

Comprehensive analysis of expression and prognostic value of the claudin family in human breast cancer

Guangda Yang¹, Liumeng Jian², Qianya Chen¹

¹Department of Cancer Chemotherapy, Zengcheng District People's Hospital of Guangzhou, Guangdong Province, China

²Department of Neurology, Zengcheng District People's Hospital of Guangzhou, Guangdong Province, China

Correspondence to: Liumeng Jian; **email:** jinlium2012@163.com, <https://orcid.org/0000-0002-0558-1655>

Keywords: claudins, breast cancer, prognosis, ONCOMINE, bc-GenExMiner v4.3

Received: December 2, 2020

Accepted: January 25, 2021

Published: March 10, 2021

Copyright: © 2021 Yang et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/3.0/) (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Claudins (CLDN) are structural components of tight junctions that function in paracellular transport and maintain the epithelial barrier function. Altered expression and distribution of members of the claudin family have been implicated in several cancers including breast cancer (BC). We performed a comprehensive analysis of the expression and prognostic value of claudins in BC using various online databases. Compared with normal tissues, CLDN3, 4, 6, 7, 9, and 14 were upregulated in BC tissues, whereas CLDN2, 5, 8, 10, 11, 15, 19, and 20 were downregulated. A high expression of CLDN2, 5, 6, 9, 10, 11, and 14–20 was associated with better relapse-free survival (RFS), whereas a high CLDN3 expression correlated with poor RFS. In addition, a high expression of CLDN3, 4, 14, and 20 was associated with poor overall survival (OS), whereas that of CLDN5 and CLDN11 was linked to a better OS. Although METABRIC and TCGA datasets revealed 22% and 27% gene alterations, respectively, in the members of the claudin family, these were not associated with survival. These findings suggest CLDN3, 5, and 11 could be promising therapeutic targets for patients with BC.

INTRODUCTION

Breast cancer (BC) is one of the most commonly diagnosed cancers worldwide. Although its incidence rates have declined continuously due to improvements in treatment strategies and early detection, BC is still the second leading cause of cancer-related deaths among women [1]. BC is a genetically heterogeneous group of tumors with a variety of morphologic features and is classified into four distinct molecular subtypes based on the immunohistochemical expression of estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (Her2): ER/PR+, Her2– (Luminal A); ER/PR+, Her2+ (Luminal B); ER/PR–, Her2+ (HER2+); and ER/PR–, Her2– (basal-like/triple-negative breast cancer [TNBC]) [2]. Endocrine therapy can improve the survival rate of patients with luminal subtype BC, whereas trastuzumab

is effective against Her2+ subtype BC [3–5]. However, chemotherapy is the only available treatment approach against basal-like BC and TNBC due to the lack of effective biomarkers. Interestingly, a novel claudin-low molecular subtype of BC has been recently identified [6], characterized by low expression of tight junction and epithelial cell–cell adhesion proteins, including claudin 3, 4, and 7, and E-cadherin [6]. In addition, claudin-low tumors preferentially display a triple-negative phenotype, with enhanced epithelial-to-mesenchymal transition (EMT) features, immune system responses, and stem cell-associated biological processes [7–10]. Moreover, patients with claudin-low BC have a poor overall survival (OS) when compared to those with luminal A subtype BC [10].

Claudins (CLDNs) are structural and functional components of tight junctions that regulate cellular

adhesion and maintain cell polarity in epithelial and endothelial cell sheets [11, 12]. The human genome consists of 23 annotated CLDN genes (they lack CLDN13) [11, 12]. Claudins are abnormally expressed in several human cancers and could be used as promising targets for cancer detection, diagnosis, and treatment [13]. Literature reports only a few studies on the expression and functions of some claudin genes in BC ([14–33] Supplementary Table 1). We believe that understanding the expression patterns, functional roles, and prognostic values of claudins would assist in identifying potential therapeutic targets and survival biomarkers for BC.

The advent of microarray technology has revolutionized the way DNA and RNA research is conducted [34]. We comprehensively analyzed different claudin genes using various online databases to determine their expression patterns, potential functions, and distinct prognostic values in patients with BC.

RESULTS

Transcriptional levels of claudins in patients with breast cancer

We first compared the mRNA levels of claudins in BC and the corresponding normal samples using the ONCOMINE database (Figure 1 and Supplementary Table 2). Details of major datasets of the claudin family in BC are shown in Supplementary Table 2 [35–46]. In total, we identified 22 claudins in BC samples.

The results indicated that the mRNA expression of CLDN2, 5, 8, 10, 11, 16, 18, 19, 22, and 23 was

downregulated in BC samples (Figure 1 and Supplementary Table 2). The decreased mRNA expression of CLDN2 was found in two datasets with a fold change of -4.357 and -2.283 . The mRNA expression of CLDN5 was only upregulated in one study but downregulated in 18 studies. The mRNA expression of CLDN8 was downregulated with a fold change ranging from -2.382 to -24.488 in all 18 studies of six published datasets and the TCGA dataset. The mRNA expression of CLDN10 was downregulated with a fold change ranging from -2.117 to -7.292 in all nine studies of two published datasets and the TCGA dataset. In addition, the mRNA expression of CLDN11 was downregulated with a fold change ranging from -2.382 to -13.276 in all 18 studies of five published datasets and the TCGA dataset. The mRNA expression of CLDN19 was downregulated with a fold change ranging from -3.652 to -10.49 in all six studies of the TCGA dataset. In addition, the TCGA dataset revealed that the mRNA expression of CLDN16 ($p = 0.001$, fold change = -2.053) decreased in intraductal cribriform breast adenocarcinoma. In the BC dataset of Turashvili’s study [38], CLDN18 was downregulated in invasive ductal breast carcinoma with a fold change of -2.301 ($p = 1.89E-04$) and CLDN23 was downregulated in invasive lobular breast carcinoma with a fold change of -3.776 ($p = 0.025$). Moreover, the TCGA dataset revealed reduced mRNA expression of CLDN22 ($p = 5.33E-06$, fold change = -2.229) in invasive ductal and lobular carcinoma.

The results showed that the mRNA expression of CLDN7, 9, and 14 was upregulated in BC samples (Figure 1 and Supplementary Table 2). The mRNA expression of CLDN7 was upregulated with a fold change ranging

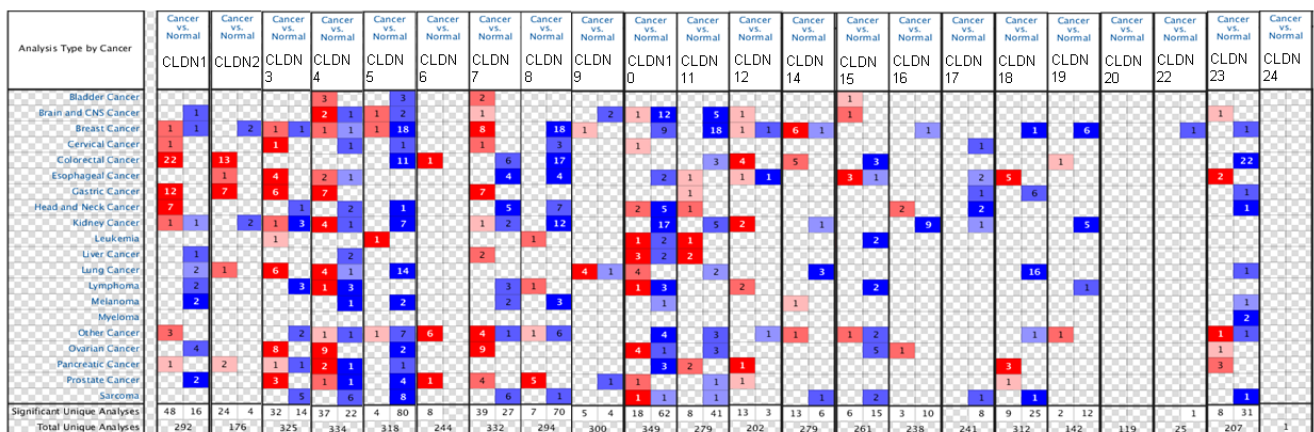


Figure 1. The mRNA expression of the claudin family in different types of cancers (ONCOMINE). Notes: The figure is generated from ONCOMINE with exact thresholds (p -value: 0.05; fold change: 2; gene rank: top 10%). The cell number represents the dataset number that meets all the thresholds, with blue for underexpression and red for overexpression. The cell color is determined by the best gene rank percentile for the analyses within the cell.

from 2.007 to 3.625 in all eight studies of three datasets. CLDN9 was overexpressed in the BC dataset of Finak's study with a fold change of 3.269 [40]. In addition, the mRNA expression of CLDN14 was only downregulated in one study but upregulated in six studies. However, the mRNA expression of CLDN1, 3, 4, and 12 was both overexpressed and underexpressed in one study. Unfortunately, we did not find any study on the mRNA expression of CLDN6, 15, 17, 20, and 24 in samples obtained from patients with BC and normal individuals (Figure 1 and Supplementary Table 2).

To further confirm the expression of members of the claudin family, we used the ULACAN database to compare the mRNA levels of claudins in the samples obtained from patients with BC with those obtained from normal individuals (Supplementary Table 3). We found that the mRNA expression of CLDN3, 4, 6, 7, 9, and 14 was higher in BC tissues than in normal tissues (Figure 2A, 2B, 2D, 2E, 2G, 2I), whereas the mRNA expression of CLDN5, 8, 11, 15, 19, and 20 was higher in normal tissues than in BC tissues (Figure 2C, 2F, 2H, 2J–2L). Therefore, comprehensive results indicated that the mRNA expression of CLDN3, 4, 6, 7, 9, and 14 was upregulated in patients with BC compared with normal individuals, whereas that of CLDN2, 5, 8, 10, 11, 15, 19, and 20 was downregulated in patients with BC. The expression of other claudin genes remains controversial.

In addition, we analyzed the promoter methylation levels of claudins in BC and normal tissues. The beta values of CLDN2, 4, 5, 6, 9, 15, 16, 17, 18, 19, and 20 indicated hypermethylation, whereas those of CLDN1, 7, 8, 10, 11, 12, and 23 indicated hypomethylation (Supplementary Table 3). However, we found that CLDN1, 5, 6, 9, 10, 11, 15, 19, and 23 had higher promoter methylation levels in BC tissues than in normal tissues, whereas CLDN2, 4, 7, 8, 12, 16, 17, 18, and 20 had lower promoter methylation levels in BC tissues than in normal tissues (Supplementary Figure 1).

Relationship between mRNA levels of claudins and the clinicopathologic parameters of patients with breast cancer

We next analyzed the relationship between claudins and the clinicopathologic parameters of BC using the bc-GenExMiner v4.3 database (Supplementary Table 4). With respect to age, the mRNA expression of CLDN2, 8, 10, 11, 19, and 23 was low in the age group above 51 years, whereas only CLDN3 mRNA was upregulated in the older group. The mRNA expression of CLDN3, 4, 7, and 15 was upregulated, whereas that of CLDN2, 10, 19, and 23 was downregulated in the node-positive BC group. In addition, the mRNA expression of CLDN3, 5,

7, 11, and 12 was upregulated, whereas that of CLDN1, 4, 6, 8, 9, 10, 14, 16, 17, 23, and 24 was downregulated in patients positive for estrogen receptor (ER). Patients positive for progesterone receptor (PR) had a higher mRNA expression of CLDN5, 7, 11, and 12 and lower expression of CLDN1, 2, 4, 6, 8, 9, 10, 14, 16, 22, 23, and 24 mRNAs as compared with normal samples. Compared with the HER2-negative group, CLDN2 and CLDN4 mRNAs were overexpressed in the human epidermal growth factor receptor 2 (HER2)-positive group. Moreover, patients with HER2-positive BC had reduced mRNA expression of CLDN5 and CLDN12.

