

## Iron: an underrated factor in aging

Dennis Mangan<sup>1,&</sup>

<sup>1</sup>MTOR LLC, Bakersfield, CA 93311, USA

**Correspondence to:** Dennis Mangan; **email:** [pdmangan@outlook.com](mailto:pdmangan@outlook.com)

**Keywords:** iron, aging, oxidative stress, calorie restriction, plasma dilution

**Received:** September 12, 2021    **Accepted:** September 27, 2021    **Published:** October 6, 2021

**Copyright:** © 2021 Mangan. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/3.0/) (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

### ABSTRACT

**Iron is an essential element for virtually all living organisms, but its reactivity also makes it potentially harmful. Iron accumulates with aging, and is associated with many age-related diseases; it also shortens the lifespans of several model organisms. Blocking iron absorption through drugs or natural products extends lifespan. Many life-extending interventions, such as rapamycin, calorie restriction, and old plasma dilution can be explained by the effects they have on iron absorption, excretion, and metabolism. Control of body iron stores so that they remain in a low normal range may be an important, lifespan- and healthspan-extending intervention.**

All life forms require the element iron as a constituent of their biochemical systems, iron being used in producing ATP in mitochondria, in cytochromes and hemoglobin, and in many other uses. Iron is essential for organismal growth and maintenance, so all life, from bacteria and algae to mammals, have developed the means to collect and store iron from their environments; this centrality of iron for all life suggests that iron may be involved in aging. Most organisms, including humans, have no systematic means of ridding themselves of excess iron. Whether this lack of ways to dispose of excess iron came about due to a relative scarcity of iron, or because the detrimental results from excess iron were relatively rare in an environment in which few organisms died from natural aging, is a question that remains to be answered. Whatever the answer to that may be, most organisms accumulate iron as they age [1–3].

A problem that organisms face in the use of iron in biological systems is protecting cells from iron damage. The very property of iron that makes it useful, its ability to accept or donate electrons, also gives it the ability to damage molecules and organelles via the Fenton reaction, in which iron reacts with hydrogen peroxide, leading to the formation of the highly reactive and toxic free radical, hydroxyl.

Most iron in cells is bound to proteins and other molecules that safely store it and prevent it from interacting with other macromolecules. In mammals, ferritin and transferrin are such proteins; hemoglobin is, however, the quantitatively most important iron depot in mammals. In theory, these storage proteins should be enough to protect organelles and macromolecules from iron's reactivity, but in practice another process becomes perhaps more important, and that is iron dysregulation. Storage proteins such as ferritin can themselves be damaged, leading to "leakage" of free iron, which can then react with and damage cellular structures, which in turn can lead to organ damage and the deterioration associated with aging [4]. Damage to ferritin can be caused by glycation due to hyperglycemia, a phenomenon seen more widely with the development of advanced glycation end products (AGEs), and with the glycation of hemoglobin (HbA1c), elevated in diabetics. The superoxide anion can also damage ferritin, leading to a vicious cycle in which leakage of free iron leads to oxidative stress, in turn leading to more iron leakage [5].

Whether this damage associated with aging is in fact a cause or consequence of aging of course remains to be determined, but as we shall see, there are several other reasons to think that iron is a driver of aging.

## Activation of mTOR

The mechanistic (or mammalian) target of rapamycin (mTOR), a molecular sensor that integrates nutritional and stress signals with growth and energy needs of the cell, plays a crucial role in aging; inhibition of mTOR with the drug rapamycin extends lifespan and healthspan in laboratory animals [6]. Nutritional and other factors that promote mTOR activation promote accelerated aging, and their absence may retard aging. For example, calorie restriction, the most robust life extension paradigm, also inhibits mTOR activation, while in humans, obesity resulting from overnutrition is a well-known cause of early morbidity and mortality [7]. In general, growth factors, whether amino acids, glucose, or fatty acids, or hormonal signals that they engender, such as insulin, promote mTOR activation, while their absence, or the presence of stress from exercise or other sources that activate the cellular energy sensor AMPK, inactivate mTOR.

Iron is one such growth factor. Iron is required for the growth of the organism, and iron activates mTOR; iron chelators, chemicals that bind free iron, inhibit mTOR activity [8]. Iron deficiency downregulates mTOR activity [9]. These data fit the paradigm of increased mTOR activity and aging, which may be promoted by excess iron.

In turn, mTOR also exerts control over iron metabolism, and the inhibition of mTOR activity by rapamycin leads to inhibition of iron accumulation via the iron-regulating hormone hepcidin [10]. Transplant patients taking sirolimus (rapamycin) often develop a microcytic anemia, which has been linked to sirolimus-induced iron deficiency [11].

Excessive activation of mTOR is seen in type 2 diabetes, and this activation is associated with insulin resistance [12]. mTOR activation in diabetes may be responsible for the accumulation of excess iron seen in this illness; alternatively, accumulation of iron might activate mTOR, leading to diabetes. Increased iron stores predict the development of type 2 diabetes, while iron depletion can protect against it [13]. Insulin resistance is associated with inadequate levels of hepcidin, an iron regulatory hormone, which could be expected to increase body iron stores [14]. So there's evidence that iron increases insulin resistance, and that in turn can lead to higher body iron in a vicious cycle. Since type 2 diabetes is an age-related disease, it can be seen how excess iron promotes aging.

However, even when ferritin is in the normal range, depletion of iron improves glucose tolerance, insulin resistance, and markers of cardiovascular disease [5].

Iron appears to have a dose-response effect starting from near iron deficiency up to iron overloading, making it a candidate driver of aging.

## Blocking iron extends lifespan

In experimental organisms and animals, blocking iron extends lifespan.

