the meta-analysis of NAS and KORA, formal replication was only considered for those associations with P < 0.05 in the discovery cohort (KORA) and with the same direction of association and P < 0.05 in the replication cohort (NAS) per accepted standards.

ACKNOWLEDGMENTS

The research described in this article has been reviewed by the Center for Public Health and Environmental Assessment, U.S. Environmental Protection Agency, and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the Agency, nor does the mention of trade names of commercial products constitute endorsement or recommendation for use. All opinions expressed in this paper are the author's and do not necessarily reflect the policies and views of EPA, DOE, or ORAU/ORISE.

CONFLICTS OF INTEREST

The authors declare no conflicts of interest

FUNDING

Funding for this study was provided in part by the U.S. Environmental Protection Agency (EPA). Miyuki Breen was supported by an appointment to the EPA Research Participation Program administered by the Oak Ridge

Institute for Science and Education (ORISE). The KORA study was initiated and financed by the Helmholtz Zentrum München - German Research Center for Environmental Health, which is funded by the German Federal Ministry of Education and Research (BMBF) and by the State of Bavaria. Furthermore, KORA research was supported within the Munich Center of Health Sciences (MC-Health), Ludwig-Maximilians-Universität, as part of LMUinnovativ. The Normative Aging Study has been supported by the National Institute of Environmental Health Sciences (grants P30ES009089, R01ES021733, R01ES025225, and R01ES027747) and by the Cooperative Studies Program/Epidemiology Research and Information Center of the U.S. Department of Veterans Affairs and is a component of the Massachusetts Veterans Epidemiology Research and Information Center in Boston, Massachusetts.

REFERENCES

- Wang M, Utell MJ, Schneider A, Zareba W, Frampton MW, Oakes D, Hopke PK, Wiltshire J, Kane C, Peters A, Breitner S, Chalupa D, Rich DQ. Does total antioxidant capacity modify adverse cardiac responses associated with ambient ultrafine, accumulation mode, and fine particles in patients undergoing cardiac rehabilitation? Environ Res. 2016; 149:15–22. <u>https://doi.org/10.1016/j.envres.2016.04.031</u> PMID:<u>27174779</u>
- Thannickal VJ, Murthy M, Balch WE, Chandel NS, Meiners S, Eickelberg O, Selman M, Pardo A, White ES, Levy BD, Busse PJ, Tuder RM, Antony VB, et al. Blue journal conference. Aging and susceptibility to lung disease. Am J Respir Crit Care Med. 2015; 191:261–69.

https://doi.org/10.1164/rccm.201410-1876PP PMID:25590812

- Aubert G, Lansdorp PM. Telomeres and aging. Physiol Rev. 2008; 88:557–79. <u>https://doi.org/10.1152/physrev.00026.2007</u> PMID:<u>18391173</u>
- Alder JK, Guo N, Kembou F, Parry EM, Anderson CJ, Gorgy AI, Walsh MF, Sussan T, Biswal S, Mitzner W, Tuder RM, Armanios M. Telomere length is a determinant of emphysema susceptibility. Am J Respir Crit Care Med. 2011; 184:904–12. <u>https://doi.org/10.1164/rccm.201103-05200C</u> PMID:<u>21757622</u>
- Nunes H, Monnet I, Kannengiesser C, Uzunhan Y, Valeyre D, Kambouchner M, Naccache JM. Is telomeropathy the explanation for combined pulmonary fibrosis and emphysema syndrome?: report of a family with TERT mutation. Am J Respir Crit Care Med. 2014; 189:753–54.