TNBC is a special type of BC with negative ER, PR, and HER2. Although the triple-negative status positively correlated with CLDN1, 6, 8, 9, 10, 16, 20, and 23, it had a negative correlation with CLDN3, 5, 7, 11, 12, and 19 mRNA expression. Moreover, basal-like status was positively correlated with CLDN1, 2, 4, 6, 8, 9, 10, 14, 16, 22, and 23 but negatively correlated with CLDN3, 5, 7, 11, 12, and 19 mRNA expression (Supplementary Table 4).

Scarff–Bloom–Richardson (SBR) grading system is considered a prognostic factor in BC. A higher SBR grade status correlated with a higher mRNA expression of CLDN3 and CLDN4 (Figure 3A, 3B) and with lower mRNA expression of CLDN5, 11, and 12 (Figure 3C, 3H, 3I). For CLDN6, 7, 9, 10, 14, 15, 16, 17, 23, and 24 (Figure 3D–3G, 3J–3O), although a substantial difference was detected in Welch's test, certain comparison groups by Dunnett's Tukey–Kramer test did not show a difference (the cutoff value of p was 0.05) (Supplementary Table 5). Other claudin genes showed no difference in the SBR grade status (Supplementary Table 5).

The Nottingham prognostic index (NPI) is another prognostic model for patients with BC. A higher NPI grade status was found to be correlated with higher mRNA expression of CLDN3 and CLDN9 (Figure 4B, 4G). For CLDN1, 4, 5, 6, 7, 10, 11, 12, and 23 (Figure 4A, 4C–4F, 4H–4K), not all pairwise comparisons in the NPI criteria were significant ($p < 0.05$) (Supplementary Table 5). Other claudin genes showed no difference in the NPI grade status (Supplementary Table 5).

Prognostic values of claudin mRNA expression in all breast cancer groups

The Kaplan–Meier plotter was used to examine the prognostic values of claudin mRNA expression in all BC groups. Figure 5 shows all relapse-free survival (RFS) curves associated with the members of the claudin family. The results revealed that a high mRNA

expression of CLDN2, 5, 6, 9, 10, 11, 14–20 was associated with better RFS (Figure 5A, 5C–5N). In contrast, a high mRNA expression of CLDN3 was associated with a poor prognosis of RFS (Figure 5B). In

addition, we analyzed the relationship between mRNA expression of claudins and other prognostic indexes, including overall survival (OS), distant metastasis-free survival (DMFS), and post-progression survival (PPS).

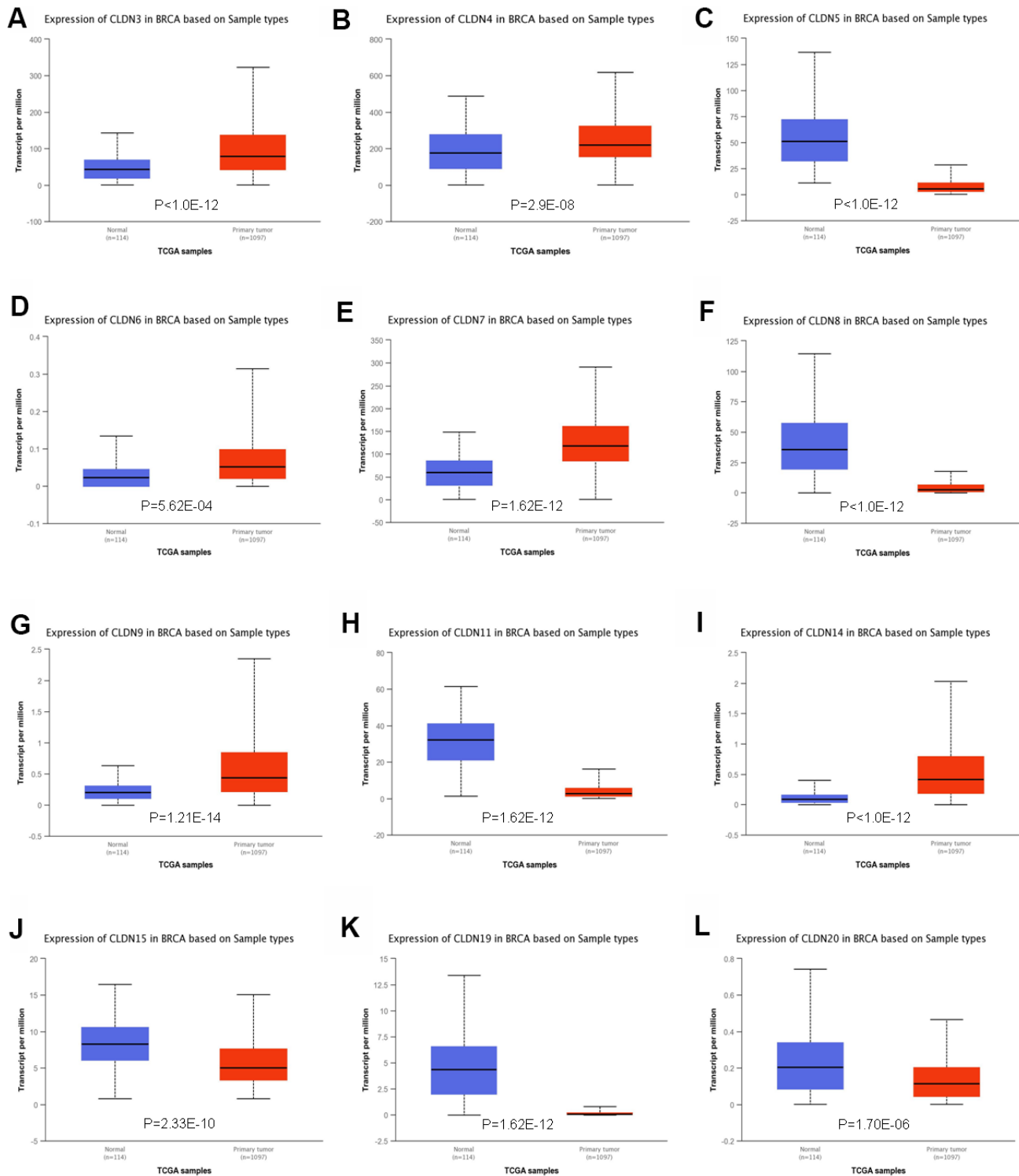


Figure 2. Significant changes in claudin mRNA expression between breast cancer and normal tissues (UALCAN database). (A) CLDN3; **(B)** CLDN4; **(C)** CLDN5; **(D)** CLDN6; **(E)** CLDN7; **(F)** CLDN8; **(G)** CLDN9; **(H)** CLDN11; **(I)** CLDN14; **(J)** CLDN15; **(K)** CLDN19; **(L)** CLDN20.

Figure 6 shows all OS curves associated with members of the claudin family. We found that a high mRNA expression of CLDN3, 4, 14, and 20 was associated with poor OS (Figure 6A, 6B, 6E, 6F), whereas a high

mRNA expression of CLDN5 and CLDN11 was associated with better OS (Figure 6C, 6D). Similarly, a high mRNA expression of CLDN3 and CLDN7 was associated with a poor prognosis of DMFS and a high

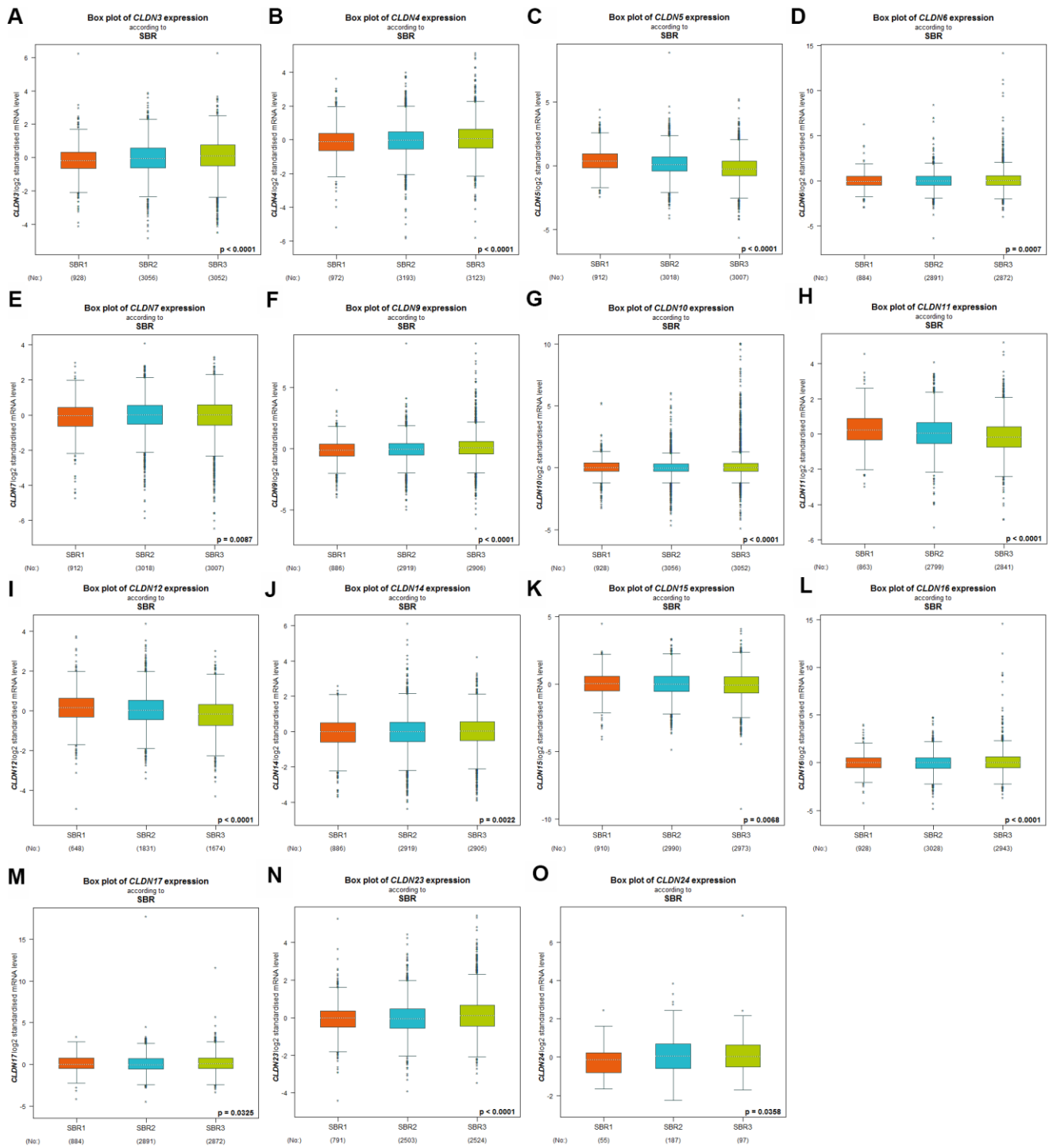


Figure 3. Relationship between the claudin family and the SBR criterion. (A) CLDN3; (B) CLDN4; (C) CLDN5; (D) CLDN6; (E) CLDN7; (F) CLDN9; (G) CLDN10; (H) CLDN11; (I) CLDN12; (J) CLDN14; (K) CLDN15; (L) CLDN16; (M) CLDN17; (N) CLDN22; (O) CLDN24. Global differences between the groups were assessed by Welch's test, and $p < 0.05$ was considered significant, with Dunnett's modified Tukey–Kramer test computed for each pairwise SBR comparison. Abbreviations: AQP, aquaporin; NPI, Nottingham Prognostic Index; SBR, Scarff–Bloom–Richardson.