In *Saccharomyces cerevisiae*, limitation of iron increases chronological lifespan via inducing autophagy [15]. Autophagy is essential for lifespan extension, so this may be the ultimate means by which iron restriction or depletion extends lifespan, and iron excess promotes aging [16].

Dietary tea extracts increase the lifespan of *Drosophila* by over 20% by blocking the absorption of iron [17].

A number of geroprotectors increase lifespan in model organisms, and many of these either block dietary iron absorption or chelate iron and remove it.

Curcumin and its metabolite tetrahydrocurcumin increase average lifespan in at least three model organisms: *C. elegans*, *Drosophila*, and mice [18]. Curcumin is a strong iron chelator; animals fed curcumin had a decline in liver ferritin [19]. Mice fed 0.2% curcumin in the diet become iron deficient; levels of zinc and copper were not affected [20].

Epigallocatechin gallate (EGCG), a compound found in green tea, extends lifespan of both *C. elegans* and *Drosophila* [21, 22].

EGCG extends the lifespan and healthspan of mice, attenuating markers of DNA damage and senescence-associated secretory profile, and increasing activation of autophagy [23]. EGCG also extends the lifespan of rats by reducing liver and kidney damage and inhibiting inflammation and oxidative stress [24].

EGCG is a strong chelator of iron [25]. EGCG protects against alcoholic liver disease in mice through decreasing the level of liver iron [26].

Aspirin extends the lifespan of *C. elegans* [27] and that of mice [28]. In humans, aspirin reduces the risk of cancer [29].

Aspirin use is associated with lower body iron stores, perhaps through an increase in gastrointestinal blood loss; observational studies have shown that regular aspirin users have lower serum ferritin; as cancer cells are notoriously iron-hungry, this might partially explain the reduced cancer risk with aspirin [30]. Aspirin also

recapitulates several features of calorie restriction, which could be expected to result in lower levels of body iron [31]. Salicylate, the main metabolite of aspirin, forms a complex with iron, and this process can be used in the quantitative detection of salicylate [32]. Bacteria elaborate siderophores in order to capture iron from their environment, and one such siderophore seen in several species of *Pseudomonas* is salicylate [33]. Aspirin increases the synthesis of ferritin in endothelial cells, which would result in lower levels of free iron, providing an antioxidant function; aspirin failed to promote ferritin synthesis in the presence of the iron chelator deferoxamine, indicating an interaction of aspirin and iron [34]. All of these data fit well with the idea that aspirin extends lifespan and inhibits cancer through decreasing body iron as at least one mechanism.

A screening of drugs for protection against neuronal glucose toxicity found six of them that reduce mortality rate in *C. elegans*: caffeine, tannic acid, ciclopirox, acetaminophen, bacitracin, and baicalein [35]. Of these, with the possible exception of caffeine, all chelate iron. Caffeine has weak iron-binding ability [36]. Tannic acid, ciclopirox, bacitracin, and baicalein are strong iron chelators [37–40]. Acetaminophen protects against iron-induced cardiac damage in gerbils [41]. Thus there is evidence that a primary mechanism of these life-extending compounds is the binding of free iron and protection against oxidant-induced damage.

Clofibrate increases lifespan in *C. elegans* [42]. When fed to Wistar rats, clofibrate led to a 50% decrease in serum iron and a reduction in transferrin mRNA [43].

Therefore, this is yet another example of a life-extending drug, the mechanism of which may at least partially involve decreased iron stores.

Berberine extends lifespan in mice [44]. Berberine also suppresses gero-conversion [45]. Berberine also has “a marked capacity” for iron-binding, and effectively chelates iron [46].

Acarbose extends lifespan in mice [47]. Acarbose increases fecal excretion of iron and has been known to be a cause of iron-deficiency anemia in humans [48, 49].

Doxycycline extends lifespan in *C. elegans* [50]. Doxycycline has a “strong iron-chelating activity” [51].

Enalapril is an angiotensin converting enzyme inhibitor that increases lifespan in rats [52]. One of the adverse effects of ACE inhibitors in humans is a dry cough, which is relieved by iron administration, indicating that one effect of these drugs concerns iron metabolism [53].

Ibuprofen extends lifespan in at least three organisms: *Saccharomyces cerevisiae*, *C. elegans*, and *Drosophila* [54]. Ibuprofen chelates iron and protects against oxidant lung injury by this means [55].

Metformin increases lifespan and healthspan in mice [56]. Many mechanisms have been proposed for the effects of metformin. One such mechanism is that, at concentrations seen in clinical use, metformin suppresses heme production in human erythrocytes, and prevents heme oxidation, thus having a role in regulating the redox status of iron [57]. In yeast, a global genetic screen showed that metformin induces a state similar to iron deficiency [58].

Quercetin, a polyphenol found in food, extends lifespan in *C. elegans*, and it appears to do so by increasing resistance to oxidative stress [59]. Quercetin is “a powerful chelating agent that can sequester iron(II) in such a way to prevent its involvement in the Fenton reaction [60].”

Thus, we can see that a large number of life-extending compounds also interact with iron, either by chelation, inhibition of absorption, or increased iron loss.

## Calorie restriction

Calorie restriction (CR) is the most robust life-extending intervention known. Many mechanisms have been proposed to explain lifespan extension by CR, such as its effects on insulin and IGF-1 signaling, mTOR, sirtuins, AMPK, adiposity, and resistance to oxidative stress [61]. CR also affects iron metabolism.

In yeast grown on a low-glucose medium, which is a model of CR, oxidative damage in the form of protein carbonylation is largely prevented. Intracellular iron concentrations changed little, whereas in yeast cells grown on non-restrictive media, iron concentrations increased up to 5-fold. The pro-oxidant effects of these increasing iron concentrations might explain the molecular damage seen in unrestricted cells, and the lower iron seen in CR yeast might explain the lower levels of damage. Thus, lower levels of iron in CR yeast can be posited as an important mechanism of increased longevity in CR [62].