https://doi.org/10.1164/rccm.201309-1724LE PMID:24628319

- Yoon YS, Jin M, Sin DD. Accelerated lung aging and chronic obstructive pulmonary disease. Expert Rev Respir Med. 2019; 13:369–380. <u>https://doi.org/10.1080/17476348.2019.1580576</u> PMID:<u>30735057</u>
- Albrecht E, Sillanpää E, Karrasch S, Alves AC, Codd V, Hovatta I, Buxton JL, Nelson CP, Broer L, Hägg S, Mangino M, Willemsen G, Surakka I, et al. Telomere length in circulating leukocytes is associated with lung function and disease. Eur Respir J. 2014; 43:983–92. <u>https://doi.org/10.1183/09031936.00046213</u> PMID:24311771
- Horvath S, Raj K. DNA methylation-based biomarkers and the epigenetic clock theory of ageing. Nat Rev Genet. 2018; 19:371–84. <u>https://doi.org/10.1038/s41576-018-0004-3</u> PMID:<u>29643443</u>
- Dhingra R, Nwanaji-Enwerem JC, Samet M, Ward-Caviness CK. DNA methylation age-environmental influences, health impacts, and its role in environmental epidemiology. Curr Environ Health Rep. 2018; 5:317–27. https://doi.org/10.1007/s40572-018-0203-2
- PMID:<u>30047075</u>
 10. Carmona JJ, Barfield RT, Panni T, Nwanaji-Enwerem JC, Just AC, Hutchinson JN, Colicino E, Karrasch S, Wahl S, Kunze S, Jafari N, Zheng Y, Hou L, et al. Metastable DNA methylation sites associated with longitudinal lung function decline and aging in humans: an epigenome-wide study in the NAS and KORA cohorts.

Epigenetics. 2018; 13:1039–55. https://doi.org/10.1080/15592294.2018.1529849 PMID:30343628

- Levine ME, Hosgood HD, Chen B, Absher D, Assimes T, Horvath S. DNA methylation age of blood predicts future onset of lung cancer in the women's health initiative. Aging (Albany NY). 2015; 7:690–700. <u>https://doi.org/10.18632/aging.100809</u> PMID:26411804
- Peng C, Cardenas A, Rifas-Shiman SL, Hivert MF, Gold DR, Platts-Mills TA, Lin X, Oken E, Avila L, Celedón JC, Weiss ST, Baccarelli AA, Litonjua AA, DeMeo DL. Epigenetic age acceleration is associated with allergy and asthma in children in project viva. J Allergy Clin Immunol. 2019; 143:2263–70.e14. <u>https://doi.org/10.1016/j.jaci.2019.01.034</u> PMID:30738172
- 13. Marioni RE, Shah S, McRae AF, Ritchie SJ, Muniz-Terrera G, Harris SE, Gibson J, Redmond P, Cox SR, Pattie A, Corley J, Taylor A, Murphy L, et al. The

epigenetic clock is correlated with physical and cognitive fitness in the lothian birth cohort 1936. Int J Epidemiol. 2015; 44:1388–96. https://doi.org/10.1093/ije/dyu277 PMID:25617346

- Horvath S. DNA methylation age of human tissues and cell types. Genome Biol. 2013; 14:R115. <u>https://doi.org/10.1186/gb-2013-14-10-r115</u> PMID:<u>24138928</u>
- Kananen L, Marttila S, Nevalainen T, Kummola L, Junttila I, Mononen N, Kähönen M, Raitakari OT, Hervonen A, Jylhä M, Lehtimäki T, Hurme M, Jylhävä J. The trajectory of the blood DNA methylome ageing rate is largely set before adulthood: evidence from two longitudinal studies. Age (Dordr). 2016; 38:65. <u>https://doi.org/10.1007/s11357-016-9927-9</u> PMID:<u>27300324</u>
- 16. Chung KF, Adcock IM. Multifaceted mechanisms in COPD: inflammation, immunity, and tissue repair and destruction. Eur Respir J. 2008; 31:1334–56. <u>https://doi.org/10.1183/09031936.00018908</u> PMID:<u>18515558</u>
- Bousquet J, Jeffery PK, Busse WW, Johnson M, Vignola AM. Asthma. From bronchoconstriction to airways inflammation and remodeling. Am J Respir Crit Care Med. 2000; 161:1720–45. <u>https://doi.org/10.1164/ajrccm.161.5.9903102</u> PMID:<u>10806180</u>
- Hancox RJ, Poulton R, Greene JM, Filsell S, McLachlan CR, Rasmussen F, Taylor DR, Williams MJ, Williamson A, Sears MR. Systemic inflammation and lung function in young adults. Thorax. 2007; 62:1064–68. <u>https://doi.org/10.1136/thx.2006.076877</u> PMID:<u>17604302</u>
- Ward-Caviness CK, Nwanaji-Enwerem JC, Wolf K, Wahl S, Colicino E, Trevisi L, Kloog I, Just AC, Vokonas P, Cyrys J, Gieger C, Schwartz J, Baccarelli AA, et al. Longterm exposure to air pollution is associated with biological aging. Oncotarget. 2016; 7:74510–25. <u>https://doi.org/10.18632/oncotarget.12903</u> PMID:<u>27793020</u>
- Nwanaji-Enwerem JC, Colicino E, Trevisi L, Kloog I, Just AC, Shen J, Brennan K, Dereix A, Hou L, Vokonas P, Schwartz J, Baccarelli AA. Long-term ambient particle exposures and blood DNA methylation age: findings from the VA normative aging study. Environ Epigenet. 2016; 2:dvw006.