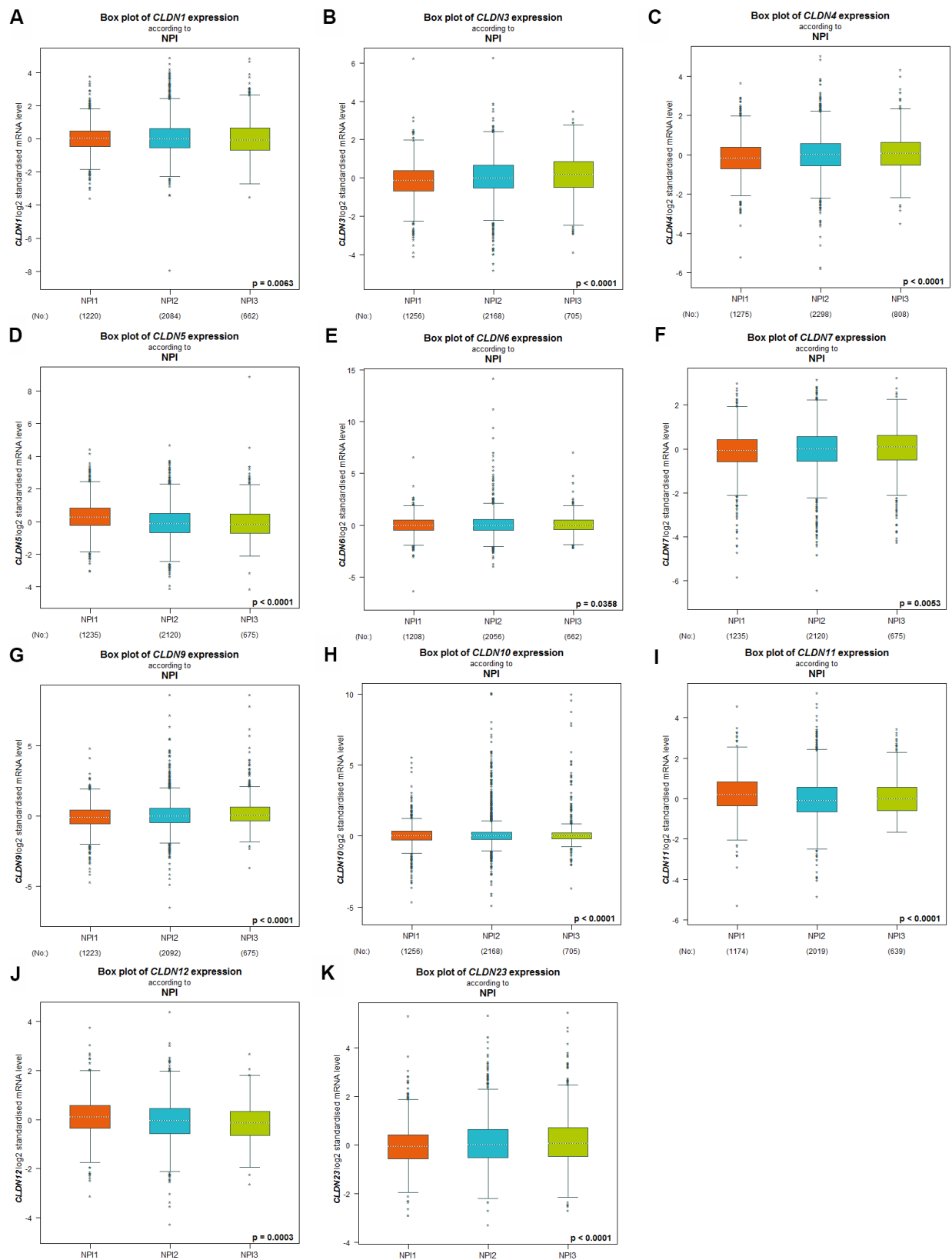


Figure 4. Relationship between the Claudin family and the NPI criterion. (A) *CLDN1*; (B) *CLDN3*; (C) *CLDN4*; (D) *CLDN5*; (E) *CLDN6*; (F) *CLDN7*; (G) *CLDN9*; (H) *CLDN10*; (I) *CLDN11*; (J) *CLDN12*; (K) *CLDN23*. Global differences between the groups were assessed by Welch's test, and $p < 0.05$ was considered significant, with Dunnett's Tukey–Kramer test computed for each pairwise comparison. Abbreviation: NPI, Nottingham Prognostic Index.

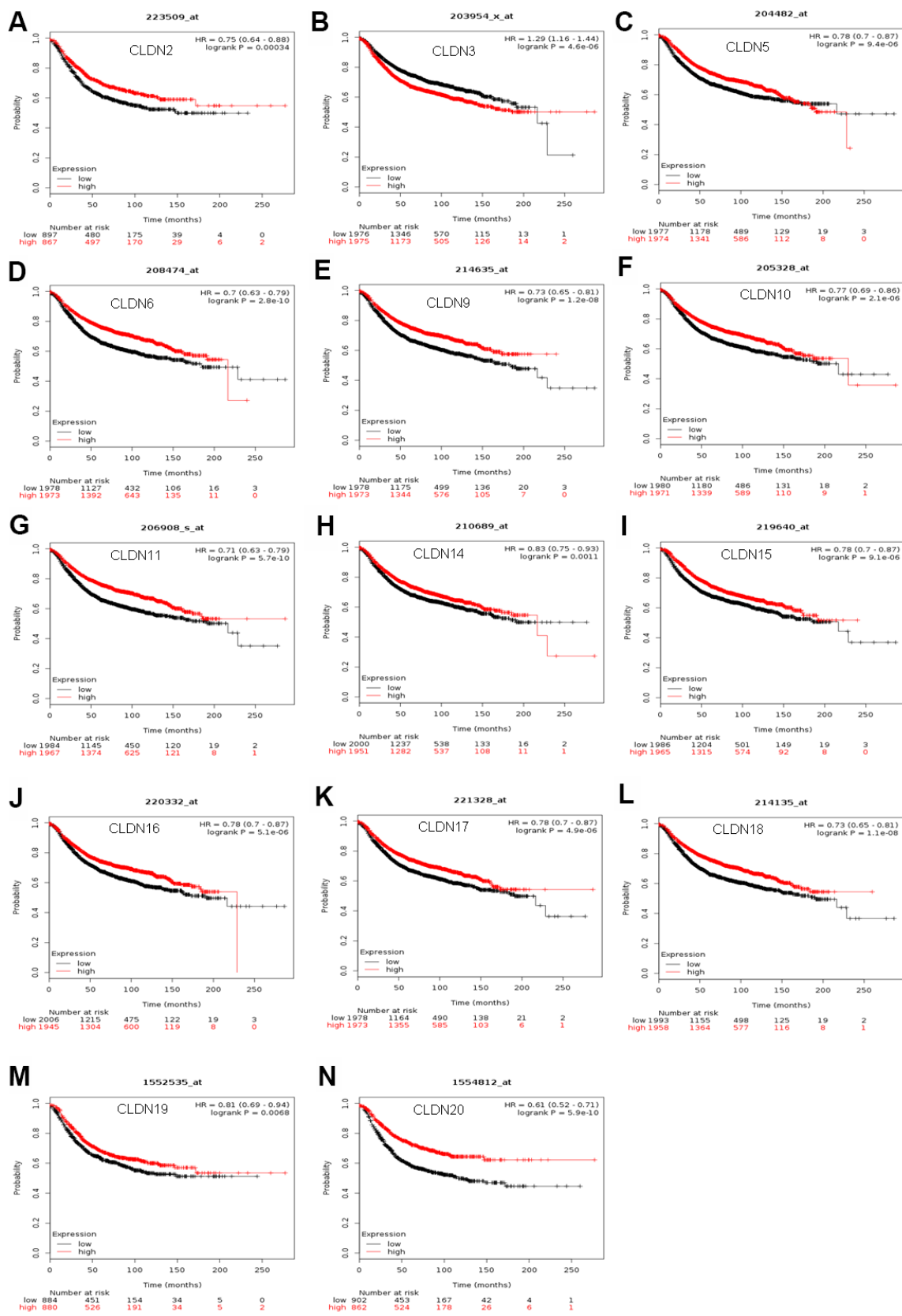


Figure 5. The RFS of mRNA levels of claudins in all patients with breast cancer (Kaplan–Meier plotter). (A) CLDN2; (B) CLDN3; (C) CLDN5; (D) CLDN6; (E) CLDN9; (F) CLDN10; (G) CLDN11; (H) CLDN14; (I) CLDN15; (J) CLDN16; (K) CLDN17; (L) CLDN18; (M) CLDN19; (N) CLDN20. Abbreviation: RFS, relapse-free survival.

CLDN2 mRNA expression indicated better DMFS (Supplementary Table 6). In addition, a high mRNA expression of CLDN3, 4, and 14 was associated with poor PPS and a high mRNA expression of CLDN6 and CLDN18 indicated better PPS (Supplementary Table 6).