CR could also be expected to result in lower ingestion of iron. When iron is the only limiting nutrient, yeast chronological lifespan is extended through induction of autophagy, which is essential for increased longevity [15].

CR was found to substantially decrease the increase in liver, kidney, and brain iron seen in rats fed ad libitum.

Lipid peroxidation was also markedly suppressed in CR animals. Thus, CR has an antioxidant effect which may be largely due to decreased levels of body iron [63].

CR downregulates expression of the iron-regulatory hormone hepcidin in the brain, and this leads to less accumulation of brain iron in aging, which is a key component of neurodegenerative diseases [64]. CR leads to less brain iron deposition in old rhesus monkeys, along with preserved motor performance [65].

Attenuation of increasing iron in liver, kidney, brain, and other tissues may be an important mechanism of the longevity-promoting effects of CR.

Increased dietary iron promotes protein insolubility and aging in *C. elegans*, while pharmacological intervention to block uptake of iron mitigated much damage and extended normal lifespan [66].

### **Heterochronic blood exchange, plasma dilution, and blood donation**

Heterochronic blood exchange between young and old mice results in “rapid inhibition of multiple tissues by old blood”, for reasons that are not clear [67]. Since old animals accumulate iron, and since they exhibit more iron dysregulation resulting in higher levels of free iron, iron may be suspected as the mechanism behind this inhibition of younger tissues by old blood.

In old animals, mere plasma dilution by exchanging it with saline and 5% albumin (an age-neutral blood exchange) leads to rejuvenation of muscle, liver, and brain in old mice [68]. Since there is no young blood involved in this exchange, this makes it doubtful that factors in young blood play an important (if any) role in rejuvenation seen in heterochronic blood exchange.

Human serum from patients who undergo therapeutic plasma exchange (TPE) was also tested for its ability to rejuvenate cells. Old human serum strongly reduced proliferation of mouse myogenic cells, while a single TPE from the same patients reversed this inhibition. Accumulation of iron delays muscle regeneration and suppresses the differentiation of myoblast cells, and this suppression can be reversed with superoxide scavenging [69].

If TPE can lead to serum becoming (or reverting to) a rejuvenating intervention, the question is, what was removed from the serum that allowed for this effect?

Iron makes a good candidate. In TPE, citrate may be used as an anticoagulant; citrate complexes with free iron, and the citrate-iron complex is the major species of

iron found in iron overloaded patients [70]. Patients undergoing TPE have a high rate of iron deficiency anemia; one study found that 60% of patients developed iron deficiency anemia, and 100% of patients had decreased serum iron [71]. This may be due to use of citrate as an anticoagulant, or due to simple removal of plasma, which contains transferrin, and replacement with an albumin solution [72]. Other components of plasma may be removed or diluted as well, but iron may be the critical element here.

Blood donation leads to lower levels of body iron; hemoglobin is the main iron depot in the body, hence its replacement after donation requires the use of body iron stores and decreases them. Several studies have found lower mortality in blood donors, even after accounting for a healthy donor effect from donors being healthier than others even before donation. When only blood donors are studied as a single class, there is an inverse association between blood donation frequency and mortality, with each additional annual donation associated with an 7.5% reduced mortality rate [73]. Blood donation is associated with a marked decrease of body iron in adult men; ferritin values of <15 µg/L (depleted) are about 8 times more common in male donors than in non-donors, and iron deficiency anemia is up to 5 times more common in donors than non-donors [74].

### **Conclusion: Iron squares the circle of life extension**

It can be seen from all of the above that iron is a common theme in many if not most life-extension interventions. This can help make sense of the seemingly disparate mechanisms of extending life by slowing aging.

As noted, autophagy is essential for lifespan extension, and autophagy activation declines with age. Ultimately, this can lead to “the garbage catastrophe of aging”, in which imperfect removal of damaged molecules leads to the accumulation of cellular “garbage”. Much of this decline in autophagy may be due to lipofuscin, a substance that is relatively difficult to remove and which “gums up” the machinery of autophagy. Importantly, iron plays a key role in the formation of lipofuscin; iron can react with polyunsaturated fatty acids and other molecules to form this material, and iron accelerates lipofuscin formation in cultured human glial cells and rat cardiomyocytes [75]. Lower levels of iron could be expected to slow the rate of lipofuscin formation.

Inhibiting the cellular integrator of nutrients and growth, mTOR, leads to longer lifespan in virtually all experimental animals tested so far. We have seen that

mTOR in turn plays a crucial role in the level of body iron stores; mTOR activation increases body iron, and iron in turn activates mTOR. That mTOR inhibition increases lifespan illustrates the fundamental trade-off between growth and longevity, and iron is a growth factor [76].

Many drugs and natural products extend lifespan by seemingly disparate mechanisms, but inhibiting iron absorption, or chelating (binding and removing) iron is a characteristic of many if not most of these substances.

Reduced iron stores can explain how calorie restriction extends lifespan.

Finally, iron can explain the free radical theory of aging. Iron catalyzes the formation of the most damaging free radical, the hydroxyl radical.

In sum, iron satisfies many of the conditions we might look for in a universally pro-aging substance. It accumulates with age; it is associated with many age-related diseases such as cardiovascular disease, cancer, and Alzheimer's disease; it catalyzes the formation of cellular junk molecules and helps to prevent their turnover; removal of iron from plasma may be rejuvenating; and people with lower levels of body iron – blood donors – have a lower mortality rate.

Iron is intimately associated with aging, and control of body iron stores may be an important way to extend human lifespan.