https://doi.org/10.1093/eep/dvw006 PMID:27453791

21. de F C Lichtenfels AJ, van der Plaat DA, de Jong K, van Diemen CC, Postma DS, Nedeljkovic I, van Duijn CM, Amin N, la Bastide-van Gemert S, de Vries M, Ward-Caviness CK, Wolf K, Waldenberger M, et al. Long-term air pollution exposure, genome-wide DNA methylation and lung function in the LifeLines cohort study. Environ Health Perspect. 2018; 126:027004. https://doi.org/10.1289/EHP2045 PMID:29410382

- 22. Sofer T, Baccarelli A, Cantone L, Coull B, Maity A, Lin X, Schwartz J. Exposure to airborne particulate matter is associated with methylation pattern in the asthma pathway. Epigenomics. 2013; 5:147–54. https://doi.org/10.2217/epi.13.16 PMID:23566092
- Lepeule J, Bind MA, Baccarelli AA, Koutrakis P, Tarantini L, Litonjua A, Sparrow D, Vokonas P, Schwartz JD. Epigenetic influences on associations between air pollutants and lung function in elderly men: the normative aging study. Environ Health Perspect. 2014; 122:566–72.

https://doi.org/10.1289/ehp.1206458 PMID:24602767

- 24. Holle R, Happich M, Löwel H, Wichmann HE, and MONICA/KORA Study Group. KORA—a research platform for population based health research. Gesundheitswesen. 2005 (Suppl 1); 67:S19–25. <u>https://doi.org/10.1055/s-2005-858235</u> PMID:<u>16032513</u>
- Rathmann W, Scheidt-Nave C, Roden M, Herder C. Type 2 diabetes: prevalence and relevance of genetic and acquired factors for its prediction. Dtsch Arztebl Int. 2013; 110:331–37. <u>https://doi.org/10.3238/arztebl.2013.0331</u> PMID:23762204
- 26. Rückert IM, Heier M, Rathmann W, Baumeister SE, Döring A, Meisinger C. Association between markers of fatty liver disease and impaired glucose regulation in men and women from the general population: the KORA-F4-study. PLoS One. 2011; 6:e22932. <u>https://doi.org/10.1371/journal.pone.0022932</u> PMID:21850244
- von Lengerke T, Mielck A, and KORA Study Group. Body weight dissatisfaction by socioeconomic status among obese, preobese and normal weight women and men: results of the cross-sectional KORA augsburg S4 population survey. BMC Public Health. 2012; 12:342.

https://doi.org/10.1186/1471-2458-12-342 PMID:<u>22571239</u>

 Gao X, Colicino E, Shen J, Just AC, Nwanaji-Enwerem JC, Wang C, Coull B, Lin X, Vokonas P, Zheng Y, Hou L, Schwartz J, Baccarelli AA. Accelerated DNA methylation age and the use of antihypertensive medication among older adults. Aging (Albany NY). 2018; 10:3210–28. https://doi.org/10.18632/aging.101626

PMID:30414594

29. Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, Crapo R, Enright P, van der Grinten CP,

Gustafsson P, Jensen R, Johnson DC, MacIntyre N, et al, and ATS/ERS Task Force. Standardisation of spirometry. Eur Respir J. 2005; 26:319–38. https://doi.org/10.1183/09031936.05.00034805 PMID:<u>16055882</u>