Prognostic values of claudin mRNA expression in different molecular subtypes of breast cancer

We next analyzed the prognostic values of claudin mRNA expression in different molecular subtypes,

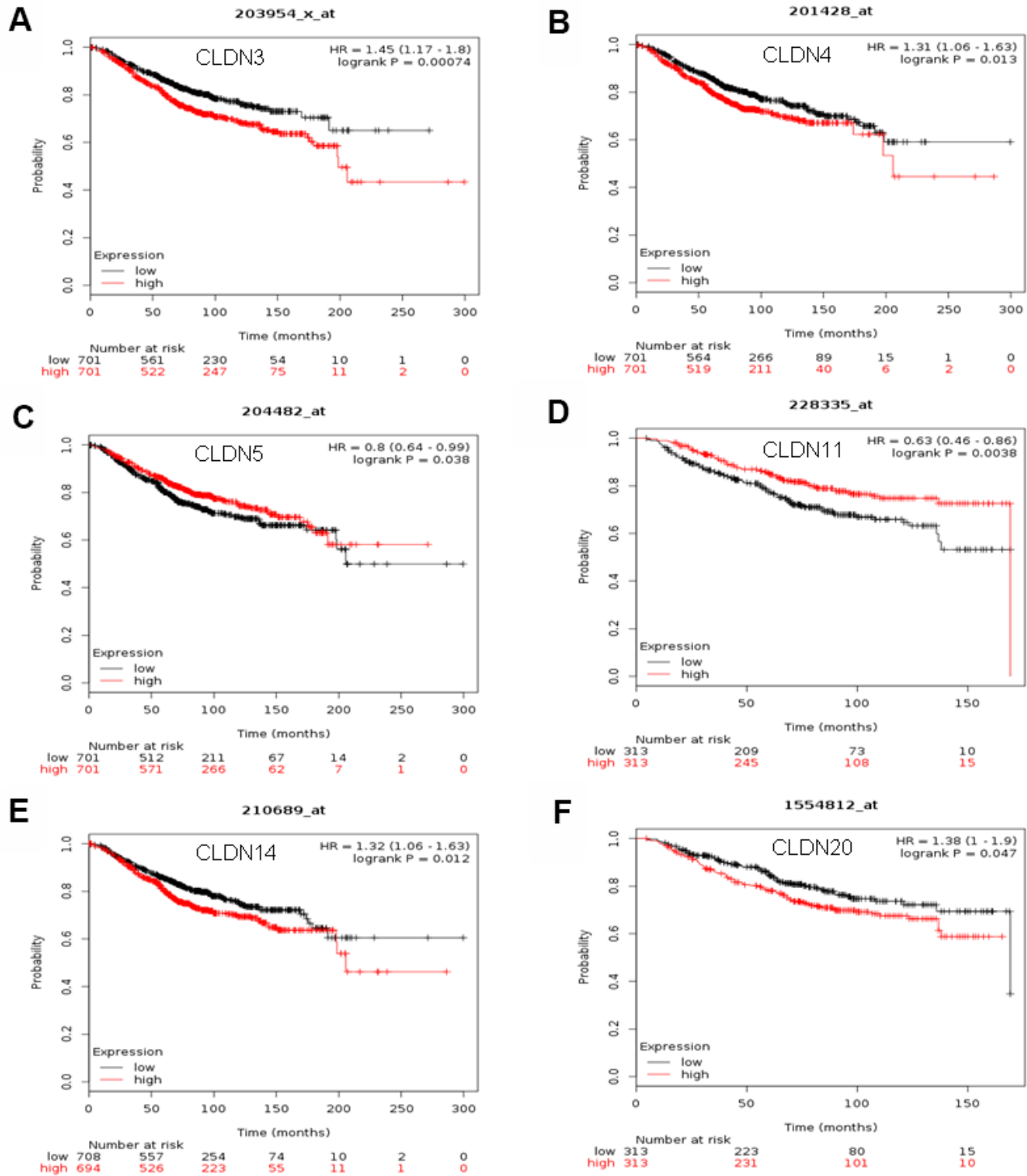


Figure 6. The OS of mRNA levels of claudins in all patients with breast cancer (Kaplan–Meier plotter). (A) CLDN3; (B) CLDN4; (C) CLDN5; (D) CLDN11; (E) CLDN14; (F) CLDN20. Abbreviations: OS, overall survival.

including basal-like, luminal A, luminal B, and HER2+ (Supplementary Table 7).

In basal-like BC, a high mRNA expression of CLDN1 and CLDN7 correlated with poor RFS, whereas a high mRNA expression of CLDN6, 9, 10, 16, and 18 correlated with better RFS. In addition, a high mRNA expression of CLDN8 indicated better OS, whereas that of CLDN10 was associated with better DMFS. Moreover, a high mRNA expression of CLDN20 indicated poor PPS. The mRNA expression of other members of the claudin family members did not correlate with prognosis in basal-like BC.

In patients with luminal A BC, a high mRNA expression of CLDN1, 2, 5, 6, 8, 9, 10, 11, and 14 to 20 correlated with better RFS, whereas only a high CLDN3 expression was associated with poor RFS. In addition, a high mRNA expression of CLDN2 was associated with the poor OS but indicated better DMFS. Other members of the claudin family were not associated with any prognosis in patients with luminal A BC.

In patients with luminal B BC, a high mRNA expression of CLDN6, 8, 9, 10, 11, 15–18, 20, and 23 correlated with better RFS. In addition, a high expression of CLDN7 was associated with poor RFS and OS. No correlation with prognosis was found in the remaining members of the claudin family.

In patients with HER2+ BC, a high mRNA expression of CLDN9 and CLDN14 correlated with better RFS, whereas only a high CLDN8 expression was associated with poor RFS. In addition, a high expression of CLDN20 was associated with poor OS. No correlation with prognosis was found in the remaining members of the claudin family.

Prognostic values of claudin mRNA expression in breast cancer with different clinicopathologic classifications

Next, we investigated the prognostic values of claudin mRNA expression in BC with different clinicopathologic classifications, including lymph node status and histologic grades (Supplementary Table 8). We found that a high mRNA expression of CLDN6, 9, 17, 18, and 20 correlated with poor PPS, whereas only high CLDN1 expression had better PPS in patients with lymph node positivity. In addition, a high CLDN11 expression was associated with better RFS in patients with lymph node positivity. A high CLDN8 expression resulted in poor DMFS, whereas a high CLDN20 expression was associated with poor OS in patients with lymph node positivity. In patients with lymph node negativity, a high CLDN3 expression correlated with

the poor OS, DMFS, and PPS, whereas a high CLDN8 expression correlated with better DMFS. In addition, a high mRNA expression of CLDN9, 18, and 19 correlated with better PPS, whereas a high CLDN20 expression was associated with poor PPS. Moreover, a high CLDN9 expression indicated a better OS.

The second clinicopathologic classification we investigated was histologic grade (Supplementary Table 9). In patients with grade 1 BC, only high CLDN11 expression indicated better RFS, whereas high expression of CLDN3 and CLDN4 correlated with poor OS in patients with grade 2 BC, whereas a high CLDN12 expression had better OS. In addition, a high CLDN3 expression correlated with poor DMFS. A high expression of CLDN3, 4, 10, 16, and 18 correlated with poor PPS, whereas a high CLDN12 expression had better PPS. In patients with grade 3 BC, a high expression of CLDN1, 3, and 7 correlated with poor RFS, whereas a high CLDN16 expression was found to have better RFS. In addition, a high expression of CLDN1 and CLDN7 correlated with poor DMFS.

Claudin gene alteration analysis

We used the cBioPortal for Cancer Genomics database to analyze the alterations in the genes of the claudin family. As for the TCGA dataset (with 963 patients), 256 (27%) patients had altered claudin genes (Figure 7A). In addition, 485 (22%) patients had altered claudin genes as per the METABRIC dataset (Figure 7B). The genetic alterations included missense mutations, truncating mutations, amplifications, and deep deletions. However, the results from the two datasets showed no differences between OS/DFS and BC patients with or without alterations in claudin genes (Figure 7C–7E).

DISCUSSION

We explored the mRNA expression, the correlation with clinicopathologic parameters, and prognostic values of 22 claudin genes in patients with BC. The majority of these genes had altered expression that could impact the survival of patients with BC. CLDN3, 5, and 11 could be used as promising therapeutic targets for BC. Compared with normal tissues, the expression of CLDN3, 4, and 7 was upregulated in BC tissues, a finding consistent with that of a previous study that reported elevated expression of these claudins in BC [22]. Although CLDN3 is known to function as a tumor suppressor in certain cancers [47–51], we found that a high CLDN3 expression was associated with poor RFS, OS, DMFS, and PPS in all patients. A high expression of CLDN3 and CLDN7 was associated with poor RFS in TNBC [23]. Moreover, CLDN3 expression correlated

with SRB and NPI. In addition, CLDN3 expression correlated with almost all clinicopathologic parameters and impacted the prognosis in patients with lymph node negativity or grade 2 BC. These results indicated that CLDN3 could serve as a potential therapeutic intervention for BC.

Although a previous study associated upregulated CLDN5 expression in BC with poor RFS [27], we found that the low expression of CLDN5 and CLDN11 was associated with poor RFS and OS in all patients. Moreover, both of them correlated with SRB and with almost all clinicopathologic parameters. Similarly, another study found upregulated CLDN11 expression in BC [21]. However, the samples in these studies were small. Moreover, overexpressed CLDN5 reduced the paracellular permeability of hCMC/D3 cells and decreased the invasion of lung adenocarcinoma A549 cells [52]. Insertion of *Claudin-5* gene in HECV cells substantially reduced the motility of the cells and their adhesiveness to the matrix, along with reduced angiogenic potential [53]. Downregulation of claudin-5 increased the permeability of the blood-tumor barrier [54, 55], suggesting it could prevent brain metastasis. Low CLDN11 expression has been used as a prognosis biomarker in certain cancers [56–58]. Moreover, miR-

99b-induced downregulation of CLDN11 promoted metastasis of hepatocellular carcinoma [59]. Similarly, miR-421 promoted the proliferation and metastasis of gastric cancer cells by targeting claudin-11 [60]. In addition, low expression of CLDN5 and CLDN11 is associated with poor RFS in patients with luminal A BC. Based on these bioinformatics data, we speculate that the downregulated expression of CLDN5 and CLDN11 could be exploited to devise therapeutic strategies against BC.

Our study had some limitations. First, the expression or prognostic values of certain claudins in BC or its subtypes are still unknown due to limited samples. Second, mechanisms regulating the expression of claudins remain elusive. Based on the literature and functional modules available in online databases, we analyzed the methylation levels of claudins in BC. The results showed that CLDN4, 5, 6, 15, and 19 were hypermethylated in BC with lower mRNA expression compared to normal tissues. However, although CLDN16, 17, and 18 were hypermethylated, their mRNA expression did not differ. Interestingly, we found that CLDN11 was hypomethylated in BC with lower mRNA expression than in normal tissues. Methylation of CLDN11 promoter is known to be