## CONFLICTS OF INTEREST

The author declares that there are no conflicts of interest.

## Editorial note

&This corresponding author has a verified history of publications using a personal email address for correspondence.

## REFERENCES

1. Massie HR, Aiello VR, Williams TR. Iron accumulation during development and ageing of *Drosophila*. *Mech Ageing Dev.* 1985; 29:215–20. [https://doi.org/10.1016/0047-6374\(85\)90020-x](https://doi.org/10.1016/0047-6374(85)90020-x) PMID:3919221
2. James SA, Roberts BR, Hare DJ, de Jonge MD, Birchall IE, Jenkins NL, Cherny RA, Bush AI, McColl G. Direct *in vivo* imaging of ferrous iron dyshomeostasis in ageing *Caenorhabditis elegans*. *Chem Sci.* 2015; 6:2952–62. <https://doi.org/10.1039/c5sc00233h>
3. Massie HR, Aiello VR, Banziger V. Iron accumulation and lipid peroxidation in aging C57BL/6J mice. *Exp Gerontol.* 1983; 18:277–85. [https://doi.org/10.1016/0531-5565\(83\)90038-4](https://doi.org/10.1016/0531-5565(83)90038-4) PMID:6667718
4. Kell DB. Iron behaving badly: inappropriate iron chelation as a major contributor to the aetiology of vascular and other progressive inflammatory and degenerative diseases. *BMC Med Genomics.* 2009; 2:2. <https://doi.org/10.1186/1755-8794-2-2> PMID:19133145
5. Facchini FS. Insulin signaling, glucose metabolism oxidative stress, and aging. *Advances in Cell Aging and Gerontology.* 2003; 14:13–33. [https://doi.org/10.1016/S1566-3124\(03\)14002-3](https://doi.org/10.1016/S1566-3124(03)14002-3)
6. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, Pahor M, Javors MA, Fernandez E, Miller RA. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature.* 2009; 460:392–95. <https://doi.org/10.1038/nature08221> PMID:19587680
7. Koubova J, Guarente L. How does calorie restriction work? *Genes Dev.* 2003; 17:313–21. <https://doi.org/10.1101/gad.1052903> PMID:12569120
8. Shang C, Zhou H, Huang S. Iron chelation inhibits mTOR activity in cancer cells. [abstract]. In: Proceedings of the 105th Annual Meeting of the American Association for Cancer Research; 2014 Apr 5-9; San Diego, CA. Philadelphia (PA): AACR; Cancer Res 2014; 74:2789. <https://doi.org/10.1158/1538-7445.AM2014-2789>
9. Ndong M, Kazami M, Suzuki T, Uehara M, Katsumata S, Inoue H, Kobayashi K, Tadokoro T, Suzuki K, Yamamoto Y. Iron deficiency down-regulates the Akt/TSC1-TSC2/mammalian Target of Rapamycin signaling pathway in rats and in COS-1 cells. *Nutr Res.* 2009; 29:640–47. <https://doi.org/10.1016/j.nutres.2009.09.007> PMID:19854379
10. Mleczko-Sanecka K, Roche F, da Silva AR, Call D, D'Alessio F, Ragab A, Lapinski PE, Ummanni R, Korf U, Oakes C, Damm G, D'Alessandro LA, Klingmüller U, et al. Unbiased RNAi screen for hepcidin regulators links hepcidin suppression to proliferative Ras/RAF and nutrient-dependent mTOR signaling. *Blood.* 2014; 123:1574–85. <https://doi.org/10.1182/blood-2013-07-515957> PMID:24385536
11. Sofroniadou S, Kassimatis T, Goldsmith D. Anaemia, microcytosis and sirolimus--is iron the missing link? PMID:28706676