- Karrasch S, Flexeder C, Behr J, Holle R, Huber RM, Jörres RA, Nowak D, Peters A, Wichmann HE, Heinrich J, Schulz H, and KORA Study Group. Spirometric reference values for advanced age from a south german population. Respiration. 2013; 85:210–19. https://doi.org/10.1159/000338780 PMID:22797415
- Quanjer PH, Tammeling GJ, Cotes JE, Pedersen OF, Peslin R, Yernault JC. Lung volumes and forced ventilatory flows. Report Working Party Standardization of Lung Function Tests, European Community for Steel and Coal. Official Statement of the European Respiratory Society. Eur Respir J Suppl. 1993; 16:5–40. PMID:<u>8499054</u>
- 32. Sparrow D, O'Connor G, Colton T, Barry CL, Weiss ST. The relationship of nonspecific bronchial responsiveness to the occurrence of respiratory symptoms and decreased levels of pulmonary function. The Normative Aging Study. Am Rev Respir Dis. 1987; 135:1255–60. https://doi.org/10.1164/arrd.1987.135.6.1255 PMID:3296893
- Ferris BG. Epidemiology Standardization Project (American Thoracic Society). Am Rev Respir Dis. 1978; 118:1–120. PMID:742764

34. Rabe KF, Hurd S, Anzueto A, Barnes PJ, Buist SA, Calverley P, Fukuchi Y, Jenkins C, Rodriguez-Roisin R, van Weel C, Zielinski J, and Global Initiative for Chronic Obstructive Lung Disease. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. Am J Respir Crit Care Med. 2007; 176:532–55.

https://doi.org/10.1164/rccm.200703-456SO PMID:<u>17507545</u>

 Quanjer PH, Stanojevic S, Cole TJ, Baur X, Hall GL, Culver BH, Enright PL, Hankinson JL, Ip MS, Zheng J, Stocks J, and ERS Global Lung Function Initiative. Multiethnic reference values for spirometry for the 3-95-yr age range: the global lung function 2012 equations. Eur Respir J. 2012; 40:1324–43. <u>https://doi.org/10.1183/09031936.00080312</u>

PMID:22743675

- R Core Team. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. 2015.
- 37. Viechtbauer W. Conducting meta-analyses in R with the metafor package. J Stat Softw. 2010; 36:1–48. https://doi.org/10.18637/jss.v036.i03

SUPPLEMENTARY MATERIALS

Supplementary Figure



Supplementary Figure 1. Scatterplot and Pearson correlation coefficient. AAD and EEAD have a negative Pearson correlation (r) with $\triangle AAD$ (A) and $\triangle EEAD$ (B) respectively. AAD = age acceleration difference; EEAD = extrinsic epigenetic age acceleration difference; $\triangle AAD$ = change in age acceleration difference between baseline and follow-up examinations; $\triangle EEAD$ = change in extrinsic epigenetic age acceleration difference between baseline and follow-up examinations; $\triangle EEAD$ = change in extrinsic epigenetic age acceleration difference between baseline and follow-up examinations; $\triangle EEAD$ = change in extrinsic epigenetic age acceleration difference between baseline and follow-up.

Supplementary Tables

Supplementary	Table 1	Clinical	covariates
Supplementary	y lable I.	Cillical	covariates.

N. 400	Baseline	Follow-up
$\mathbf{N} = 408$	Mean (SD)	Mean (SD)
Age (yr)	71.3 (6.36)	77.6 (6.52)
Follow-up time (yr)	-	6.26 (3.33)
Weight (kg)	85.5 (14.1)	83.4 (14.5)
Height (cm)	173.9 (6.81)	173.3 (7.06)
BMI (kg/m2)	28.2 (4.03)	27.7 (4.17)
DNAmAge (yr)	72.1 (7.07)	75.5 (7.00)
EEAA (yr)	4.81(5.62)	4.02 (7.78)
AAD (yr)	0.77 (5.41)	-2.02 (7.72)
EEAD (yr)	4.81 (5.62)	4.02 (7.78)
FVC (liter)	3.39 (0.66)	3.37 (0.71)
FEV1 (liter)	2.57 (0.58)	2.48 (0.59)
FEF25-75 (liter/s)	4.24 (1.83)	3.54 (1.53)
FEV1/FVC	0.76 (0.07)	0.74 (0.07)
Binary Variables	N (%)	N (%)
Female	0 (0)	0 (0)
COPD GOLD (Incidence)	-	0 (0)
COPD GLI (Incidence)	-	15 (3.68)
White	399 (97.8)	399 (97.8)
Black	5 (1.23)	5 (1.23)
Hispanic	4 (0.98)	4 (0.98)
Ex-smoker	260 (63.7)	261 (64.0)
Current-smoker	17 (4.17)	17 (4.17)