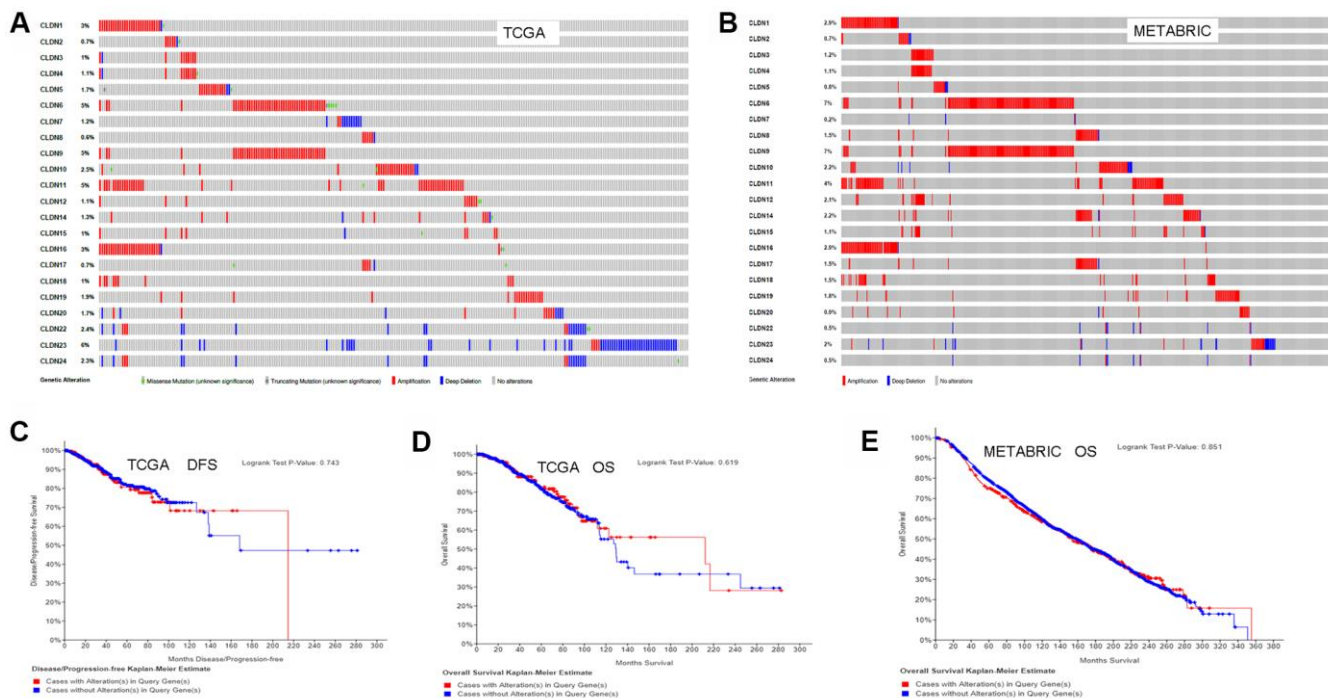


Figure 7. Analysis of alterations in claudins in breast cancer (using cBioPortal for Cancer Genomics). (A) OncoPrint of the TCGA dataset in cBioPortal. (B) OncoPrint of the METABRIC dataset in cBioPortal. (C) DFS analysis in cases with or without alterations in claudin genes of the TCGA dataset. (D) OS analysis in cases with or without alterations in claudin genes of the TCGA dataset. (E) OS analysis in cases with or without alterations in claudin genes of the METABRIC dataset. OncoPrint represents the distribution and proportion of samples with different kinds of alterations in the claudin family. Abbreviations: DFS, disease/progression-free survival; OS, overall survival.

associated with the development and poor survival in several cancers [56–58]. Moreover, miRNA-induced reduced expression of CLDN11 promoted metastasis [59, 60], implying DNA methylation as one of the underlying regulatory mechanisms of at least certain claudins in BC. Third, this study showed that certain claudins functioned as tumor suppressors. Fourth, the results of bioinformatics analysis need to be validated by performing biological experiments.

In summary, we systematically analyzed the expression and prognostic value of claudins in BC. Our findings suggested that CLDN3, 5, and 11 could be used as promising therapeutic targets for BC.

MATERIALS AND METHODS

ONCOMINE analysis

ONCOMINE (<http://www.oncomine.com>), an online cancer microarray database, was used to analyze the mRNA levels of the claudin family in different cancers [61]. The search filters were set to the following: differential analysis (cancer vs. normal), cancer type (breast cancer), sample type (clinical specimen), data type (mRNA), and genes (CLDN1-12, CLDN14-20, and CLDN22-24). Thresholds were set as the following: gene rank, 10%; fold change, 2; and *p*-value, 0.05.

UALCAN database

UALCAN (<http://ualcan.path.uab.edu/index.html>) is a portal for facilitating tumor subgroup gene expression and survival analyses [62]. It was used to evaluate the mRNA levels and promoter methylation levels of the claudin family in patients with BC and normal individuals. In addition, we evaluated the effects of the claudin family on the survival of patients with BC. The beta value indicates the level of DNA methylation ranging from 0 (unmethylated) to 1 (fully methylated). Different beta cut-off values have been considered to indicate hypermethylation (beta value: 0.7–0.5) or hypo-methylation (beta-value: 0.3–0.25). A *p*-value < 0.05 was considered significant.

Breast cancer gene expression miner (bc-GenExMiner) v4.3

bc-GenExMiner v4.3 (<http://bcgenex.centregauducheau.fr/BC-GEM/GEM-Accueil.php?js=1>) [63], a statistical mining tool of published annotated BC transcriptomic data, was used to assess the correlation of expression of members of the claudin family with specific clinicopathologic features of BC, including age, nodal status, hormonal receptors status (ER and PR), HER2,

pathologic subtype, NPI, and SBR grade. A *p*-value < 0.05 was considered significant.

Kaplan–Meier plotter database

Kaplan–Meier plotter (<https://kmplot.com/analysis/>) [64], an online database established using gene expression data and survival information of 1,809 patients with BC downloaded from GEO, was used to analyze the prognostic values of members of the claudin family in all patients, in different molecular subtypes of BC, and different kinds of clinicopathologic classifications of BC. Only the best probe set of the claudin family was selected for this analysis. The hazard ratio (HR) with 95% confidence intervals and log-rank *p*-value were calculated and shown.

The cancer genome atlas (TCGA) data and cBioPortal for cancer genomics database

The Cancer Genome Atlas consists of both sequencing and pathological data on 30 different cancers [65]. The cBioPortal for Cancer Genomics (<http://www.cbioportal.org>) contains large-scale cancer genomics datasets with functions such as visualization, downloading, and analysis [66]. We selected two BC datasets with most patients (TCGA, Provisional and METABRIC, Nature 2012 and Nat Commun, 2016) for further analysis using cBioPortal. The OncoPrint, OS, and DFS of the claudin family were analyzed online.

Statistical analysis

Gene expression data from the Oncomine database were analyzed using the *p*-values, fold changes, and ranks. Gene expression and methylation levels from the UALCAN or bc-GenExMiner v4.3 database were compared using the *t*-test. Survival curves were generated using the Kaplan–Meier plots. The pairwise comparison in the SBR and NPI criteria was performed using Dunnett’s Tukey–Kramer test. *P*-values < 0.05 were considered significant. The details can be found on the webpage of the databases used.

Abbreviations

BC: breast cancer; CLDNs: claudins; bc-GenExMiner: breast cancer gene expression miner; TCGA: The Cancer Genome Atlas; ER: estrogen receptor; HER2: human epidermal growth factor receptor 2; IHC: immunohistochemistry; PR: progesterone receptor; TNBC: triple-negative breast cancer; RFS: relapse-free survival; OS: overall survival; DMFS: distant metastasis-free survival; PPS: post-progression survival; SBR: Scarff–Bloom–Richardson; NPI: Nottingham

Prognostic Index; EMT: epithelial-to-mesenchymal transition.

AUTHOR CONTRIBUTIONS

Conceptualization: Y.G.D. and J.L.M.; methodology: Y.G.D. and J.L.M.; investigation: Y.G.D., J.L.M., and C.Q.Y.; writing–original draft: Y.G.D. and C.Q.Y.; writing–review and editing: J.L.M.; visualization: Y.G.D., J.L.M., and C.Q.Y.; supervision: J.L.M.; funding acquisition: Y.G.D.

CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

FUNDING

This study was supported by Guangzhou Zengcheng District Science and Technology Innovation fund project (ZCKJ2019-008).

REFERENCES

1. DeSantis CE, Ma J, Gaudet MM, Newman LA, Miller KD, Goding Sauer A, Jemal A, Siegel RL. Breast cancer statistics, 2019. *CA Cancer J Clin.* 2019; 69:438–51.
<https://doi.org/10.3322/caac.21583> PMID:31577379
2. Onitilo AA, Engel JM, Greenlee RT, Mukesh BN. Breast cancer subtypes based on ER/PR and Her2 expression: comparison of clinicopathologic features and survival. *Clin Med Res.* 2009; 7:4–13.
<https://doi.org/10.3121/cmr.2009.825> PMID:19574486
3. Semiglazov VF, Semiglazov VV, Dashyan GA, Ziltsova EK, Ivanov VG, Bozhok AA, Melnikova OA, Paltuev RM, Kletzel A, Berstein LM. Phase 2 randomized trial of primary endocrine therapy versus chemotherapy in postmenopausal patients with estrogen receptor-positive breast cancer. *Cancer.* 2007; 110:244–54.
<https://doi.org/10.1002/cncr.22789> PMID:17538978
4. Barton MK. Exemestane is effective for the chemoprevention of breast cancer. *CA Cancer J Clin.* 2011; 61:363–64.
<https://doi.org/10.3322/caac.20131> PMID:21898372
5. O’Sullivan CC, Bradbury I, Campbell C, Spielmann M, Perez EA, Joensuu H, Costantino JP, Delaloge S, Rastogi P, Zardavas D, Ballman KV, Holmes E, de Azambuja E, et al. Efficacy of adjuvant trastuzumab for patients with human epidermal growth factor receptor 2-positive early breast cancer and tumors ≤ 2 cm: a meta-analysis of the randomized trastuzumab trials. *J Clin Oncol.* 2015; 33:2600–08.
<https://doi.org/10.1200/JCO.2015.60.8620> PMID:26101239
6. Herschkowitz JI, Simin K, Weigman VJ, Mikaelian I, Usary J, Hu Z, Rasmussen KE, Jones LP, Assefnia S, Chandrasekharan S, Backlund MG, Yin Y, Khramtsov AI, et al. Identification of conserved gene expression features between murine mammary carcinoma models and human breast tumors. *Genome Biol.* 2007; 8:R76.
<https://doi.org/10.1186/gb-2007-8-5-r76> PMID:17493263
7. Prat A, Parker JS, Karginova O, Fan C, Livasy C, Herschkowitz JI, He X, Perou CM. Phenotypic and molecular characterization of the claudin-low intrinsic subtype of breast cancer. *Breast Cancer Res.* 2010; 12:R68.
<https://doi.org/10.1186/bcr2635> PMID:20813035
8. Taube JH, Herschkowitz JI, Komurov K, Zhou AY, Gupta S, Yang J, Hartwell K, Onder TT, Gupta PB, Evans KW, Hollier BG, Ram PT, Lander ES, et al. Core epithelial-to-mesenchymal transition interactome gene-expression signature is associated with claudin-low and metaplastic breast cancer subtypes. *Proc Natl Acad Sci USA.* 2010; 107:15449–54.
<https://doi.org/10.1073/pnas.1004900107> PMID:20713713
9. Hennessy BT, Gonzalez-Angulo AM, Stemke-Hale K, Gilcrease MZ, Krishnamurthy S, Lee JS, Fridlyand J, Sahin A, Agarwal R, Joy C, Liu W, Stivers D, Baggerly K, et al. Characterization of a naturally occurring breast cancer subset enriched in epithelial-to-mesenchymal transition and stem cell characteristics. *Cancer Res.* 2009; 69:4116–24.
<https://doi.org/10.1158/0008-5472.CAN-08-3441> PMID:19435916
10. Dias K, Dvorkin-Gheva A, Hallett RM, Wu Y, Hassell J, Pond GR, Levine M, Whelan T, Bane AL. Claudin-low breast cancer; clinical and pathological characteristics. *PLoS One.* 2017; 12:e0168669.
<https://doi.org/10.1371/journal.pone.0168669> PMID:28045912
11. Lal-Nag M, Morin PJ. The claudins. *Genome Biol.* 2009; 10:235.
<https://doi.org/10.1186/gb-2009-10-8-235> PMID:19706201
12. Tsukita S, Tanaka H, Tamura A. The claudins: from tight junctions to biological systems. *Trends Biochem Sci.* 2019; 44:141–52.
<https://doi.org/10.1016/j.tibs.2018.09.008> PMID:30665499