- Nephrol Dial Transplant. 2010; 25:1667–75.  
<https://doi.org/10.1093/ndt/gfp674>  
PMID:[20054028](https://pubmed.ncbi.nlm.nih.gov/20054028/)
12. Blagosklonny MV. TOR-centric view on insulin resistance and diabetic complications: perspective for endocrinologists and gerontologists. *Cell Death Dis.* 2013; 4:e964.  
<https://doi.org/10.1038/cddis.2013.506>  
PMID:[24336084](https://pubmed.ncbi.nlm.nih.gov/24336084/)
  13. Fernández-Real JM, López-Bermejo A, Ricart W. Cross-talk between iron metabolism and diabetes. *Diabetes.* 2002; 51:2348–54.  
<https://doi.org/10.2337/diabetes.51.8.2348>  
PMID:[12145144](https://pubmed.ncbi.nlm.nih.gov/12145144/)
  14. Sam AH, Busbridge M, Amin A, Webber L, White D, Franks S, Martin NM, Sleeth M, Ismail NA, Daud NM, Papamargaritis D, Le Roux CW, Chapman RS, et al. Hepcidin levels in diabetes mellitus and polycystic ovary syndrome. *Diabet Med.* 2013; 30:1495–99.  
<https://doi.org/10.1111/dme.12262>  
PMID:[23796160](https://pubmed.ncbi.nlm.nih.gov/23796160/)
  15. Montella-Manuel S, Pujol-Carrion N, Mechoud MA, de la Torre-Ruiz MA. Bulk autophagy induction and life extension is achieved when iron is the only limited nutrient in *Saccharomyces cerevisiae*. *Biochem J.* 2021; 478:811–37.  
<https://doi.org/10.1042/BCJ20200849>  
PMID:[33507238](https://pubmed.ncbi.nlm.nih.gov/33507238/)
  16. Madeo F, Zimmermann A, Maiuri MC, Kroemer G. Essential role for autophagy in life span extension. *J Clin Invest.* 2015; 125:85–93.  
<https://doi.org/10.1172/JCI73946>  
PMID:[25654554](https://pubmed.ncbi.nlm.nih.gov/25654554/)
  17. Massie HR, Aiello VR, Williams TR. Inhibition of iron absorption prolongs the life span of *Drosophila*. *Mech Ageing Dev.* 1993; 67:227–37.  
[https://doi.org/10.1016/0047-6374\(93\)90001-8](https://doi.org/10.1016/0047-6374(93)90001-8)  
PMID:[8326745](https://pubmed.ncbi.nlm.nih.gov/8326745/)
  18. Shen LR, Parnell LD, Ordovas JM, Lai CQ. Curcumin and aging. *Biofactors.* 2013; 39:133–40.  
<https://doi.org/10.1002/biof.1086>  
PMID:[23325575](https://pubmed.ncbi.nlm.nih.gov/23325575/)
  19. Jiao Y, Wilkinson J 4th, Christine Pietsch E, Buss JL, Wang W, Planalp R, Torti FM, Torti SV. Iron chelation in the biological activity of curcumin. *Free Radic Biol Med.* 2006; 40:1152–60.  
<https://doi.org/10.1016/j.freeradbiomed.2005.11.003>  
PMID:[16545682](https://pubmed.ncbi.nlm.nih.gov/16545682/)
  20. Chin D, Huebbe P, Frank J, Rimbach G, Pallauf K. Curcumin may impair iron status when fed to mice for six months. *Redox Biol.* 2014; 2:563–69.  
<https://doi.org/10.1016/j.redox.2014.01.018>  
PMID:[24634837](https://pubmed.ncbi.nlm.nih.gov/24634837/)
  21. Abbas S, Wink M. Epigallocatechin gallate from green tea (*Camellia sinensis*) increases lifespan and stress resistance in *Caenorhabditis elegans*. *Planta Med.* 2009; 75:216–21.  
<https://doi.org/10.1055/s-0028-1088378>  
PMID:[19085685](https://pubmed.ncbi.nlm.nih.gov/19085685/)
  22. Wagner AE, Piegholdt S, Rabe D, Baenas N, Schloesser A, Eggersdorfer M, Stocker A, Rimbach G. Epigallocatechin gallate affects glucose metabolism and increases fitness and lifespan in *Drosophila melanogaster*. *Oncotarget.* 2015; 6:30568–78.  
<https://doi.org/10.18632/oncotarget.5215>  
PMID:[26375250](https://pubmed.ncbi.nlm.nih.gov/26375250/)
  23. Sharma R, Kumar R, Sharma A, Goel A, Padwad Y. Long term consumption of green tea EGCG enhances healthspan and lifespan in mice by mitigating multiple aspects of cellular senescence in mitotic and post-mitotic tissues, gut dysbiosis and immunosenescence. *bioRxiv.* 2021. [Epub ahead of print].  
<https://doi.org/10.1101/2021.01.01.425058>
  24. Niu Y, Na L, Feng R, Gong L, Zhao Y, Li Q, Li Y, Sun C. The phytochemical, EGCG, extends lifespan by reducing liver and kidney function damage and improving age-associated inflammation and oxidative stress in healthy rats. *Aging Cell.* 2013; 12:1041–49.  
<https://doi.org/10.1111/acer.12133> PMID:[23834676](https://pubmed.ncbi.nlm.nih.gov/23834676/)
  25. Mandel SA, Amit T, Kalfon L, Reznichenko L, Weinreb O, Youdim MB. Cell signaling pathways and iron chelation in the neurorestorative activity of green tea polyphenols: special reference to epigallocatechin gallate (EGCG). *J Alzheimers Dis.* 2008; 15:211–22.  
<https://doi.org/10.3233/jad-2008-15207>  
PMID:[18953110](https://pubmed.ncbi.nlm.nih.gov/18953110/)
  26. Ren Y, Deng F, Zhu H, Wan W, Ye J, Luo B. Effect of epigallocatechin-3-gallate on iron overload in mice with alcoholic liver disease. *Mol Biol Rep.* 2011; 38:879–86.  
<https://doi.org/10.1007/s11033-010-0180-5>  
PMID:[20490691](https://pubmed.ncbi.nlm.nih.gov/20490691/)
  27. Ayyadevara S, Bharill P, Dandapat A, Hu C, Khaidakov M, Mitra S, Shmookler Reis RJ, Mehta JL. Aspirin inhibits oxidant stress, reduces age-associated functional declines, and extends lifespan of *Caenorhabditis elegans*. *Antioxid Redox Signal.* 2013; 18:481–90.  
<https://doi.org/10.1089/ars.2011.4151>  
PMID:[22866967](https://pubmed.ncbi.nlm.nih.gov/22866967/)
  28. Strong R, Miller RA, Astle CM, Floyd RA, Flurkey K, Hensley KL, Javors MA, Leeuwenburgh C, Nelson JF, Ongini E, Nadon NL, Warner HR, Harrison DE. Nordihydroguaiaretic acid and aspirin increase lifespan