Clinical covariates for the NAS study participants. BMI = body mass index; DNAmAge = DNA methylation age; EEAA = extrinsic epigenetic age acceleration; IEAA = intrinsic epigenetic age acceleration; AAD = age acceleration difference; EEAD = extrinsic epigenetic age acceleration difference; FEV1 = forced expiratory volume in 1 s; FVC = forced vital capacity; FEF25-75 = forced expiratory flow at 25-75% of FVC; COPD = Chronic obstructive pulmonary disease; GOLD = Global Initiative for Chronic Obstructive Lung Disease; GLI = Global Lung Initiative.

Supplementary Table 2. Associations between epigenetic aging biomarkers and COPD (a) and lung function (b) in KORA.

a. COPD					
	Epigenetic aging measure	Odds ratio	LCI	UCI	Р
COPD	AAD	1.00	0.99	1.02	0.63
COPD	EEAD	1.01	1.00	1.03	0.04
COPD	ΔΑΑD	1.01	0.98	1.03	0.62
COPD	ΔΕΕΑD	1.02	1.00	1.04	0.04
b. Lung function					

	Epigenetic aging measure	Effect estimate	LCI	UCI	Р
FEV1 (% difference from median)	AAD	-0.43	-1.64	0.77	0.48
FVC (% difference from median)	AAD	-0.17	-1.20	0.86	0.74
FEV1/FCV (% difference from median)	AAD	-0.22	-0.81	0.37	0.46
FEF25-75 (% difference from median)	AAD	-1.61	-4.05	0.83	0.20
FEV1 (% difference from median)	EEAD	-0.91	-1.89	0.07	0.07
FVC (% difference from median)	EEAD	-0.62	-1.45	0.22	0.15
FEV1/FCV (% difference from median)	EEAD	-0.32	-0.80	0.16	0.19
FEF25-75 (% difference from median)	EEAD	-1.34	-3.32	0.65	0.19
FEV1 (% difference from median)	ΔAAD	-0.85	-2.41	0.71	0.29
FVC (% difference from median)	ΔΑΑD	-0.65	-1.99	0.68	0.34
FEV1/FCV (% difference from median)	ΔΑΑD	-0.10	-0.86	0.66	0.80
FEF25-75 (% difference from median)	ΔΑΑD	-0.85	-3.98	2.27	0.59
FEV1 (% difference from median)	ΔΕΕΑD	-0.54	-1.82	0.74	0.41
FVC (% difference from median)	ΔΕΕΑD	-0.10	-1.19	0.99	0.86
FEV1/FCV (% difference from median)	ΔΕΕΑD	-0.54	-1.16	0.08	0.09
FEF25-75 (% difference from median)	ΔEEAD	-1.17	-3.74	1.39	0.37

AAD = age acceleration difference; ΔAAD = change in age acceleration difference between baseline and follow-up exams; COPD = chronic obstructive pulmonary disease; EEAD = extrinsic epigenetic age acceleration difference; $\Delta EEAD$ = change in extrinsic epigenetic age acceleration difference between baseline and follow-up exams; FEF25-75 = forced expiratory flow at 25-75% of pulmonary volume; FEV1 = forced expiratory volume in the first second; FVC = forced vital capacity; LCI = lower 95% confidence interval; P = p-value; UCI = upper 95% confidence interval.