13. Morin PJ. Claudin proteins in human cancer: promising new targets for diagnosis and therapy. *Cancer Res.* 2005; 65:9603–06.
<https://doi.org/10.1158/0008-5472.CAN-05-2782>
PMID:[16266975](https://pubmed.ncbi.nlm.nih.gov/16266975/)
14. Tokés AM, Kulka J, Paku S, Szik A, Páska C, Novák PK, Szilák L, Kiss A, Bögi K, Schaff Z. Claudin-1, -3 and -4 proteins and mRNA expression in benign and Malignant breast lesions: a research study. *Breast Cancer Res.* 2005; 7:R296–305.
<https://doi.org/10.1186/bcr983> PMID:[15743508](https://pubmed.ncbi.nlm.nih.gov/15743508/)
15. Morohashi S, Kusumi T, Sato F, Odagiri H, Chiba H, Yoshihara S, Hakamada K, Sasaki M, Kijima H. Decreased expression of claudin-1 correlates with recurrence status in breast cancer. *Int J Mol Med.* 2007; 20:139–43.
PMID:[17611630](https://pubmed.ncbi.nlm.nih.gov/17611630/)
16. Ma F, Ding X, Fan Y, Ying J, Zheng S, Lu N, Xu B. A CLDN1-negative phenotype predicts poor prognosis in triple-negative breast cancer. *PLoS One.* 2014; 9:e112765.
<https://doi.org/10.1371/journal.pone.0112765>
PMID:[25393310](https://pubmed.ncbi.nlm.nih.gov/25393310/)
17. Blanchard AA, Ma X, Dueck KJ, Penner C, Cooper SC, Mulhall D, Murphy LC, Leygue E, Myal Y. Claudin 1 expression in basal-like breast cancer is related to patient age. *BMC Cancer.* 2013; 13:268.
<https://doi.org/10.1186/1471-2407-13-268>
PMID:[23721519](https://pubmed.ncbi.nlm.nih.gov/23721519/)
18. Zhao X, Zou Y, Gu Q, Zhao G, Gray H, Pfeffer LM, Yue J. Lentiviral vector mediated Claudin1 silencing inhibits epithelial to mesenchymal transition in breast cancer cells. *Viruses.* 2015; 7:2965–79.
<https://doi.org/10.3390/v7062755> PMID:[26067567](https://pubmed.ncbi.nlm.nih.gov/26067567/)
19. Kim TH, Huh JH, Lee S, Kang H, Kim GI, An HJ. Down-regulation of claudin-2 in breast carcinomas is associated with advanced disease. *Histopathology.* 2008; 53:48–55.
<https://doi.org/10.1111/j.1365-2559.2008.03052.x>
PMID:[18479414](https://pubmed.ncbi.nlm.nih.gov/18479414/)
20. Kimbung S, Kovács A, Bendahl PO, Malmström P, Fernö M, Hatschek T, Hedenfalk I. Claudin-2 is an independent negative prognostic factor in breast cancer and specifically predicts early liver recurrences. *Mol Oncol.* 2014; 8:119–28.
<https://doi.org/10.1016/j.molonc.2013.10.002>
PMID:[24287398](https://pubmed.ncbi.nlm.nih.gov/24287398/)
21. Jia H, Chai X, Li S, Wu D, Fan Z. Identification of claudin-2, -6, -11 and -14 as prognostic markers in human breast carcinoma. *Int J Clin Exp Pathol.* 2019; 12:2195–204.
PMID:[31934042](https://pubmed.ncbi.nlm.nih.gov/31934042/)
22. Hewitt KJ, Agarwal R, Morin PJ. The claudin gene family: expression in normal and neoplastic tissues. *BMC Cancer.* 2006; 6:186.
<https://doi.org/10.1186/1471-2407-6-186>
PMID:[16836752](https://pubmed.ncbi.nlm.nih.gov/16836752/)
23. Jääskeläinen A, Soini Y, Jukkola-Vuorinen A, Auvinen P, Haapasaari KM, Karihtala P. High-level cytoplasmic claudin 3 expression is an independent predictor of poor survival in triple-negative breast cancer. *BMC Cancer.* 2018; 18:223.
<https://doi.org/10.1186/s12885-018-4141-z>
PMID:[29482498](https://pubmed.ncbi.nlm.nih.gov/29482498/)
24. Kulka J, Szász AM, Németh Z, Madaras L, Schaff Z, Molnár IA, Tokés AM. Expression of tight junction protein claudin-4 in basal-like breast carcinomas. *Pathol Oncol Res.* 2009; 15:59–64.
<https://doi.org/10.1007/s12253-008-9089-x>
PMID:[18752049](https://pubmed.ncbi.nlm.nih.gov/18752049/)
25. Lanigan F, McKiernan E, Brennan DJ, Hegarty S, Millikan RC, McBryan J, Jirstrom K, Landberg G, Martin F, Duffy MJ, Gallagher WM. Increased claudin-4 expression is associated with poor prognosis and high tumour grade in breast cancer. *Int J Cancer.* 2009; 124:2088–97.
<https://doi.org/10.1002/ijc.24159> PMID:[19142967](https://pubmed.ncbi.nlm.nih.gov/19142967/)
26. Duarte GM, Almeida NR, Tocchet F, Espinola J, Barreto CT, Pinto GA, Soares FA, Marshall P, Russano de Paiva Silva G. Claudin-4 expression is associated with disease-free survival in breast carcinoma-in-situ: mean follow-up of 8.2 years. *Clin Breast Cancer.* 2018; 18:e1111–16.
<https://doi.org/10.1016/j.clbc.2018.06.005>
PMID:[30025999](https://pubmed.ncbi.nlm.nih.gov/30025999/)
27. Sugimoto H, Nagahara M, Bae Y, Nakagawa T, Ishikawa T, Sato T, Uetake H, Eishi Y, Sugihara K. Clinicopathologic relevance of claudin 5 expression in breast cancer. *Am J Clin Pathol.* 2015; 143:540–46.
<https://doi.org/10.1309/AJCPWGBZ6D00AIV>
PMID:[25780006](https://pubmed.ncbi.nlm.nih.gov/25780006/)
28. Xu X, Jin H, Liu Y, Liu L, Wu Q, Guo Y, Yu L, Liu Z, Zhang T, Zhang X, Dong X, Quan C. The expression patterns and correlations of claudin-6, methy-CpG binding protein 2, DNA methyltransferase 1, histone deacetylase 1, acetyl-histone H3 and acetyl-histone H4 and their clinicopathological significance in breast invasive ductal carcinomas. *Diagn Pathol.* 2012; 7:33.
<https://doi.org/10.1186/1746-1596-7-33>
PMID:[22455563](https://pubmed.ncbi.nlm.nih.gov/22455563/)
29. Yang M, Li Y, Shen X, Ruan Y, Lu Y, Jin X, Song P, Guo Y, Zhang X, Qu H, Shao Y, Quan C. CLDN6 promotes chemoresistance through GSTP1 in human breast cancer. *J Exp Clin Cancer Res.* 2017; 36:157.
<https://doi.org/10.1186/s13046-017-0627-9>
PMID:[29116019](https://pubmed.ncbi.nlm.nih.gov/29116019/)