- of genetically heterogeneous male mice. *Aging Cell*. 2008; 7:641–50.  
<https://doi.org/10.1111/j.1474-9726.2008.00414.x>  
PMID:18631321
29. Rothwell PM, Fowkes FG, Belch JF, Ogawa H, Warlow CP, Meade TW. Effect of daily aspirin on long-term risk of death due to cancer: analysis of individual patient data from randomised trials. *Lancet*. 2011; 377:31–41.  
[https://doi.org/10.1016/S0140-6736\(10\)62110-1](https://doi.org/10.1016/S0140-6736(10)62110-1)  
PMID:21144578
30. Mascitelli L, Pezzetta F, Sullivan JL. Aspirin-associated iron loss as an anticancer mechanism. *Med Hypotheses*. 2010; 74:78–80.  
<https://doi.org/10.1016/j.mehy.2009.07.047>  
PMID:19692186
31. Pietrocchi F, Castoldi F, Markaki M, Lachkar S, Chen G, Enot DP, Durand S, Bossut N, Tong M, Malik SA, Loos F, Dupont N, Mariño G, et al. Aspirin Recapitulates Features of Caloric Restriction. *Cell Rep*. 2018; 22:2395–407.  
<https://doi.org/10.1016/j.celrep.2018.02.024>  
PMID:29490275
32. Reid KR, Meyerhoff ME, Mitchell-Koch JT. Salicylate detection by complexation with iron (III) and optical absorbance spectroscopy. an undergraduate quantitative analysis experiment. *J Chem Educ*. 2008; 85:1658.  
<https://doi.org/10.1021/ed085p1658>
33. Visca P, Ciervo A, Sanfilippo V, Orsi N. Iron-regulated salicylate synthesis by *Pseudomonas* spp. *J Gen Microbiol*. 1993; 139:1995–2001.  
<https://doi.org/10.1099/00221287-139-9-1995>  
PMID:7504066
34. Oberle S, Polte T, Abate A, Podhasky HP, Schröder H. Aspirin increases ferritin synthesis in endothelial cells: a novel antioxidant pathway. *Circ Res*. 1998; 82:1016–20.  
<https://doi.org/10.1161/01.res.82.9.1016>  
PMID:9598599
35. Lublin A, Isoda F, Patel H, Yen K, Nguyen L, Hajje D, Schwartz M, Mobbs C. FDA-approved drugs that protect mammalian neurons from glucose toxicity slow aging dependent on cbp and protect against proteotoxicity. *PLoS One*. 2011; 6:e27762.  
<https://doi.org/10.1371/journal.pone.0027762>  
PMID:22114686
36. Kolaylı S, Ocak M, Küçük M, Abbasoğlu R. Does caffeine bind to metal ions? *Food Chemistry*. 2004; 84:383–88.  
[https://doi.org/10.1016/S0308-8146\(03\)00244-9](https://doi.org/10.1016/S0308-8146(03)00244-9)
37. Phiwchai I, Yuensook W, Sawaengsiriphon N, Krunghanuch S, Pilapong C. Tannic acid (TA): A molecular tool for chelating and imaging labile iron. *Eur J Pharm Sci*. 2018; 114:64–73.  
<https://doi.org/10.1016/j.ejps.2017.12.004>  
PMID:29225106
38. Eberhard Y, McDermott SP, Wang X, Gronda M, Venugopal A, Wood TE, Hurren R, Datti A, Batey RA, Wrana J, Antholine WE, Dick JE, Schimmer AD. Chelation of intracellular iron with the antifungal agent ciclopirox olamine induces cell death in leukemia and myeloma cells. *Blood*. 2009; 114:3064–73.  
<https://doi.org/10.1182/blood-2009-03-209965>  
PMID:19589922
39. Quinlan GJ, Gutteridge JM. Bacitracin and a bacitracin-zinc complex damage DNA and carbohydrate in the presence of iron and copper salts. *Free Radic Res Commun*. 1989; 7:37–44.  
<https://doi.org/10.3109/10715768909088160>  
PMID:2509300
40. Perez CA, Wei Y, Guo M. Iron-binding and anti-Fenton properties of baicalein and baicalin. *J Inorg Biochem*. 2009; 103:326–32.  
<https://doi.org/10.1016/j.jinorgbio.2008.11.003>  
PMID:19108897
41. Walker EM Jr, Epling CP, Parris C, Cansino S, Ghosh P, Desai DH, Morrison RG, Wright GL, Wehner P, Mangiarua EI, Walker SM, Blough ER. Acetaminophen protects against iron-induced cardiac damage in gerbils. *Ann Clin Lab Sci*. 2007; 37:22–33.  
PMID:17311866
42. Brandstädt S, Schmeisser K, Zarse K, Ristow M. Lipid-lowering fibrates extend *C. elegans* lifespan in a NHR-49/PPARalpha-dependent manner. *Aging (Albany NY)*. 2013; 5:270–75.  
<https://doi.org/10.18632/aging.100548>  
PMID:23603800
43. Huang HL, Shaw NS. Role of hypolipidemic drug clofibrate in altering iron regulatory proteins IRP1 and IRP2 activities and hepatic iron metabolism in rats fed a low-iron diet. *Toxicol Appl Pharmacol*. 2002; 180:118–28.  
<https://doi.org/10.1006/taap.2002.9378>  
PMID:11969379
44. Dang Y, An Y, He J, Huang B, Zhu J, Gao M, Zhang S, Wang X, Yang B, Xie Z. Berberine ameliorates cellular senescence and extends the lifespan of mice via regulating p16 and cyclin protein expression. *Aging Cell*. 2020; 19:e13060.  
<https://doi.org/10.1111/acer.13060> PMID:31773901
45. Zhao H, Halicka HD, Li J, Darzynkiewicz Z. Berberine suppresses gero-conversion from cell cycle arrest to senescence. *Aging (Albany NY)*. 2013; 5:623–36.  
<https://doi.org/10.18632/aging.100593>