a. COPD					
	Epigenetic aging measure	Odds ratio	LCI	UCI	Р
COPD	ΔEEAD	1.02	1.00	1.03	0.008044
COPD	ΔAAD	1.02	1.00	1.03	0.02502
COPD	AAD	1.02	1.00	1.04	0.03089
COPD	EEAD	1.02	0.99	1.04	0.485577
b. Lung function		1.01	0.77	1.02	0.105577
	Epigenetic aging measure	Effect estimate	LCI	UCI	Р
FEF25-75	ΔEEAD	-2.55	-5.21	0.10	0.06042
FEF25-75	ΔAAD	-2.33	-5.19	0.53	0.111603
FEF25-75	EEAD	-2.23	-5.50	1.04	0.182474
FEF25-75	AAD	-0.65	-4.20	2.89	0.717814
FEV1	ΔAAD	-1.32	-2.79	0.15	0.07916
FEV1	EEAD	-1.10	-2.75	0.56	0.194244
FEV1	ΔEEAD	-0.73	-2.10	0.64	0.297151
FEV1	AAD	0.28	-1.51	2.07	0.761846
FEV1/FVC	ΔEEAD	-0.79	-1.38	-0.20	0.008856
FEV1/FVC	ΔAAD	-0.79	-1.42	-0.15	0.015798
FEV1/FVC	EEAD	-0.59	-1.31	0.13	0.108924
FEV1/FVC	AAD	-0.49	-1.27	0.30	0.223369
FVC	AAD	0.96	-0.66	2.58	0.244423
FVC	ΔAAD	-0.49	-1.83	0.85	0.47577
FVC	EEAD	-0.31	-1.81	1.19	0.683633
FVC	ΔEEAD	0.10	-1.15	1.35	0.877678

Supplementary Table 3. Associations between epigenetic aging measures and COPD (a) and lung function (b) in NAS.

AAD = age acceleration difference; ΔAAD = change in age acceleration difference between baseline and follow-up exams; COPD = chronic obstructive pulmonary disease; EEAD = extrinsic epigenetic age acceleration difference; $\Delta EEAD$ = change in extrinsic epigenetic age acceleration difference between baseline and follow-up exams; FEF25-75 = forced expiratory flow at 25-75% of pulmonary volume; FEV1 = forced expiratory volume in the first second; FVC = forced vital capacity; LCI = lower 95% confidence interval; P = p-value; UCI = upper 95% confidence interval.

a. COPD					
Outcome	Epigenetic aging measure	OR	OR LCI	OR UCI	Р
COPD	EEAD	1.01	1.00	1.02	0.043004
COPD	AAD	1.01	1.00	1.02	0.06543
COPD	ΔAAD	1.01	1.00	1.03	0.031643
COPD	ΔΕΕΑD	1.02	1.01	1.03	0.000706
b. Lung function					
	Epigenetic aging measure	Effect estimate	LCI	UCI	Р
FEV1	AAD	-0.21	-1.21	0.79	0.677415
FEV1	EEAD	-0.96	-1.80	-0.12	0.025818
FEV1	ΔAAD	-1.10	-2.17	-0.03	0.044241
FEV1	ΔEEAD	-0.63	-1.56	0.31	0.1883
FVC	AAD	0.16	-0.71	1.02	0.72534
FVC	EEAD	-0.54	-1.27	0.19	0.145254
FVC	ΔAAD	-0.57	-1.52	0.37	0.236804
FVC	ΔΕΕΑD	-0.01	-0.84	0.81	0.973866
FEV1/FVC	AAD	-0.32	-0.79	0.15	0.185455
FEV1/FVC	EEAD	-0.40	-0.80	-0.01	0.046866
FEV1/FVC	ΔAAD	-0.50	-0.99	-0.02	0.04325
FEV1/FVC	ΔEEAD	-0.67	-1.10	-0.25	0.001988
FEF25-75	AAD	-1.30	-3.31	0.71	0.204109
FEF25-75	EEAD	-1.58	-3.27	0.12	0.068706
FEF25-75	ΔAAD	-1.65	-3.76	0.46	0.124265
FEF25-75	ΔEEAD	-1.84	-3.68	0.01	0.050812

Supplementary Table 4. Meta-analysis of KORA and NAS for COPD (a) and lung function (b) outcomes.

AAD = age acceleration difference; Δ AAD = change in age acceleration difference between baseline and follow-up exams; COPD = chronic obstructive pulmonary disease; EEAD = extrinsic epigenetic age acceleration difference; Δ EEAD = change in extrinsic epigenetic age acceleration difference between baseline and follow-up exams; FEF25-75 = forced expiratory flow at 25-75% of pulmonary volume; FEV1 = forced expiratory volume in the first second; FVC = forced vital capacity; LCI = lower 95% confidence interval; P = p-value; UCI = upper 95% confidence interval.