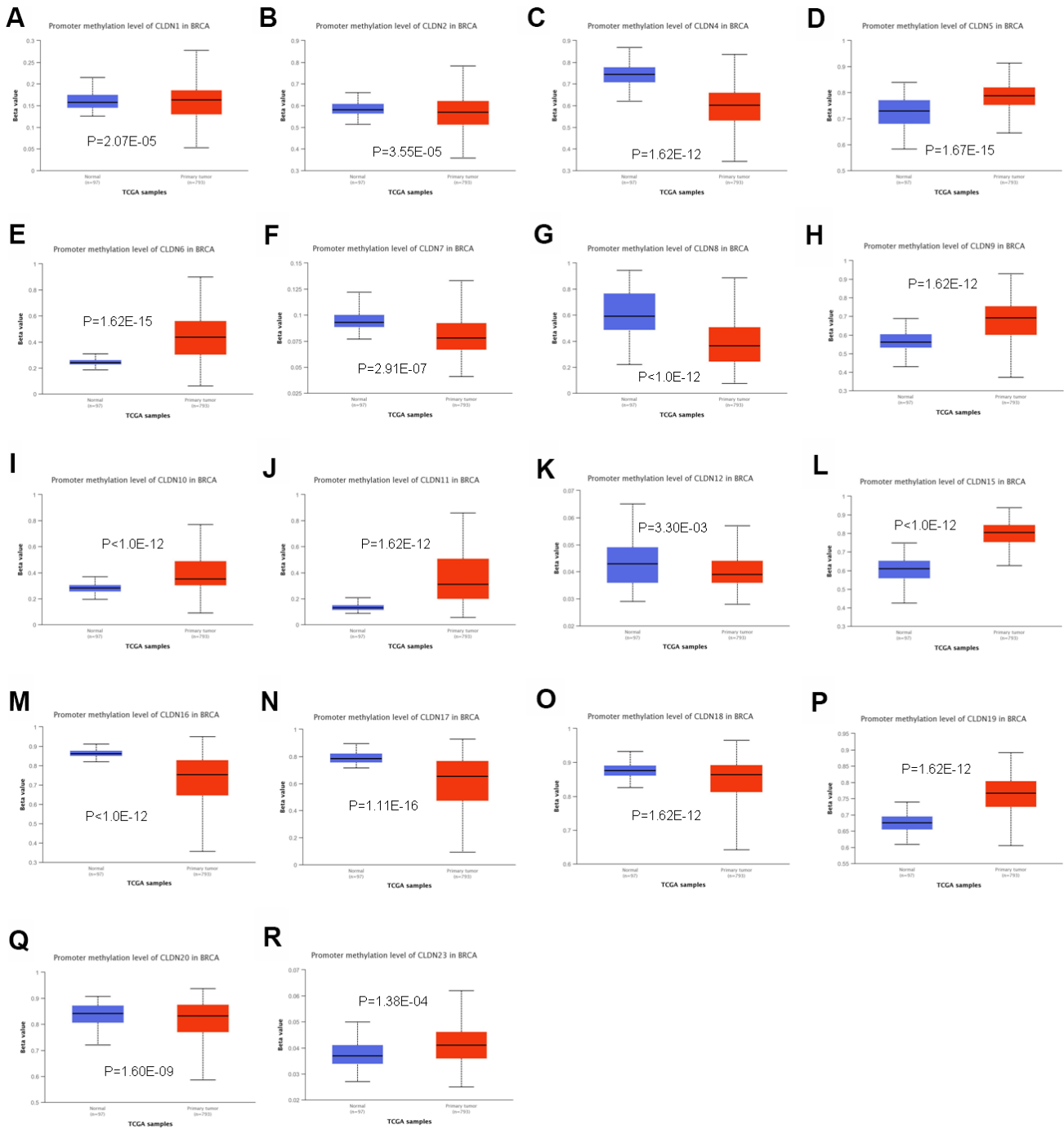
30. Yang M, Li Y, Ruan Y, Lu Y, Lin D, Xie Y, Dong B, Dang Q, Quan C. CLDN6 enhances chemoresistance to ADM via AF-6/ERKs pathway in TNBC cell line MDAMB231. *Mol Cell Biochem.* 2018; 443:169–80.
<https://doi.org/10.1007/s11010-017-3221-8>
PMID:[29159771](https://pubmed.ncbi.nlm.nih.gov/29159771/)
31. Iravani O, Yip GW, Thike AA, Chua PJ, Jane Scully O, Tan PH, Bay BH. Prognostic significance of claudin 12 in estrogen receptor-negative breast cancer. *J Clin Pathol.* 2016; 69:878–83.
<https://doi.org/10.1136/jclinpath-2015-203265>
PMID:[26926102](https://pubmed.ncbi.nlm.nih.gov/26926102/)
32. Kuo SJ, Chien SY, Lin C, Chan SE, Tsai HT, Chen DR. Significant elevation of CLDN16 and HAPLN3 gene expression in human breast cancer. *Oncol Rep.* 2010; 24:759–66.
<https://doi.org/10.3892/or.00000918> PMID:[20664984](https://pubmed.ncbi.nlm.nih.gov/20664984/)
33. Martin TA, Harrison GM, Watkins G, Jiang WG. Claudin-16 reduces the aggressive behavior of human breast cancer cells. *J Cell Biochem.* 2008; 105:41–52.
<https://doi.org/10.1002/jcb.21797> PMID:[18442037](https://pubmed.ncbi.nlm.nih.gov/18442037/)
34. Sealfon SC, Chu TT. RNA and DNA microarrays. *Methods Mol Biol.* 2011; 671:3–34.
https://doi.org/10.1007/978-1-59745-551-0_1
PMID:[20967621](https://pubmed.ncbi.nlm.nih.gov/20967621/)
35. Ma XJ, Dahiya S, Richardson E, Erlander M, Sgroi DC. Gene expression profiling of the tumor microenvironment during breast cancer progression. *Breast Cancer Res.* 2009; 11:R7.
<https://doi.org/10.1186/bcr2222> PMID:[19187537](https://pubmed.ncbi.nlm.nih.gov/19187537/)
36. Curtis C, Shah SP, Chin SF, Turashvili G, Rueda OM, Dunning MJ, Speed D, Lynch AG, Samarajiwa S, Yuan Y, Gräf S, Ha G, Haffari G, et al, and METABRIC Group. The genomic and transcriptomic architecture of 2,000 breast tumours reveals novel subgroups. *Nature.* 2012; 486:346–52.
<https://doi.org/10.1038/nature10983> PMID:[22522925](https://pubmed.ncbi.nlm.nih.gov/22522925/)
37. Radvanyi L, Singh-Sandhu D, Gallichan S, Lovitt C, Pedyczak A, Mallo G, Gish K, Kwok K, Hanna W, Zubovits J, Armes J, Venter D, Hakimi J, et al. The gene associated with trichorhinophalangeal syndrome in humans is overexpressed in breast cancer. *Proc Natl Acad Sci USA.* 2005; 102:11005–10.
<https://doi.org/10.1073/pnas.0500904102>
PMID:[16043716](https://pubmed.ncbi.nlm.nih.gov/16043716/)
38. Turashvili G, Bouchal J, Baumforth K, Wei W, Dziechciarkova M, Ehrmann J, Klein J, Fridman E, Skarda J, Srovnal J, Hajdich M, Murray P, Kolar Z. Novel markers for differentiation of lobular and ductal invasive breast carcinomas by laser microdissection and microarray analysis. *BMC Cancer.* 2007; 7:55.
<https://doi.org/10.1186/1471-2407-7-55>
PMID:[17389037](https://pubmed.ncbi.nlm.nih.gov/17389037/)
39. Perou CM, Sørlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, Fluge O, Pergamenschikov A, Williams C, et al. Molecular portraits of human breast tumours. *Nature.* 2000; 406:747–52.
<https://doi.org/10.1038/35021093> PMID:[10963602](https://pubmed.ncbi.nlm.nih.gov/10963602/)
40. Finak G, Bertos N, Pepin F, Sadekova S, Souleimanova M, Zhao H, Chen H, Omeroglu G, Meterissian S, Omeroglu A, Hallett M, Park M. Stromal gene expression predicts clinical outcome in breast cancer. *Nat Med.* 2008; 14:518–27.
<https://doi.org/10.1038/nm1764> PMID:[18438415](https://pubmed.ncbi.nlm.nih.gov/18438415/)
41. Glück S, Ross JS, Royce M, McKenna EF Jr, Perou CM, Avisar E, Wu L. TP53 genomics predict higher clinical and pathologic tumor response in operable early-stage breast cancer treated with docetaxel-capecitabine ± trastuzumab. *Breast Cancer Res Treat.* 2012; 132:781–91.
<https://doi.org/10.1007/s10549-011-1412-7>
PMID:[21373875](https://pubmed.ncbi.nlm.nih.gov/21373875/)
42. Richardson AL, Wang ZC, De Nicolo A, Lu X, Brown M, Miron A, Liao X, Iglehart JD, Livingston DM, Ganesan S. X chromosomal abnormalities in basal-like human breast cancer. *Cancer Cell.* 2006; 9:121–32.
<https://doi.org/10.1016/j.ccr.2006.01.013>
PMID:[16473279](https://pubmed.ncbi.nlm.nih.gov/16473279/)
43. Sørlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A, Deng S, Johnsen H, Pesich R, Geisler S, Demeter J, Perou CM, Lønning PE, et al. Repeated observation of breast tumor subtypes in independent gene expression data sets. *Proc Natl Acad Sci USA.* 2003; 100:8418–23.
<https://doi.org/10.1073/pnas.0932692100>
PMID:[12829800](https://pubmed.ncbi.nlm.nih.gov/12829800/)
44. Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, Richardson AL, Polyak K, Tubo R, Weinberg RA. Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature.* 2007; 449:557–63.
<https://doi.org/10.1038/nature06188> PMID:[17914389](https://pubmed.ncbi.nlm.nih.gov/17914389/)
45. Sørlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, van de Rijn M, Jeffrey SS, Thorsen T, Quist H, Matese JC, et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci USA.* 2001; 98:10869–74.
<https://doi.org/10.1073/pnas.191367098>
PMID:[11553815](https://pubmed.ncbi.nlm.nih.gov/11553815/)
46. Zhao H, Langerød A, Ji Y, Nowels KW, Nesland JM, Tibshirani R, Bukholm IK, Kåresen R, Botstein D,

- Børresen-Dale AL, Jeffrey SS. Different gene expression patterns in invasive lobular and ductal carcinomas of the breast. *Mol Biol Cell*. 2004; 15:2523–36.
<https://doi.org/10.1091/mbc.e03-11-0786>
PMID:15034139
47. Jiang L, Yang YD, Fu L, Xu W, Liu D, Liang Q, Zhang X, Xu L, Guan XY, Wu B, Sung JJ, Yu J. CLDN3 inhibits cancer aggressiveness via Wnt-EMT signaling and is a potential prognostic biomarker for hepatocellular carcinoma. *Oncotarget*. 2014; 5:7663–76.
<https://doi.org/10.18632/oncotarget.2288>
PMID:25277196
48. Che J, Yue D, Zhang B, Zhang H, Huo Y, Gao L, Zhen H, Yang Y, Cao B. Claudin-3 inhibits lung squamous cell carcinoma cell epithelial-mesenchymal transition and invasion via suppression of the Wnt/ β -catenin signaling pathway. *Int J Med Sci*. 2018; 15:339–51.
<https://doi.org/10.7150/ijms.22927> PMID:29511369
49. Lin X, Shang X, Manorek G, Howell SB. Regulation of the epithelial-mesenchymal transition by claudin-3 and claudin-4. *PLoS One*. 2013; 8:e67496.
<https://doi.org/10.1371/journal.pone.0067496>
PMID:23805314
50. Shang X, Lin X, Alvarez E, Manorek G, Howell SB. Tight junction proteins claudin-3 and claudin-4 control tumor growth and metastases. *Neoplasia*. 2012; 14:974–85.
<https://doi.org/10.1593/neo.12942> PMID:23097631
51. Ahmad R, Kumar B, Chen Z, Chen X, Müller D, Lele SM, Washington MK, Batra SK, Dhawan P, Singh AB. Loss of claudin-3 expression induces IL6/gp130/Stat3 signaling to promote colon cancer Malignancy by hyperactivating Wnt/ β -catenin signaling. *Oncogene*. 2017; 36:6592–604.
<https://doi.org/10.1038/onc.2017.259> PMID:28783170
52. Ma SC, Li Q, Peng JY, Zhouwen JL, Diao JF, Niu JX, Wang X, Guan XD, Jia W, Jiang WG. Claudin-5 regulates blood-brain barrier permeability by modifying brain microvascular endothelial cell proliferation, migration, and adhesion to prevent lung cancer metastasis. *CNS Neurosci Ther*. 2017; 23:947–60.
<https://doi.org/10.1111/cns.12764> PMID:28961379
53. Escudero-Esparza A, Jiang WG, Martin TA. Claudin-5 participates in the regulation of endothelial cell motility. *Mol Cell Biochem*. 2012; 362:71–85.
<https://doi.org/10.1007/s11010-011-1129-2>
PMID:22038628
54. Guo J, Cai H, Zheng J, Liu X, Liu Y, Ma J, Que Z, Gong W, Gao Y, Tao W, Xue Y. Long non-coding RNA NEAT1 regulates permeability of the blood-tumor barrier via miR-181d-5p-mediated expression changes in ZO-1, occludin, and claudin-5. *Biochim Biophys Acta Mol Basis Dis*. 2017; 1863:2240–54.
<https://doi.org/10.1016/j.bbadis.2017.02.005>
PMID:28185956
55. Zhao L, Wang P, Liu Y, Ma J, Xue Y. miR-34c regulates the permeability of blood-tumor barrier via MAZ-mediated expression changes of ZO-1, occludin, and claudin-5. *J Cell Physiol*. 2015; 230:716–31.
<https://doi.org/10.1002/jcp.24799> PMID:25201524
56. Li HP, Peng CC, Wu CC, Chen CH, Shih MJ, Huang MY, Lai YR, Chen YL, Chen TW, Tang P, Chang YS, Chang KP, Hsu CL. Inactivation of the tight junction gene CLDN11 by aberrant hypermethylation modulates tubulin polymerization and promotes cell migration in nasopharyngeal carcinoma. *J Exp Clin Cancer Res*. 2018; 37:102.
<https://doi.org/10.1186/s13046-018-0754-y>
PMID:29747653
57. Li J, Zhou C, Ni S, Wang S, Ni C, Yang P, Ye M. Methylated claudin-11 associated with metastasis and poor survival of colorectal cancer. *Oncotarget*. 2017; 8:96249–62.
<https://doi.org/10.18632/oncotarget.21997>
PMID:29221203
58. Shen Z, Cao B, Lin L, Zhou C, Ye D, Qiu S, Li Q, Cui X. The clinical signification of claudin-11 promoter hypermethylation for laryngeal squamous cell carcinoma. *Med Sci Monit*. 2017; 23:3635–40.
<https://doi.org/10.12659/msm.904751>
PMID:28743857
59. Yang J, Liu X, Yuan X, Wang Z. miR-99b promotes metastasis of hepatocellular carcinoma through inhibition of claudin 11 expression and may serve as a prognostic marker. *Oncol Rep*. 2015; 34:1415–23.
<https://doi.org/10.3892/or.2015.4104> PMID:26134929
60. Yang P, Zhang M, Liu X, Pu H. MicroRNA-421 promotes the proliferation and metastasis of gastric cancer cells by targeting claudin-11. *Exp Ther Med*. 2017; 14:2625–32.
<https://doi.org/10.3892/etm.2017.4798>
PMID:28962204
61. Rhodes DR, Yu J, Shanker K, Deshpande N, Varambally R, Ghosh D, Barrette T, Pandey A, Chinnaiyan AM. ONCOMINE: a cancer microarray database and integrated data-mining platform. *Neoplasia*. 2004; 6:1–6.
[https://doi.org/10.1016/s1476-5586\(04\)80047-2](https://doi.org/10.1016/s1476-5586(04)80047-2)
PMID:15068665
62. Chandrashekar DS, Bashel B, Balasubramanya SA, Creighton CJ, Ponce-Rodriguez I, Chakravarthi BV, Varambally S. UALCAN: a portal for facilitating tumor