- PMID:[23974852](#)
46. Shirwaikar A, Shirwaikar A, Rajendran K, Punitha IS. In vitro antioxidant studies on the benzyl tetra isoquinoline alkaloid berberine. *Biol Pharm Bull.* 2006; 29:1906–10.  
<https://doi.org/10.1248/bpb.29.1906> PMID:[16946507](#)
  47. Harrison DE, Strong R, Allison DB, Ames BN, Astle CM, Atamna H, Fernandez E, Flurkey K, Javors MA, Nadon NL, Nelson JF, Pletcher S, Simpkins JW, et al. Acarbose, 17- $\alpha$ -estradiol, and nordihydroguaiaretic acid extend mouse lifespan preferentially in males. *Aging Cell.* 2014; 13:273–82.  
<https://doi.org/10.1111/acer.12170> PMID:[24245565](#)
  48. Yoo WH, Park TS, Baek HS. Marked weight loss in a type 2 diabetic patient treated with acarbose. *Diabetes Care.* 1999; 22:645–46.  
<https://doi.org/10.2337/diacare.22.4.645> PMID:[10189546](#)
  49. Couet C, Ulmer M, Hamdaoui M, Bau HM, Debry G. Metabolic effects of acarbose in young healthy men. *Eur J Clin Nutr.* 1989; 43:187–96.  
PMID:[2659314](#)
  50. Houtkooper RH, Mouchiroud L, Ryu D, Moullan N, Katsyuba E, Knott G, Williams RW, Auwerx J. Mitonuclear protein imbalance as a conserved longevity mechanism. *Nature.* 2013; 497:451–57.  
<https://doi.org/10.1038/nature12188> PMID:[23698443](#)
  51. Grenier D, Huot MP, Mayrand D. Iron-chelating activity of tetracyclines and its impact on the susceptibility of *Actinobacillus actinomycetemcomitans* to these antibiotics. *Antimicrob Agents Chemother.* 2000; 44:763–66.  
<https://doi.org/10.1128/AAC.44.3.763-766.2000> PMID:[10681353](#)
  52. Santos EL, de Picoli Souza K, da Silva ED, Batista EC, Martins PJ, D’Almeida V, Pesquero JB. Long term treatment with ACE inhibitor enalapril decreases body weight gain and increases life span in rats. *Biochem Pharmacol.* 2009; 78:951–58.  
<https://doi.org/10.1016/j.bcp.2009.06.018> PMID:[19549507](#)
  53. Lee SC, Park SW, Kim DK, Lee SH, Hong KP. Iron supplementation inhibits cough associated with ACE inhibitors. *Hypertension.* 2001; 38:166–70.  
<https://doi.org/10.1161/01.hyp.38.2.166> PMID:[11509470](#)
  54. He C, Tsuchiyama SK, Nguyen QT, Plyusnina EN, Terrill SR, Sahibzada S, Patel B, Faulkner AR, Shaposhnikov MV, Tian R, Tsuchiya M, Kaeberlein M, Moskalev AA, et al. Enhanced longevity by ibuprofen, conserved in multiple species, occurs in yeast through inhibition of tryptophan import. *PLoS Genet.* 2014; 10:e1004860.  
<https://doi.org/10.1371/journal.pgen.1004860> PMID:[25521617](#)
  55. Kennedy TP, Rao NV, Noah W, Michael JR, Jafri MH Jr, Gurtner GH, Hoidal JR. Ibuprofen prevents oxidant lung injury and *in vitro* lipid peroxidation by chelating iron. *J Clin Invest.* 1990; 86:1565–73.  
<https://doi.org/10.1172/JCI114876> PMID:[2173723](#)
  56. Martin-Montalvo A, Mercken EM, Mitchell SJ, Palacios HH, Mote PL, Scheibye-Knudsen M, Gomes AP, Ward TM, Minor RK, Blouin MJ, Schwab M, Pollak M, Zhang Y, et al. Metformin improves healthspan and lifespan in mice. *Nat Commun.* 2013; 4:2192.  
<https://doi.org/10.1038/ncomms3192> PMID:[23900241](#)
  57. Li X, Wang X, Snyder MP. Metformin Affects Heme Function as a Possible Mechanism of Action. *G3 (Bethesda).* 2019; 9:513–22.  
<https://doi.org/10.1534/g3.118.200803> PMID:[30554148](#)
  58. Stynen B, Abd-Rabbo D, Kowarzyk J, Miller-Fleming L, Aulakh SK, Garneau P, Ralser M, Michnick SW. Changes of Cell Biochemical States Are Revealed in Protein Homomeric Complex Dynamics. *Cell.* 2018; 175:1418–29.e9.  
<https://doi.org/10.1016/j.cell.2018.09.050> PMID:[30454649](#)
  59. Kampkötter A, Timpel C, Zurawski RF, Ruhl S, Chovolou Y, Proksch P, Wätjen W. Increase of stress resistance and lifespan of *Caenorhabditis elegans* by quercetin. *Comp Biochem Physiol B Biochem Mol Biol.* 2008; 149:314–23.  
<https://doi.org/10.1016/j.cbpb.2007.10.004> PMID:[18024103](#)
  60. Leopoldini M, Russo N, Chiodo S, Toscano M. Iron chelation by the powerful antioxidant flavonoid quercetin. *J Agric Food Chem.* 2006; 54:6343–51.  
<https://doi.org/10.1021/jf060986h> PMID:[16910729](#)
  61. Al-Regaiey KA. The effects of calorie restriction on aging: a brief review. *Eur Rev Med Pharmacol Sci.* 2016; 20:2468–73.  
PMID:[27338076](#)
  62. Reverter-Branchat G, Cabiscol E, Tamarit J, Ros J. Oxidative damage to specific proteins in replicative and chronological-aged *Saccharomyces cerevisiae*: common targets and prevention by calorie restriction. *J Biol Chem.* 2004; 279:31983–89.  
<https://doi.org/10.1074/jbc.M404849200> PMID:[15166233](#)
  63. Cook CI, Yu BP. Iron accumulation in aging:



- modulation by dietary restriction. *Mech Ageing Dev.* 1998; 102:1–13.  
[https://doi.org/10.1016/s0047-6374\(98\)00005-0](https://doi.org/10.1016/s0047-6374(98)00005-0)  
PMID:[9663787](https://pubmed.ncbi.nlm.nih.gov/9663787/)
64. Wei S, Shi W, Li M, Gao Q. Calorie restriction down-regulates expression of the iron regulatory hormone hepcidin in normal and D-galactose-induced aging mouse brain. *Rejuvenation Res.* 2014; 17:19–26.  
<https://doi.org/10.1089/rej.2013.1450>  
PMID:[24044515](https://pubmed.ncbi.nlm.nih.gov/24044515/)
65. Kastman EK, Willette AA, Coe CL, Bendlin BB, Kosmatka KJ, McLaren DG, Xu G, Canu E, Field AS, Alexander AL, Voytko ML, Beasley TM, Colman RJ, et al. A calorie-restricted diet decreases brain iron accumulation and preserves motor performance in old rhesus monkeys. *J Neurosci.* 2010; 30:7940–47.  
<https://doi.org/10.1523/JNEUROSCI.0835-10.2010>  
PMID:[20534842](https://pubmed.ncbi.nlm.nih.gov/20534842/)
66. Klang IM, Schilling B, Sorensen DJ, Sahu AK, Kapahi P, Andersen JK, Swoboda P, Killilea DW, Gibson BW, Lithgow GJ. Iron promotes protein insolubility and aging in *C. elegans*. *Aging (Albany NY).* 2014; 6:975–91.  
<https://doi.org/10.18632/aging.100689>  
PMID:[25554795](https://pubmed.ncbi.nlm.nih.gov/25554795/)
67. Rebo J, Mehdipour M, Gathwala R, Causey K, Liu Y, Conboy MJ, Conboy IM. A single heterochronic blood exchange reveals rapid inhibition of multiple tissues by old blood. *Nat Commun.* 2016; 7:13363.  
<https://doi.org/10.1038/ncomms13363>  
PMID:[27874859](https://pubmed.ncbi.nlm.nih.gov/27874859/)
68. Mehdipour M, Skinner C, Wong N, Lieb M, Liu C, Etienne J, Kato C, Kiproff D, Conboy MJ, Conboy IM. Rejuvenation of three germ layers tissues by exchanging old blood plasma with saline-albumin. *Aging (Albany NY).* 2020; 12:8790–819.  
<https://doi.org/10.18632/aging.103418>  
PMID:[32474458](https://pubmed.ncbi.nlm.nih.gov/32474458/)
69. Ikeda Y, Satoh A, Horinouchi Y, Hamano H, Watanabe H, Imao M, Imanishi M, Zamami Y, Takechi K, Izawa-Ishizawa Y, Miyamoto L, Hirayama T, Nagasawa H, et al. Iron accumulation causes impaired myogenesis correlated with MAPK signaling pathway inhibition by oxidative stress. *FASEB J.* 2019; 33:9551–64.  
<https://doi.org/10.1096/fj.201802724RR>  
PMID:[31145863](https://pubmed.ncbi.nlm.nih.gov/31145863/)
70. Grootveld M, Bell JD, Halliwell B, Aruoma OI, Bomford A, Sadler PJ. Non-transferrin-bound iron in plasma or serum from patients with idiopathic hemochromatosis. Characterization by high performance liquid chromatography and nuclear magnetic resonance spectroscopy. *J Biol Chem.* 1989; 264:4417–22.  
PMID:[2466835](https://pubmed.ncbi.nlm.nih.gov/2466835/)
71. Compton F, Salazar E, Klein K, Tint H, Castillo B, Bai Y. New Onset Iron Deficiency Anemia in Chronic Therapeutic Plasma Exchange Patients. *Ann Clin Lab Sci.* 2018; 48:273–78.  
PMID:[29970428](https://pubmed.ncbi.nlm.nih.gov/29970428/)
72. Ahmadpoor P, Aglae C, Cariou S, Pambrun E, Renaud S, Garo F, Darmon R, Schultz C, Prelipcean C, Reboul P, Moranne O. Physiological role of plasma and its components and the clinical implications of different methods of apheresis: A narrative review. *Ther Apher Dial.* 2021; 25:262–72.  
<https://doi.org/10.1111/1744-9987.13567>  
PMID:[32710797](https://pubmed.ncbi.nlm.nih.gov/32710797/)
73. Ullum H, Rostgaard K, Kamper-Jørgensen M, Reilly M, Melbye M, Nyrén O, Norda R, Edgren G, Hjalgrim H. Blood donation and blood donor mortality after adjustment for a healthy donor effect. *Transfusion.* 2015; 55:2479–85.  
<https://doi.org/10.1111/trf.13205> PMID:[26098293](https://pubmed.ncbi.nlm.nih.gov/26098293/)
74. Milman N, Kirchhoff M. Influence of blood donation on iron stores assessed by serum ferritin and haemoglobin in a population survey of 1433 Danish males. *Eur J Haematol.* 1991; 47:134–39.  
<https://doi.org/10.1111/j.1600-0609.1991.tb00136.x>  
PMID:[1889481](https://pubmed.ncbi.nlm.nih.gov/1889481/)
75. Terman A. Garbage catastrophe theory of aging: imperfect removal of oxidative damage? *Redox Rep.* 2001; 6:15–26.  
<https://doi.org/10.1179/135100001101535996>  
PMID:[11333111](https://pubmed.ncbi.nlm.nih.gov/11333111/)
76. Blagosklonny MV, Hall MN. Growth and aging: a common molecular mechanism. *Aging (Albany NY).* 2009; 1:357–62.  
<https://doi.org/10.18632/aging.100040>  
PMID:[20157523](https://pubmed.ncbi.nlm.nih.gov/20157523/)