- subgroup gene expression and survival analyses. *Neoplasia*. 2017; 19:649–58.
<https://doi.org/10.1016/j.neo.2017.05.002>
PMID:[28732212](https://pubmed.ncbi.nlm.nih.gov/28732212/)
63. Jézéquel P, Campone M, Gouraud W, Guérin-Charbonnel C, Leux C, Ricolleau G, Campion L. bc-GenExMiner: an easy-to-use online platform for gene prognostic analyses in breast cancer. *Breast Cancer Res Treat*. 2012; 131:765–75.
<https://doi.org/10.1007/s10549-011-1457-7>
PMID:[21452023](https://pubmed.ncbi.nlm.nih.gov/21452023/)
64. Györfy B, Lanczky A, Eklund AC, Denkert C, Budczies J, Li Q, Szallasi Z. An online survival analysis tool to rapidly assess the effect of 22,277 genes on breast cancer prognosis using microarray data of 1,809 patients. *Breast Cancer Res Treat*. 2010; 123:725–31.
<https://doi.org/10.1007/s10549-009-0674-9>
PMID:[20020197](https://pubmed.ncbi.nlm.nih.gov/20020197/)
65. Cancer Genome Atlas Network. Comprehensive molecular portraits of human breast tumours. *Nature*. 2012; 490:61–70.
<https://doi.org/10.1038/nature11412> PMID:[23000897](https://pubmed.ncbi.nlm.nih.gov/23000897/)
66. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, Sun Y, Jacobsen A, Sinha R, Larsson E, Cerami E, Sander C, Schultz N. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal*. 2013; 6:pl1.
<https://doi.org/10.1126/scisignal.2004088>
PMID:[23550210](https://pubmed.ncbi.nlm.nih.gov/23550210/)

SUPPLEMENTARY MATERIALS

Supplementary Figure



Supplementary Figure 1. Significant changes in claudin promoter methylation levels between breast cancer and normal tissues (UALCAN database). (A) CLDN1; (B) CLDN2; (C) CLDN4; (D) CLDN5; (E) CLDN6; (F) CLDN7; (G) CLDN8; (H) CLDN9; (I) CLDN10; (J) CLDN11; (K) CLDN12; (L) CLDN15; (M) CLDN16; (N) CLDN17; (O) CLDN18; (P) CLDN19; (Q) CLDN20; (R) CLDN23. The beta value indicates the level of DNA methylation ranging from 0 (unmethylated) to 1 (fully methylated). Different beta cut-off values have been considered to indicate hypermethylation (beta value: 0.7–0.5) or hypomethylation (beta-value: 0.3–0.25). A p-value < 0.05 was considered significant.

Supplementary Tables

Please browse Full Text version to see the data of Supplementary Tables 2–4, 6–9.

Supplementary Table 1. Expression and functions of claudin in breast cancer.

Genes	Types of cancer	Expression level	Function role	References
CLDN1	breast cancer vs normal	down	NA	[14]
	recurrent group BC vs non-recurrent group	down	correlated with disease-free interval and lymph node metastasis	[15]
	TNBC	NA	associated with worse relapse-free survival (RFS) and overall survival	[16]
	basal-like breast cancer	up	involved in epithelial-mesenchymal-transition (EMT)	[17]
	breast cancer cells	NA	silencing CLDN1 inhibited epithelial to mesenchymal transition (EMT)	[18]
CLDN2	TNBC	NA	associated with the risk of recurrence in the lymph node positive subgroup	[16]
	breast cancer vs normal	down	associated with lymph node metastasis	[19]
	BC liver metastases compared to other metastatic sites	up	associated with metastasis-free interval	[20]
CLDN3	breast cancer vs normal	down	NA	[21]
	breast cancer vs normal	no difference	NA	[14]
	breast cancer	up	NA	[22]
CLDN4	TNBC	strong cytoplasmic claudin 3 expression	associated with survival.	[23]
	breast cancer vs normal	no difference	NA	[14]
	recurrent group BC vs non-recurrent group	no significance	NA	[15]
	TNBC	NA	not associated with survival.	[16]
	breast cancer	up	NA	[22]
	basal-like	up	NA	[22]
	group BC as compared to not basal-like	up	NA	[24]
	breast cancer	NA	correlated with tumour grade, Her2, ER and survival	[25]
CLDN5	breast carcinoma in situ	up	had a worse prognosis in carcinoma in situ	[26]
	recurrent breast cancer	up	with lower relapse-free survival (RFS)	[27]
CLDN6	breast invasive ductal carcinomas	down	correlated with lymph node metastasis	[28]
	MCF-7/MDR cells	NA	High expression of CLDN6 confers chemoresistance on breast cancer	[29]
	TNBC cell line MDAMB231	NA	CLDN6 enhances chemoresistance to ADM	[30]
CLDN7	breast cancer vs normal	down	NA	[21]
	TNBC	NA	not associated with survival.	[16]
	breast cancer	up	NA	[22]
CLDN11	TNBC	strong cytoplasmic claudin 7 expression	associated with survival	[23]
	breast cancer vs normal	up	NA	[21]
CLDN12	estrogen receptor (ER)-negative breast cancer	NA	associated with survival.	[31]
CLDN16	breast cancer vs normal	up	no significant association with overall survival	[32]
	node positive tumors compared to negative	down	associated with aggressive phenotype	[33]
	breast cancer vs normal	down	NA	[21]

Note: “up” means high expression; “down” means low expression; BC; breast cancer; TNBC, triple negative breast cancer; NA, not available.

Supplementary Table 2. Datasets of the claudin family in breast cancer (ONCOMINE database).

Supplementary Table 3. The expression of the claudin family of breast cancer (UALCAN database).

Supplementary Table 4. Relationship between the claudin family and the clinicopathologic parameters of breast cancer (bc-GenExMiner v4.3).

Supplementary Table 5. Results of Dunnett's Tukey-Kramer test for pairwise comparison in Scarff bloom and Richardson and Nottingham prognostic index criteria.

Gene	Pairwise comparison of SBR	P-value	Pairwise comparison of NPI	P-value
CLDN1	No significant	0.0553	NPI2 > NPI1	< 0.01
			NPI3 = NPI1	> 0.10
			NPI3 = NPI2	> 0.10
CLDN2	No significant	0.568	No significant	0.2713
CLDN3	SBR2 > SBR1	<0.0001	NPI2 > NPI1	<0.0001
	SBR3 > SBR1	<0.0001	NPI3 > NPI1	<0.0001
	SBR3 > SBR2	<0.0001	NPI3 > NPI2	<0.0001
CLDN4	SBR3 > SBR1	<0.0001	NPI2 > NPI1	<0.0001
	SBR3 > SBR2	<0.0001	NPI3 > NPI1	<0.0001
	SBR2 > SBR1	<0.0001	NPI3 = NPI2	>0.1
CLDN5	SBR2 < SBR1	<0.0001	NPI2 < NPI1	<0.0001
	SBR3 < SBR1	<0.0001	NPI3 < NPI1	<0.0001
	SBR3 < SBR2	<0.0001	NPI3 = NPI2	>0.1
CLDN6	SBR3 > SBR1	<0.01	NPI2 > NPI1	<0.05
	SBR3 > SBR2	<0.01	NPI3 = NPI1	>0.1
	SBR2 = SBR1	>0.1	NPI3 = NPI2	>0.1
CLDN7	SBR2 > SBR1	<0.05	NPI2 > NPI1	<0.05
	SBR3 < SBR2	<0.1	NPI3 > NPI1	<0.05
	SBR3 = SBR1	>0.1	NPI3 = NPI2	>0.1
CLDN8	No significant	0.3213	No significant	0.1274
CLDN9	SBR3 > SBR1	<0.0001	NPI2 > NPI1	<0.0001
	SBR3 > SBR2	<0.0001	NPI3 > NPI1	<0.0001
	SBR2 = SBR1	>0.1	NPI3 > NPI2	<0.01
CLDN10	SBR3 > SBR1	<0.0001	NPI2 > NPI1	<0.0001
	SBR3 > SBR2	<0.0001	NPI3 > NPI1	<0.001
	SBR2 = SBR1	>0.1	NPI3 = NPI2	>0.1
CLDN11	SBR2 < SBR1	<0.0001	NPI2 < NPI1	<0.0001
	SBR3 < SBR1	<0.0001	NPI3 < NPI1	<0.01
	SBR3 < SBR2	<0.0001	NPI3 = NPI2	>0.1
CLDN12	SBR3 < SBR1	<0.0001	NPI2 < NPI1	<0.01
	SBR3 < SBR2	<0.0001	NPI3 < NPI1	<0.01
	SBR2 < SBR1	<0.1	NPI3 = NPI2	>0.1
CLDN14	SBR3 > SBR1	<0.01	No significant	0.1355
	SBR3 > SBR2	<0.05		
	SBR2 = SBR1	>0.1		
CLDN15	SBR3 < SBR1	<0.05	No significant	0.0732
	SBR3 < SBR2	<0.05		
	SBR2 = SBR1	>0.1		
CLDN16	SBR3 > SBR2	<0.0001	No significant	0.6659

	SBR3 > SBR1	<0.05		
	SBR2 = SBR1	>0.1		
CLDN17	SBR3 > SBR2	<0.05	No significant	0.6285
	SBR2 = SBR1	>0.1		
	SBR3 = SBR1	>0.1		
CLDN18	No significant	0.5717	No significant	0.9423
CLDN19	No significant	0.5957	No significant	0.1165
CLDN20	No significant	0.0907	No significant	0.8386
CLDN22	No significant	0.8599	No significant	0.7234
CLDN23	SBR3 > SBR1	<0.0001	NPI2 > NPI1	<0.0001
	SBR3 > SBR2	<0.0001	NPI3 > NPI1	<0.0001
	SBR2 = SBR1	>0.1	NPI3 = NPI2	>0.01
CLDN24	SBR2 > SBR1	<0.1	No significant	0.7341
	SBR3 > SBR1	<0.1		
	SBR3 = SBR2	>0.1		

Supplementary Table 6. Survival analyses of the claudin family in all patients with breast cancer (Kaplan–Meier plotter).

Supplementary Table 7. Survival analyses of the claudin family with different molecular subtypes in breast cancer (Kaplan–Meier plotter).

Supplementary Table 8. Survival analyses of the claudin family with different lymph node status in breast cancer (Kaplan–Meier plotter).

Supplementary Table 9. Survival analyses of the claudin family with different histologic grades in breast cancer (Kaplan–Meier plotter).